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(57) Abstract

The invention provides isolated nucleic acids molecules, designated CSAPK nucleic acid molecules, which encode novel protein kinases. The invention also provides antisense nucleic acid molecules, recombinant expression vectors containing CSAPK nucleic acid molecules, host cells into which the expression vectors have been introduced, and nonhuman transgenic animals in which a CSAPK gene has been introduced or disrupted. The invention still further provides isolated CSAPK proteins, fusion proteins, antigenic peptides and anti-CSAPK antibodies. Diagnostic, screening, and therapeutic methods utilizing compositions of the invention are also provided.

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NOVEL PROTEIN KINASE MOLECULES AND USES THEREFOR

Background of the Invention

Phosphate tightly associated with protein has been known since the late nineteenth century. Since then, a variety of covalent linkages of phosphate to proteins have been found. The most common involve esterification of phosphate to serine, threonine, and tyrosine with smaller amounts being linked to lysine, arginine, histidine, aspartic acid, glutamic acid, and cysteine. The occurrence of phosphorylated proteins implies the existence of one or more protein kinases capable of phosphorylating amino acid residues on proteins, and also of protein phosphatases capable of hydrolyzing phosphorylated amino acid residues on proteins.

Protein kinases play critical roles in the regulation of biochemical and morphological changes associated with cellular growth and division (D'Urso, G. et al. (1990) Science 250: 786-791; Birchmeier. C. et al. (1993) Bioessays 15: 185-189). They serve as growth factor receptors and signal transducers and have been implicated in cellular transformation and malignancy (Hunter, T. et al. (1992) Cell 70: 375-387; Posada, J. et al. (1992) Mol. Biol. Cell 3: 583-592, Hunter, T. et al. (1994) Cell 79: 573-582). For example, protein kinases have been shown to participate in the transmission of signals from growth-factor receptors (Sturgill, T. W. et al. (1988) Nature 344: 715-718; Gomez, N. et al. (1991) Nature 353: 170-173), control of entry of cells into mitosis (Nurse, P. (1990) Nature 344: 503-508; Maller, J. L. (1991) Curr. Opin. Cell Biol. 3: 269-275) and regulation of actin bundling (Husain-Chishti, A. et al. (1988) Nature 334: 718-721). Protein kinases can be divided into two main groups based on either amino acid sequence similarity or specificity for either serine/threonine or tyrosine residues. A small number of dual-specificity kinases are structurally like the serine/threoninespecific group. Within the broad classification, kinases can be further sub-divided into families whose members share a higher degree of catalytic domain amino acid sequence identity and also have similar biochemical properties. Most protein kinase family members also share structural features outside the kinase domain that reflect their particular cellular roles. These include regulatory domains that control kinase activity or interaction with other proteins (Hanks, S.K. et al. (1988) Science 241: 42-52).

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Summary of the Invention

The present invention is based, at least in part, on the discovery of novel nucleic acid molecules and proteins encoded by such nucleic acid molecules, referred to herein as "Cardiovascular System Associated Protein Kinase" ("CSAPK") proteins. The CSAPK nucleic acid and protein molecules of the present invention are useful as modulating agents in regulating a variety of cellular processes, e.g., cardiac cellular processes. Accordingly, in one aspect, this invention provides isolated nucleic acid molecules encoding CSAPK proteins or biologically active portions thereof, as well as nucleic acid fragments suitable as primers or hybridization probes for the detection of CSAPK-encoding nucleic acids.

In one embodiment, a CSAPK nucleic acid molecule of the invention is at least 60%, 62%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to a nucleotide sequence (e.g., to the entire length of the nucleotide sequence) including SEQ ID NO:1, SEO ID NO:3, or a complement thereof. In another embodiment, a CSAPK nucleic acid molecule is 50%, 54%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 86%, 90%, 95%, 98% homologous to a nucleotide sequence including SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a complement thereof. In yet another embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 97%, 98% homologous to a nucleotide sequence including SEQ ID NO:7, SEQ ID NO:9, or a complement thereof. In yet another embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 73%, 75%, 80%, 85%, 86%, 90%, 95%, 98% homologous to a nucleotide sequence including SEQ ID NO:10, SEQ ID NO:12, or a complement thereof. In a further embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 75%, 78%, 80%, 85%, 90%, 95%, 98% homologous to a nucleotide sequence including SEQ ID NO:13, SEQ ID NO:15, or a complement thereof.

In a preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:1 or 3, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:3 and nucleotides 1-296 of SEQ ID NO:1. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:3 and nucleotides 1202-4137 of SEQ ID NO:1. In another preferred embodiment,

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the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:1 or 3. In another preferred embodiment, the nucleic acid molecule comprises a fragment of at least 509 nucleotides of the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, or a complement thereof.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:4, 6, 16, 17, 18, 19, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:6 and nucleotides 1-46 of SEQ ID NO:4. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:6 and nucleotides 1411-2120 of SEQ ID NO:4. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:4, 6, 16, 17, 18, 19.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:7 or 9, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:9 and nucleotides 1-50 of SEQ ID NO:7. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:9 and nucleotides 1793-2454 of SEQ ID NO:7. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:7 or 9.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:10 or 12, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:12 and nucleotides 1-274 of SEQ ID NO:10. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:12 and nucleotides 755-1864 of SEQ ID NO:10. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:10 or 12.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown in SEQ ID NO:13 or 15, or a complement thereof. In yet another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:13 or 15

In another embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or

SEQ ID NO:14. In a preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:2 (e.g., the entire amino acid sequence of SEQ ID NO:2). In another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 42%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:5 (e.g., the entire amino acid sequence of SEQ ID NO:5). In yet another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 41%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:8 (e.g., the entire amino acid sequence of SEQ ID NO:8). In a further preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 59%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:11 (e.g., the entire amino acid sequence of SEQ·ID NO:11). In another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:14).

In another preferred embodiment, an isolated nucleic acid molecule encodes the amino acid sequence of a human CSAPK. In yet another preferred embodiment, the nucleic acid molecule includes a nucleotide sequence encoding a protein which includes the amino acid sequence of SEQ ID NO: 2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14. In yet another preferred embodiment, the nucleic acid molecule includes a nucleotide sequence encoding a protein having the amino acid sequence of SEQ ID NO: 2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14.

Another embodiment of the invention features nucleic acid molecules, preferably CSAPK nucleic acid molecules, which specifically detect CSAPK nucleic acid molecules relative to nucleic acid molecules encoding non-CSAPK proteins. For

example, in one embodiment, such a nucleic acid molecule is at least 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, or 800 nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, 19, or a complement thereof.

In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:2, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:1 or SEQ ID NO:3 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:5, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:8, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:7 or SEQ ID NO:9 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:10 or SEQ ID NO:12 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:14, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:13 or SEQ ID NO:15 under stringent conditions.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to a CSAPK nucleic acid molecule, e.g., the coding strand of a CSAPK nucleic acid molecule.

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Another aspect of the invention provides a vector comprising a CSAPK nucleic acid molecule. In certain embodiments, the vector is a recombinant expression vector. In another embodiment, the invention provides a host cell containing a vector of the invention. The invention also provides a method for producing a protein, preferably a CSAPK protein, by culturing in a suitable medium, a host cell, e.g., a mammalian host cell such as a non-human mammalian cell, of the invention containing a recombinant expression vector, such that the protein is produced.

Another aspect of this invention features isolated or recombinant CSAPK proteins and polypeptides. In one embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATPbinding site. In another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 51%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 99% or more homologous to an amino acid sequence including SEQ ID NO:2. In yet another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATPbinding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, at least one leucine zipper-basic region, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity

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kinase catalytic domain and at least one ATP-binding site and has an amino acid sequence which is at least 42%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:5. In yet another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, and at least one ATPbinding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, and at least one ATPbinding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity. kinase catalytic domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

In yet another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 41%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:8. In yet another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which

hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:7 or SEQ ID NO:9.

In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 59%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:11. In yet another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:10 or SEQ ID NO:12.

In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In yet another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:14. In yet another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated

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with cell cycle regulation. In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is able to interact with Nck (described in Lehman et al. (1990) Nucleic Acids Res. 18:1048). In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:13 or SEQ ID NO:15.

In another embodiment, the isolated protein, preferably a CSAPK protein, has an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14. In a preferred embodiment, the protein, preferably a CSAPK protein, has an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEO ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11. or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14). In another embodiment, the invention features fragments of the proteins having the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14, wherein the fragment comprises at least 15 amino acids (e.g., contiguous amino acids) of the amino acid sequence of SEQ ID NO:2, SEQ 20 ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14, respectively. In another embodiment, the protein, preferably a CSAPK protein, has the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14.

Another embodiment of the invention features an isolated protein, preferably a CSAPK protein, which is encoded by a nucleic acid molecule having a nucleotide sequence at least about 50%, 54%, 55%, 60%, 62%, 65%, 70%, 75%, 78%, 80%, 85%, 86%, 90%, 95%, 97%, 98% or more homologous to a nucleotide sequence (e.g., to the entire length of the nucleotide sequence) including SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, or a complement thereof. This invention further features an isolated protein,

preferably a CSAPK protein, which is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a complement thereof.

The proteins of the present invention or biologically active portions thereof, can be operatively linked to a non-CSAPK polypeptide (e.g., heterologous amino acid sequences) to form fusion proteins. The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind proteins of the invention, preferably CSAPK proteins. In addition, the CSAPK proteins or biologically active portions thereof can be incorporated into pharmaceutical compositions, which optionally include pharmaceutically acceptable carriers.

In another aspect, the present invention provides a method for detecting the presence of a CSAPK nucleic acid molecule, protein or polypeptide in a biological sample by contacting the biological sample with an agent capable of detecting a CSAPK nucleic acid molecule, protein or polypeptide such that the presence of a CSAPK nucleic acid molecule, protein or polypeptide is detected in the biological sample.

In another aspect, the present invention provides a method for detecting the presence of CSAPK activity in a biological sample by contacting the biological sample with an agent capable of detecting an indicator of CSAPK activity such that the presence of CSAPK activity is detected in the biological sample.

In another aspect, the invention provides a method for modulating CSAPK activity comprising contacting a cell capable of expressing CSAPK with an agent that modulates CSAPK activity such that CSAPK activity in the cell is modulated. In one embodiment, the agent inhibits CSAPK activity. In another embodiment, the agent stimulates CSAPK activity. In one embodiment, the agent is an antibody that specifically binds to a CSAPK protein. In another embodiment, the agent modulates expression of CSAPK by modulating transcription of a CSAPK gene or translation of a CSAPK mRNA. In yet another embodiment, the agent is a nucleic acid molecule having

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a nucleotide sequence that is antisense to the coding strand of a CSAPK mRNA or a CSAPK gene.

In one embodiment, the methods of the present invention are used to treat a subject having a disorder characterized by aberrant CSAPK protein or nucleic acid expression or activity by administering an agent which is a CSAPK modulator to the subject. In one embodiment, the CSAPK modulator is a CSAPK protein. In another embodiment the CSAPK modulator is a CSAPK nucleic acid molecule. In yet another embodiment, the CSAPK modulator is a peptide, peptidomimetic, or other small molecule. In a preferred embodiment, the disorder characterized by aberrant CSAPK protein or nucleic acid expression is a cellular growth related disorder, e.g., a cardiovascular disorder.

The present invention also provides a diagnostic assay for identifying the presence or absence of a genetic alteration characterized by at least one of (i) aberrant modification or mutation of a gene encoding a CSAPK protein; (ii) mis-regulation of the gene; and (iii) aberrant post-translational modification of a CSAPK protein, wherein a wild-type form of the gene encodes a protein with a CSAPK activity.

In another aspect the invention provides a method for identifying a compound that binds to or modulates the activity of a CSAPK protein, by providing an indicator composition comprising a CSAPK protein having CSAPK activity, contacting the indicator composition with a test compound, and determining the effect of the test compound on CSAPK activity in the indicator composition to identify a compound that modulates the activity of a CSAPK protein.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

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Brief Description of the Drawings

Figure 1 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-1. The nucleotide sequence corresponds to nucleic acids 1 to 4137 of SEQ ID NO:1. The amino acid sequence corresponds to amino acids 1 to 302 of SEQ ID NO:2. The coding region without the 5' and 3' untranslated regions of the human CSAPK-1 gene is shown in SEQ ID NO:3.

Figure 2 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-2. The nucleotide sequence corresponds to nucleic acids 1 to 2120 of SEQ ID NO:4. The amino acid sequence corresponds to amino acids 1 to 455 of SEQ ID NO:5. The coding region without the 5' and 3' untranslated regions of the human CSAPK-2 gene is shown in SEQ ID NO:6.

Figure 3 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-3. The nucleotide sequence corresponds to nucleic acids 1 to 2454 of SEQ ID NO:7. The amino acid sequence corresponds to amino acids 1 to 581 of SEQ ID NO:8. The coding region without the 5' and 3' untranslated regions of the human CSAPK-3 gene is shown in SEQ ID NO:9.

Figure 4 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-4. The nucleotide sequence corresponds to nucleic acids 1 to 1864 of SEQ ID NO:10. The amino acid sequence corresponds to amino acids 1 to 160 of SEQ ID NO:11. The coding region without the 5' and 3' untranslated regions of the human CSAPK-4 gene is shown in SEQ ID NO:12.

Figure 5 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-5. The nucleotide sequence corresponds to nucleic acids 1 to 1333 of SEQ ID NO:13. The amino acid sequence corresponds to amino acids 1 to 444 of SEQ ID NO:14. The coding region without the 5' and 3' untranslated regions of the human CSAPK-5 gene is shown in SEQ ID NO:15.

Figure 6 depicts a schematic diagram of the MLK (CSAPK-2) overexpression construct used to create the MLK (CSAPK-2) transgenic animals.

Figure 7 is a diagramatic depiction of the MLK (CSAPK-2) cDNA splice variants.

Figure 8 is a diagramatic depiction of the resulting proteins from the MLK (CSAPK-2) splice variants.

Figure 9 depicts the cDNA sequence of the human splice variant CSAPK-2a. The nucleotide sequence corresponds to nucleic acids 1 to 2194 of SEQ ID NO:16.

Figure 10 depicts the cDNA sequence of the human splice variant CSAPK-2b.

The nucleotide sequence corresponds to nucleic acids 1 to 2254 of SEQ ID NO:17.

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Figure 11 depicts the cDNA sequence of the human splice variant CSAPK-2c. The nucleotide sequence corresponds to nucleic acids 1 to 2272 of SEQ ID NO:18.

Figure 12 depicts the cDNA sequence of the human splice variant CSAPK-2d. The nucleotide sequence corresponds to nucleic acids 1 to 2069 of SEQ ID NO:19.

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Detailed Description of the Invention

The present invention is based, at least in part, on the discovery of novel molecules, referred to herein as "Cardiovascular System Associated Protein Kinase" or "CSAPK" nucleic acid and polypeptide molecules, which play a role in or function in signalling pathways associated with cellular growth. In one embodiment, the CSAPK molecules modulate the activity of one or more proteins involved in cellular growth or differentiation, e.g., cardiac cell growth or differentiation. In another embodiment, the CSAPK molecules of the present invention are capable of modulating the phosphorylation state of a CSAPK molecule or one or more proteins involved in cellular growth or differentiation, e.g., cardiac cell growth or differentiation.

growth or differentiation, e.g., cardiac cell growth or differentiation.

In a preferred embodiment, the CSAPK molecules are protein kinases which are

expressed and/or function in cells of the cardiovascular system, e.g., cells of the heart,

the blood vessels, and/or the blood.

As used herein, the term "protein kinase" includes a protein or polypeptide which is capable of modulating its own phosphorylation state or the phosphorylation state of another protein or polypeptide. Protein kinases can have a specificity for (i.e., a specificity to phosphorylate) serine/threonine residues, tyrosine residues, or both serine/threonine and tyrosine residues, e.g., the dual specificity kinases. As referred to herein, protein kinases preferably include a catalytic domain of about 200-400 amino acid residues in length, preferably about 200-300 amino acid residues in length, or more preferably about 250-300 amino acid residues in length, which includes preferably 5-20, more preferably 5-15, or preferably 11 highly conserved motifs or subdomains separated by sequences of amino acids with reduced or minimal conservation.

Specificity of a protein kinase for phosphorylation of either tyrosine or serine/threonine can be predicted by the sequence of two of the subdomains (VIb and VIII) in which different residues are conserved in each class (as described in, for example, Hanks et al.

(1988) Science 241:42-52) the contents of which are incorporated herein by reference). These subdomains are also described in further detail herein.

Protein kinases play a role in signalling pathways associated with cellular growth. For example, protein kinases are involved in the regulation of signal transmission from cellular receptors, e.g., growth-factor receptors; entry of cells into mitosis; and the regulation of cytoskeleton function, e.g., actin bundling. Thus, the CSAPK molecules of the present invention may be involved in: 1) the regulation of transmission of signals from cellular receptors, e.g., cardiac cell growth factor receptors; 2) the modulation of the entry of cells, e.g., cardiac precursor cells, into mitosis; 3) the modulation of cellular differentiation; 4) the modulation of cell death; and 5) the regulation of cytoskeleton function, e.g., actin bundling.

Inhibition or over stimulation of the activity of protein kinases involved in signaling pathways associated with cellular growth can lead to perturbed cellular growth, which can in turn lead to cellular growth related disorders. As used herein, a "cellular growth related disorder" includes a disorder, disease, or condition characterized by a deregulation, e.g., an upregulation or a downregulation, of cellular growth. Cellular growth deregulation may be due to a deregulation of cellular proliferation, cell cycle progression, cellular differentiation and/or cellular hypertrophy. Examples of cellular growth related disorders include cardiovascular disorders such as heart failure, hypertension, atrial fibrillation, dilated cardiomyopathy, idiopathic cardiomyopathy, or angina; proliferative disorders or differentiative disorders such as cancer, e.g., melanoma, prostate cancer, cervical cancer, breast cancer, colon cancer, or sarcoma.

The present invention is based, at least in part, on the discovery of novel molecules, referred to herein as CSAPK protein and nucleic acid molecules, which comprise a family of molecules having certain conserved structural and functional features. The term "family" when referring to the protein and nucleic acid molecules of the invention is intended to mean two or more proteins or nucleic acid molecules having a common structural domain or motif and having sufficient amino acid or nucleotide sequence homology as defined herein. Such family members can be naturally or non-naturally occurring and can be from either the same or different species. For example, a family can contain a first protein of human origin, as well as other, distinct proteins of

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human origin or alternatively, can contain homologues of non-human origin. Members of a family may also have common functional characteristics.

One embodiment of the invention features CSAPK nucleic acid molecules, preferably human CSAPK molecules, e.g., CSAPK-1, CSAPK-2, CSAPK-2a, CSAPK-2b, CSAPK-2c, CSAPK-2d, CSAPK-3, CSAPK-4, and CSAPK-5, which were identified from cDNA libraries made from hearts of patients with congestive heart failure (CHF) of ischemic and idiopathic origin. The CSAPK nucleic acid and protein molecules of the invention are described in further detail in the following subsections.

10 A. The CSAPK-1 Nucleic Acid and Protein Molecules

In one embodiment, the isolated proteins of the present invention, preferably CSAPK-1 proteins, are identified based on the presence of at least one "Ser/Thr kinase site" and at least one "ATP-binding region." As used herein, the term "Ser/Thr kinase site" includes an amino acid sequence of about 200-400 amino acid residues in length, preferably 200-300 amino acid residues in length, and more preferably 250-300 amino acid residues in length, which is conserved in kinases which phosphorylate serine and threonine residues and found in the catalytic domain of Ser/Thr kinases. Preferably, the Ser/Thr kinase site includes the following amino acid consensus sequence X9-g-X-G-X₄-V-X₁₂-K-X-(10-19)-E-X₆₆-h-X₈-h-r-D-X-K-X₂-N-X₁₇-K-X₂-D-f-g-X₂₁-p-X₁₃w-X3-g-X55-R-X14-h-X3 (SEQ ID NO:22) (where invariant residues are indicated by upper case letters and nearly invariant residues are indicated by lower case letters). The nearly invariant residues are usually found in most Ser/Thr kinase sites, but can be replaced by other amino acids which, preferably, have similar characteristics. For example, a nearly invariant hydrophobic amino acid in the above amino acid consensus sequence would most likely be replaced by another hydrophobic amino acid. Ser/Thr kinase domains are described in, for example, Levin D.E. et al. (1990) Proc. Natl. Acad. Sci. USA 87:8272-76, the contents of which are incorporated herein by reference.

As used herein, the term "ATP-binding region" includes an amino acid sequence of about 20-40, preferably 20-30, and more preferably 25-30 amino acid residues in length, present in enzymes which activate their substrates by phosphorylation, and involved in binding adenosine triphosphate (ATP). ATP-binding regions preferably

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include the following amino acid consensus sequence: G-X-G-X-X-G-X(15-23)-K(SEQ ID NO:23). ATP-binding regions are described in, for example, Samuel K.P. et al. (1987) FEBS Let. 218(1): 81-86, the contents of which are incorporated herein by reference. Amino acid residues 40 to 63 of CSAPK-1 comprise an ATP-binding region.

Isolated proteins of the present invention, preferably CSAPK-1 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:1 or SEQ ID NO:3. As used herein, the term "sufficiently homologous" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain) amino acid residues or nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences share common structural domains or motifs and/or a common functional activity. For example, amino acid or nucleotide sequences which share common structural domains have at least 30%, 40%, or 50% homology, preferably 60% homology, more preferably 70%-80%, and even more preferably 90-95% homology across the amino acid sequences of the domains and contain at least one and preferably two structural domains or motifs, are defined herein as sufficiently homologous. Furthermore, amino acid or nucleotide sequences which share at least 30%, 40%, or 50%, preferably 60%, more preferably 70-80%, or 90-95% homology and share a common functional activity are defined herein as sufficiently homologous.

As used interchangeably herein a "CSAPK-1 activity", "biological activity of CSAPK-1" or "functional activity of CSAPK-1", refers to an activity exerted by a CSAPK-1 protein, polypeptide or nucleic acid molecule on a CSAPK-1 responsive cell or a CSAPK-1 protein substrate, as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-1 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-1 proteins and polypeptides having a CSAPK-1 activity. Preferred proteins are CSAPK-1 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region and, preferably, a CSAPK-1 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent

hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

The nucleotide sequence of the isolated human CSAPK-1 cDNA and the predicted amino acid sequence of the human CSAPK-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-1 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203308. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-1 gene, which is approximately 4137 nucleotides in length, encodes a protein having a molecular weight of approximately 34.7 kD and which is approximately 302 amino acid residues in length. The CSAPK-1 gene is expressed predominantly in heart, skeletal muscle, or the placenta.

B. The CSAPK-2 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-2 proteins, are identified based on the presence of at least one dual specificity kinase catalytic domain and at least one ATP-binding region. In yet a further embodiment, the isolated proteins of the present invention, preferably CSAPK-2 proteins, are identified based on the presence of at least one dual specificity kinase catalytic domain, at least one ATP-binding region, and at least one leucine zipper region.

As used herein, the term "dual specificity kinase catalytic domain" includes an amino acid sequence of 200-400 amino acid residues in length, preferably 200-300 amino acid residues in length, and more preferably 250-300 amino acid residues in length, which includes a kinase catalytic domain whose primary sequence is a hybrid between a serine/threonine kinase catalytic domain and a tyrosine kinase catalytic domain. Kinases containing the dual specificity kinase catalytic domain are capable of phosphorylating both serine/threonine and tyrosine residues. Preferably, a dual

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specificity kinase catalytic domain includes the following amino acid consensus sequence X9-G-X-G-X2-G-X-V-X12-K-X-(10-19)-E-X34-G-X40-H-R-D-X-K-X2-N-X17-K-X2-D-F-G-X19-W-X-A-P-E-X13-W-X7-E-X6-P-X36-C-W-X6-R-P-X-F-X14 (SEQ ID NO:24). Dual specificity kinase catalytic domains are described in, for example, Holzman L. B. et al. (1994) *J. Biol. Chem.* 269:30808-817, the contents of which are incorporated herein by reference. Amino acid residues 31-277 of the CSAPK-2 protein comprise a dual specificity kinase catalytic domain.

As used herein, the term "leucine zipper region" includes a protein domain which contains either leucine or isoleucine residues in every seventh position over a stretch of at least 20-30 amino acid residues, more preferably at least 20-25 amino acid residues, and most preferably at least 22 amino acid residues. Typically, the leucine zipper domain is rich in charged residues and forms a helical structure with a hydrophobic ridge of leucines or isoleucines down one face of the helix. The leucine zipper promotes dimerization through hydrophobic interactions between the leucine residues. Leucine zipper domains are described in, for example, Landschultz et al. (1988) Science 240: 1759-1764, the contents of which are incorporated herein by reference. Amino acid residues 294 to 322 of the CSAPK-2 protein comprise a leucine zipper region. Optionally, the leucine zipper region is found adjacent to a basic region. As used herein, the term "basic region" includes an amino acid sequence of 5-30, preferably 5-20, more preferably 5-15 amino acid residues in length, which is composed of a series of basic and hydrophobic amino acid residues, at either the N-terminus or the C-terminus of the leucine zipper region. For example, a 15 amino acid basic domain can contain at least 6, 8, or 10 basic residues and/or at least 1, 2, or 4 hydrophobic residues. Amino acid residues 407-421 of the CSAPK-2 protein comprise a basic region.

Isolated proteins of the present invention, preferably CSAPK-2 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:5 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:4 or SEQ ID NO:6.

As used interchangeably herein a "CSAPK-2 activity", "biological activity of CSAPK-2" or "functional activity of CSAPK-2", refers to an activity exerted by a CSAPK-2 protein, polypeptide or nucleic acid molecule on a CSAPK-2 responsive cell

or a CSAPK-2 protein substrate, as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-2 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-2 proteins and polypeptides having a CSAPK-2 activity. Preferred proteins are CSAPK-2 proteins having at least one dual specificity kinase catalytic domain and at least one ATP-binding region. Yet further preferred CSAPK-2 proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region, and at least one leucine zipper region. Preferred proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region and, preferably, a CSAPK-2 activity. Additional preferred proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

The nucleotide sequence of the isolated human CSAPK-2 cDNA and the predicted amino acid sequence of the human CSAPK-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-2 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203306. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-2 gene, which is approximately 2120 nucleotides in length, encodes a protein having a molecular weight of approximately 52.3 kD and which is approximately 455 amino acid residues in length. The CSAPK-2 gene is expressed predominantly in muscle, e.g., skeletal or cardiac muscle. Four splice variants of CSAPK-2 were also identified (see Figures 7-12).

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C. The CSAPK-3 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-3 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. The Ser/Thr kinase site and the ATP-binding region are described herein. Amino acid residues 5-164 of the CSAPK-3 protein comprise a Ser/Thr kinase domain.

Isolated proteins of the present invention, preferably CSAPK-3 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:8 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:7 or SEQ ID NO:9.

As used interchangeably herein a "CSAPK-3 activity", "biological activity of CSAPK-3" or "functional activity of CSAPK-3", refers to an activity exerted by a CSAPK-3 protein, polypeptide or nucleic acid molecule on a CSAPK-3 responsive cell or a CSAPK-3 protein substrate as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-3 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-3 proteins and polypeptides having a CSAPK-3 activity. Preferred proteins are CSAPK-3 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-3 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:7 or SEQ ID NO:9.

The nucleotide sequence of the isolated human CSAPK-3 cDNA and the predicted amino acid sequence of the human CSAPK-3 polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-3 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203309. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of

Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-3 gene, which is approximately 2454 nucleotides in length, encodes a protein having a molecular weight of approximately 66.8 kD and which is approximately 581 amino acid residues in length. The CSAPK-3 gene is expressed predominantly in heart, skeletal muscle, brain, placenta, lung, liver, kidney, and pancreas.

10 D. The CSAPK-4 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-4 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. The Ser/Thr kinase site and the ATP-binding region are described herein. Amino acid residues 11 to 75 of the CSAPK-4 protein comprise a Ser/Thr kinase domain.

Isolated proteins of the present invention, preferably CSAPK-4 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:11 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:10 or SEQ ID NO:12.

As used interchangeably herein a "CSAPK-4 activity", "biological activity of CSAPK-4" or "functional activity of CSAPK-4", refers to an activity exerted by a CSAPK-4 protein, polypeptide or nucleic acid molecule on a CSAPK-4 responsive cell or a CSAPK-4 protein substrate as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-4 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-4 proteins and polypeptides having a CSAPK-4 activity. Preferred proteins are CSAPK-4 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-4 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes

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under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:10 or SEQ ID NO:12.

The nucleotide sequence of the isolated human CSAPK-4 cDNA and the predicted amino acid sequence of the human CSAPK-4 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-4 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203307. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The human CSAPK-4 gene, which is approximately 1864 nucleotides in length, encodes a protein having a molecular weight of approximately 18.4 kD and which is approximately 160 amino acid residues in length. The CSAPK-4 gene is expressed predominantly in skeletal muscle.

E. The CSAPK-5 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-5 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. In yet another embodiment, the isolated proteins of the present invention, preferably CSAPK-5 proteins, are identified based on the presence of at least one Ser/Thr kinase site, and at least one ATP-binding region.

Isolated proteins of the present invention, preferably CSAPK-5 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:14 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:13 or SEQ ID NO:15.

As used interchangeably herein a "CSAPK-5 activity", "biological activity of CSAPK-5" or "functional activity of CSAPK-5", refers to an activity exerted by a CSAPK-5 protein, polypeptide or nucleic acid molecule on a CSAPK-5 responsive cell

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or a CSAPK-5 protein substrate as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-5 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-5 proteins and polypeptides having a CSAPK-5 activity. Preferred proteins are CSAPK-5 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-5 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:13 or SEQ ID NO:15.

The nucleotide sequence of the isolated human CSAPK-5 cDNA and the predicted amino acid sequence of the human CSAPK-5 polypeptide are shown in Figure 5 and in SEQ ID NOs:13 and 14, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-1 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203305. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The human CSAPK-5 gene, which is approximately 1333 nucleotides in length, encodes a protein having a molecular weight of approximately 51 kD and which is approximately 444 amino acid residues in length.

Various aspects of the invention are described in further detail in the following subsections:

I. Isolated Nucleic Acid Molecules

One aspect of the invenuon pertains to isolated nucleic acid molecules that

encode CSAPK proteins or biologically active portions thereof, as well as nucleic acid
fragments sufficient for use as hybridization probes to identify CSAPK-encoding nucleic

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acids (e.g., CSAPK mRNA) and fragments for use as PCR primers for the amplification or mutation of CSAPK nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (e.g., cDNA or genomic DNA) and RNA molecules (e.g., mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. For example, with regards to genomic DNA, the term "isolated" includes nucleic acid molecules which are separated from the chromosome with which the genomic DNA is naturally associated. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated CSAPK nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0 1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion thereof, can be isolated using standard molecular biology techniques and the sequence information provided herein. For example, using all or portion of the nucleic acid sequence of SEQ ID NO:1, or the nucleotide sequence of SEQ ID NO:3, as a hybridization probe, CSAPK nucleic acid molecules can be isolated using standard hybridization and cloning techniques (e.g., as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring

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Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

Moreover, a nucleic acid molecule encompassing all or a portion of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to CSAPK nucleotide sequences can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:1. The sequence of SEQ ID NO:1 corresponds to the partial human CSAPK-1 cDNA. This cDNA comprises sequences encoding the human CSAPK-1 protein (i.e., "the coding region", from nucleotides 297-1202), as well as 5' untranslated sequences (nucleotides 1-296) and 3' untranslated sequences (nucleotides 1203-4137). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:1 (e.g., nucleotides 297-1202, corresponding to SEQ ID NO:3).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:4. The sequence of SEQ ID NO:4 corresponds to the partial human CSAPK-2 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-2 protein (i.e., "the coding region", from nucleotides 47-1411), as well as 5' untranslated sequences (nucleotides 1-46) and 3' untranslated sequences (nucleotides 1412-2120). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:4 (e.g., nucleotides 47-

1411, corresponding to SEQ ID NO:6). In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:7. The sequence of SEQ ID NO:7 corresponds to the partial human CSAPK-3 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-3 protein (i.e., "the coding region", from nucleotides 51-1793), as well as 5' untranslated sequences (nucleotides 1-50) and 3' untranslated sequences (nucleotides 1794-2454). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:7 (e.g., nucleotides 51-1793, corresponding to SEQ ID NO:9).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:10. The sequence of SEQ ID NO:10 corresponds to the partial human CSAPK-4 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-4 protein (i.e., "the coding region", from nucleotides 275-754), as well as 5' untranslated sequences (nucleotides 1-274) and 3' untranslated sequences (nucleotides 755-1864). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:10 (e.g., nucleotides 275-754, corresponding to SEQ ID NO:12).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:13. The sequence of SEQ ID NO:13 corresponds to the partial human CSAPK-5 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-5 protein (i.e., "the coding region", from nucleotides 2-1333).

In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion of any of these nucleotide sequences. A nucleic acid molecule which is complementary to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ

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ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 is one which is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively, such that it can hybridize to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively, thereby forming a stable duplex.

In still another preferred embodiment, an isolated nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least about 50%, 54%, 55%, 60%, 62%, 65%, 70%, 75%, 78%, 80%, 85%, 86%, 90%, 95%, 97%, 98% or more homologous to the nucleotide sequence (e.g., to the entire length of the nucleotide sequence) shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion of any of these nucleotide sequences.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 for example a fragment which can be used as a probe or primer or a fragment encoding a biologically active portion of a CSAPK protein. The nucleotide sequence determined from the cloning of the CSAPK gene allows for the generation of probes and primers designed for use in identifying and/or cloning other CSAPK family members, as well as CSAPK homologues from other species. The probe/primer typically comprises substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12 or 15, preferably about 20 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, or 75 consecutive nucleotides of a sense sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID

NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 of an anti-sense sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 or of a naturally occurring allelic variant or mutant of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In an exemplary embodiment, a nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least 350, 400, 450, 500, 550, 600, 650, 700, 750, or 800 nucleotides in length and hybridizes under stringent hybridization conditions to a nucleic acid molecule of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

Probes based on the CSAPK nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments, the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissues which misexpress a CSAPK protein, such as by measuring a level of a CSAPK-encoding nucleic acid in a sample of cells from a subject e.g., detecting CSAPK mRNA levels or determining whether a genomic CSAPK gene has been mutated or deleted.

A nucleic acid fragment encoding a "biologically active portion of a CSAPK protein" can be prepared by isolating a portion of the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, which encodes a polypeptide having a CSAPK biological activity (the biological activities of the CSAPK proteins are described herein), expressing the encoded portion of the CSAPK protein (e.g., by

recombinant expression *in vitro*) and assessing the activity of the encoded portion of the CSAPK protein.

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 due to the degeneracy of the genetic code and, thus, encode the same CSAPK proteins as those encoded by the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14.

In addition to the CSAPK nucleotide sequences shown in SEQ ID NO:1, SEQ ID NO:3, SEO ID NO:4, SEO ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the CSAPK proteins may exist within a population (e.g., the human population). Such genetic polymorphism in the CSAPK genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which include an open reading frame encoding an CSAPK protein, preferably a mammalian CSAPK protein, and can further include non-coding regulatory sequences, and introns. Such natural allelic variations include both functional and non-functional CSAPK proteins and can typically result in 1-5% variance in the nucleotide sequence of a CSAPK gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in CSAPK genes that are the result of natural allelic variation and that do not alter the functional activity of a CSAPK protein are intended to be within the scope of the invention.

Moreover, nucleic acid molecules encoding other CSAPK family members and, thus, which have a nucleotide sequence which differs from the CSAPK sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are intended to be within the scope of the invention. For example, another CSAPK cDNA can be identified based on the nucleotide sequence of human CSAPK. Moreover, nucleic acid molecules encoding CSAPK proteins from different species, and thus which have a nucleotide sequence which differs from the CSAPK sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are intended to be within the scope of the invention. For example, a mouse CSAPK cDNA can be identified based on the nucleotide sequence of a human CSAPK.

Nucleic acid molecules corresponding to natural allelic variants and homologues of the CSAPK cDNAs of the invention can be isolated based on their homology to the CSAPK nucleic acids disclosed herein using the cDNAs disclosed herein, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15, 20, 25, 30 or more nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In other embodiment, the nucleic acid is at least 30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, or 600 nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 30%, 40%, 50%, or 60% homologous to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 70%, more preferably at least about 80%, even more preferably at least about 85%

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or 90% homologous to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50°C, 55°C, 60°C, or 65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In addition to naturally-occurring allelic variants of the CSAPK sequences that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, thereby leading to changes in the amino acid sequence of the encoded CSAPK proteins, without altering the functional ability of the CSAPK proteins. For example, nucleotide substitutions leading to amino acid substitutions at "nonessential" amino acid residues can be made in the sequence oSEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of CSAPK (e.g., the sequence of SEQ ID NO:2. SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14) without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the CSAPK proteins of the present invention, are predicted to be particularly unamenable to alteration. Furthermore, additional amino acid residues that are conserved between the

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CSAPK proteins of the present invention and other CSAPK family members are not likely to be amenable to alteration.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding CSAPK proteins that contain changes in amino acid residues that are not essential for activity. Such CSAPK proteins differ in amino acid sequence from SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, or SEQ ID NO:14).

An isolated nucleic acid molecule encoding a CSAPK protein homologous to the protein of SEO ID NO:2, SEO ID NO:5, SEO ID NO:8, SEO ID NO:11, or SEO ID NO:14 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15, respectively, such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15 by standard techniques, such as site-directed mutagenesis and PCRmediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histudine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine,

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phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in a CSAPK protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a CSAPK coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for CSAPK biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15, the encoded protein can be expressed recombinantly and the activity of the protein can be determined.

In a preferred embodiment, a mutant CSAPK protein can be assayed for the ability to: 1) regulate trasmission of signals from cellular receptors, e.g., cardiac cell growth factor receptors; 2) control entry of cells, e.g., cardiac cells, into mitosis; 3) modulate cellular differentiation; 4) modulate cell death; or 5) regulate cytoskeleton function, e.g., actin bundling.

In addition to the nucleic acid molecules encoding CSAPK proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire CSAPK coding strand, or only to a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding CSAPK. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (e.g., the coding region of human CSAPK-1 corresponds to SEQ ID NO:3). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding CSAPK. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

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Given the coding strand sequences encoding CSAPK disclosed herein (e.g., SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9, SEQ ID NO:12, or SEQ ID NO:15), antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of CSAPK mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding or noncoding region of CSAPK mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of CSAPK mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-Dgalactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5- oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (i.e., RNA transcribed from

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the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated in situ such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding a CSAPK protein to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention include direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an α-anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β-units, the strands run parallel to each other (Gaultier et al. (1987) *Nucleic Acids*. *Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to

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catalytically cleave CSAPK mRNA transcripts to thereby inhibit translation of CSAPK mRNA. A ribozyme having specificity for a CSAPK-encoding nucleic acid can be designed based upon the nucleotide sequence of a CSAPK cDNA disclosed herein (i.e., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19). For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a CSAPK-encoding mRNA. See, e.g., Cech et al. U.S. Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, CSAPK mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, e.g., Bartel, D. and Szostak, J.W. (1993) *Science* 261:1411-1418.

Alternatively, CSAPK gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the CSAPK (e.g., the CSAPK promoter and/or enhancers) to form triple helical structures that prevent transcription of the CSAPK gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. et al. (1992) *Ann. NY Acad Sci* 660:27-36; and Maher, L.J. (1992) *Bioassays* 14(12):807-15.

In yet another embodiment, the CSAPK nucleic acid molecules of the present invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acid molecules can be modified to generate peptide nucleic acids (see Hyrup B. et al. (1996) *Bioorganic & Medicinal Chemistry* 4 (1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup B. et al. (1996) *supra*; Perry-O'Keefe et al. Proc. Natl. Acad. Sci. 93: 14670-675.

PNAs of CSAPK nucleic acid molecules can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, for example, inducing transcription or translation arrest or inhibiting replication. PNAs of CSAPK nucleic acid molecules can also be used in the analysis of single base pair mutations in a gene, (e.g., by PNA-directed PCR clamping); as 'artificial restriction enzymes' when used in combination with other enzymes, (e.g., S1 nucleases (Hyrup B. (1996) supra)); or as probes or primers for DNA sequencing or hybridization (Hyrup B. et al. (1996) supra; Perry-O'Keefe supra).

In another embodiment, PNAs of CSAPK can be modified, (e.g., to enhance their stability or cellular uptake), by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of CSAPK nucleic acid molecules can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, (e.g., RNAse H and DNA polymerases), to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup B. (1996) supra). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup B. (1996) supra and Finn P.J. et al. (1996) Nucleic Acids Res. 24 (17): 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs, e.g., 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used as a between the PNA and the 5' end of DNA (Mag, M. et al. (1989) Nucleic Acid Res. 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn P.J. et al. (1996) supra). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser, K.H. et al. (1975) Bioorganic Med. Chem. Lett. 5: 1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) *Proc. Natl. Acad. Sci. US.* 86:6553-6556; Lemaitre et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (See, e.g., Krol et al. (1988) *Bio-Techniques* 6:958-976) or intercalating agents. (See, e.g., Zon (1988) *Pharm. Res.* 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, (e.g., a peptide, hybridization triggered cross-linking agent, transport agent, or hybridization-triggered cleavage agent).

II. Isolated CSAPK Proteins and Anti-CSAPK Antibodies

One aspect of the invention pertains to isolated CSAPK proteins, and biologically active portions thereof, as well as polypeptide fragments suitable for use as immunogens to raise anti-CSAPK antibodies. In one embodiment, native CSAPK proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, CSAPK proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, a CSAPK protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the CSAPK protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of CSAPK protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of CSAPK protein having less than about 30% (by dry weight) of non-CSAPK protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-CSAPK protein, still more preferably less than about 10% of non-CSAPK protein, and most preferably less than

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about 5% non-CSAPK protein. When the CSAPK protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, i.e., culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of CSAPK protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of CSAPK protein having less than about 30% (by dry weight) of chemical precursors or non-CSAPK chemicals, more preferably less than about 20% chemical precursors or non-CSAPK chemicals, still more preferably less than about 10% chemical precursors or non-CSAPK chemicals, and most preferably less than about 5% chemical precursors or non-CSAPK chemicals.

Biologically active portions of a CSAPK protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the CSAPK protein, e.g., the amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, which include less amino acids than the full length CSAPK proteins, and exhibit at least one activity of a CSAPK protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the CSAPK protein. A biologically active portion of a CSAPK protein can be a polypeptide which is, for example, at least 10, 25, 50, 100 or more amino acids in length.

In a preferred embodiment, the CSAPK protein has an amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14. In other embodiments, the CSAPK protein is substantially homologous to SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, and retains the functional activity of the protein of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail in subsection I above.

Accordingly, in another embodiment, the CSAPK protein is a protein which comprises

an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14).

To determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a preferred embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, or 90% of the length of the reference sequence (e.g., when aligning a second sequence to the CSAPK amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14 having 177 amino acid residues, at least 80, preferably at least 100, more preferably at least 120, even more preferably at least 140, and even more preferably at least 150, 160 or 170 amino acid residues are aligned). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a preferred embodiment, the percent identity between two amino acid sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using either a Blossom 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another preferred

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embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6.

The nucleic acid and protein sequences of the present invention can further be used as a "query sequence" to perform a search against public databases to, for example, identify other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (1990) *J. Mol. Biol.* 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences homologous to CSAPK nucleic acid molecules of the invention. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength = 3 to obtain amino acid sequences homologous to CSAPK protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., (1997) *Nucleic Acids Res* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used. See http://www.ncbi.nlm.nih.gov.

The invention also provides CSAPK chimeric or fusion proteins. As used herein, a CSAPK "chimeric protein" or "fusion protein" comprises a CSAPK polypeptide operatively linked to a non-CSAPK polypeptide. An "CSAPK polypeptide" refers to a polypeptide having an amino acid sequence corresponding to CSAPK, whereas a "non-CSAPK polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the CSAPK protein, e.g., a protein which is different from the CSAPK protein and which is derived from the same or a different organism. Within a CSAPK fusion protein the CSAPK polypeptide can correspond to all or a portion of a CSAPK protein. In a preferred embodiment, a CSAPK fusion protein comprises at least one biologically active portion of a CSAPK protein. In another preferred embodiment, a CSAPK fusion protein comprises at least two biologically active portions of a CSAPK protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the CSAPK polypeptide and the

non-CSAPK polypeptide are fused in-frame to each other. The non-CSAPK polypeptide can be fused to the N-terminus or C-terminus of the CSAPK polypeptide.

For example, in one embodiment, the fusion protein is a GST-CSAPK fusion protein in which the CSAPK sequences are fused to the C-terminus of the GST sequences. Such fusion proteins can facilitate the purification of recombinant CSAPK.

In another embodiment, the fusion protein is a CSAPK protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of CSAPK can be increased through use of a heterologous signal sequence.

The CSAPK fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject *in vivo*. The CSAPK fusion proteins can be used to affect the bioavailability of a CSAPK substrate. Use of CSAPK fusion proteins may be useful therapeutically for the treatment of cellular growth related disorders, e.g., cardiovascular disorers. Moreover, the CSAPK-fusion proteins of the invention can be used as immunogens to produce anti-CSAPK antibodies in a subject, to purify CSAPK ligands and in screening assays to identify molecules which inhibit the interaction of CSAPK with a CSAPK substrate.

Preferably, a CSAPK chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Current Protocols in Molecular Biology, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression vectors are commercially

available that already encode a fusion moiety (e.g., a GST polypeptide). A CSAPK-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the CSAPK protein.

The present invention also pertains to variants of the CSAPK proteins which function as either CSAPK agonists (mimetics) or as CSAPK antagonists. Variants of the CSAPK proteins can be generated by mutagenesis, e.g., discrete point mutation or truncation of a CSAPK protein. An agonist of the CSAPK proteins can retain substantially the same, or a subset, of the biological activities of the naturally occurring form of a CSAPK protein. An antagonist of a CSAPK protein can inhibit one or more of the activities of the naturally occurring form of the CSAPK protein by, for example, competitively modulating a cardiovascular system activity of a CSAPK protein. Thus, specific biological effects can be elicited by treatment with a variant of limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the protein has fewer side effects in a subject relative to treatment with the naturally occurring form of the CSAPK protein.

In one embodiment, variants of a CSAPK protein which function as either CSAPK agonists (mimetics) or as CSAPK antagonists can be identified by screening combinatorial libraries of mutants, e.g., truncation mutants, of a CSAPK protein for CSAPK protein agonist or antagonist activity. In one embodiment, a variegated library of CSAPK variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of CSAPK variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential CSAPK sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage display) containing the set of CSAPK sequences therein. There are a variety of methods which can be used to produce libraries of potential CSAPK variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding

the desired set of potential CSAPK sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, e.g., Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura et al. (1984) *Annu. Rev. Biochem.* 53:323; Itakura et al. (1984) *Science* 198:1056; Ike et al. (1983) *Nucleic Acid Res.* 11:477.

In addition, libraries of fragments of a CSAPK protein coding sequence can be used to generate a variegated population of CSAPK fragments for screening and subsequent selection of variants of a CSAPK protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of a CSAPK coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the CSAPK protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of CSAPK proteins. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recrusive ensemble mutagenesis (REM), a new technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify CSAPK variants (Arkin and Yourvan (1992) *Proc. Natl. Acad. Sci. USA* 89:7811-7815; Delgrave et al. (1993) *Protein Engineering* 6(3):327-331).

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In one embodiment, cell based assays can be exploited to analyze a variegated CSAPK library. For example, a library of expression vectors can be transfected into a cell line which ordinarily synthesizes and secretes CSAPK. The transfected cells are then cultured such that CSAPK and a particular mutant CSAPK are secreted and the effect of expression of the mutant on CSAPK activity in cell supernatants can be detected, e.g., by any of a number of enzymatic assays. Plasmid DNA can then be recovered from the cells which score for inhibition, or alternatively, potentiation of CSAPK activity, and the individual clones further characterized.

An isolated CSAPK protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind CSAPK using standard techniques for polyclonal and monoclonal antibody preparation. A full-length CSAPK protein can be used or, alternatively, the invention provides antigenic peptide fragments of CSAPK for use as immunogens. The antigenic peptide of CSAPK comprises at least 8 amino acid residues of the amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 and encompasses an epitope of CSAPK such that an antibody raised against the peptide forms a specific immune complex with CSAPK. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues.

Preferred epitopes encompassed by the antigenic peptide are regions of CSAPK that are located on the surface of the protein, e.g., hydrophilic regions.

A CSAPK immunogen typically is used to prepare antibodies by immunizing a suitable subject, (e.g., rabbit, goat, mouse or other mammal) with the immunogen. An appropriate immunogenic preparation can contain, for example, recombinantly expressed CSAPK protein or a chemically synthesized CSAPK polypeptide. The preparation can further include an adjuvant, such as Freund's complete or incomplete adjuvant, or similar immunostimulatory agent. Immunization of a suitable subject with an immunogenic CSAPK preparation induces a polyclonal anti-CSAPK antibody response.

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Accordingly, another aspect of the invention pertains to anti-CSAPK antibodies. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antigen binding site which specifically binds (immunoreacts with) an antigen, such as CSAPK. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind CSAPK. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of CSAPK. A monoclonal antibody composition thus typically displays a single binding affinity for a particular CSAPK protein with which it immunoreacts.

Polyclonal anti-CSAPK antibodies can be prepared as described above by immunizing a suitable subject with a CSAPK immunogen. The anti-CSAPK antibody 15 titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized CSAPK. If desired, the antibody molecules directed against CSAPK can be isolated from the mammal (e.g., from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction. At an appropriate time after immunization, e.g., when the anti-CSAPK antibody titers are highest, antibodyproducing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) Nature 256:495-497) (see also, Brown et al. (1981) JImmunol. 127:539-46; Brown et al. (1980) J. Biol. Chem .255:4980-83; Yeh et al. (1976) Proc. Natl. Acad. Sci. USA 76:2927-31; and Yeh et al. (1982) Int. J Cancer 29:269-75), the more recent human B cell hybridoma technique (Kozbor et al. (1983) Immunol Today 4:72), the EBV-hybridoma technique (Cole et al. (1985), Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing monoclonal antibody hybridomas is well known (see generally R. H. Kenneth, in Monoclonal Antibodies. A New Dimension In Biological

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Analyses, Plenum Publishing Corp., New York, New York (1980); E. A. Lerner (1981) Yale J. Biol. Med., 54:387-402; M. L. Gefter et al. (1977) Somatic Cell Genet.

3:231-36). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with a CSAPK immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds CSAPK.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating an anti-CSAPK monoclonal antibody (see, e.g., G. Galfre et al. (1977) Nature 266:55052; Gefter et al. Somatic Cell Genet., cited supra; Lerner, Yale J. Biol. Med., cited supra; Kenneth, Monoclonal Antibodies, cited supra). Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods which also would be useful. Typically, the immortal cell line (e.g., a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line. Preferred immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a number of myeloma cell lines can be used as a fusion partner according to standard techniques, e.g., the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days 25 because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind CSAPK, e.g., using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal anti-CSAPK antibody can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with CSAPK to thereby isolate immunoglobulin library members that bind

CSAPK. Kits for generating and screening phage display libraries are commercially available (e.g., the Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; and the Stratagene SurfZAPTM Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, Ladner et al. U.S. Patent No. 5,223,409; Kang et al. PCT International Publication No. WO 92/18619; Dower et al. PCT International Publication No. WO 91/17271; Winter et al. PCT International Publication WO 92/20791; Markland et al. PCT International Publication No. WO 92/15679; Breitling et al. PCT International Publication WO 93/01288; McCafferty et al. PCT International Publication No. WO 92/01047; Garrard 10 et al. PCT International Publication No. WO 92/09690; Ladner et al. PCT International Publication No. WO 90/02809; Fuchs et al. (1991) Bio/Technology 9:1370-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; Griffiths et al. (1993) EMBO J 12:725-734; Hawkins et al. (1992) J. Mol. Biol. 226:889-896; Clarkson et al. (1991) Nature 352:624-628; Gram et al. (1992) Proc. Natl. 15 Acad. Sci. USA 89:3576-3580; Garrad et al. (1991) Bio/Technology 9:1373-1377; Hoogenboom et al. (1991) Nuc. Acid Res. 19:4133-4137; Barbas et al. (1991) Proc. Natl. Acad. Sci. USA 88:7978-7982; and McCafferty et al. Nature (1990) 348:552-554.

Additionally, recombinant anti-CSAPK antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson et al. International Application No. PCT/US86/02269; Akira, et al. European Patent Application 184,187; Taniguchi, M., European Patent Application 171,496; Morrison et al. European Patent Application 173,494; Neuberger et al. PCT International Publication No. WO 86/01533; Cabilly et al. U.S. Patent No. 4,816,567; Cabilly et al. European Patent Application 125,023; Better et al. (1988) Science 240:1041-1043; Liu et al. (1987) Proc. Natl. Acad. Sci. USA 84:3439-3443; Liu et al. (1987) J Immunol. 139:3521-3526; Sun et al. (1987) Proc. Natl. Acad. Sci. USA 84:214-218; Nishimura et al. (1987) Canc Res. 47:999-1005; Wood et al. (1985) Nature 314:446-449; and Shaw

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et al. (1988) J. Natl. Cancer Inst. 80:1553-1559); Morrison, S. L. (1985) Science 229:1202-1207; Oi et al. (1986) BioTechniques 4:214; Winter U.S. Patent 5,225,539; Jones et al. (1986) Nature 321:552-525; Verhoeyan et al. (1988) Science 239:1534; and Beidler et al. (1988) J. Immunol. 141:4053-4060.

An anti-CSAPK antibody (e.g., monoclonal antibody) can be used to isolate CSAPK by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-CSAPK antibody can facilitate the purification of natural CSAPK from cells and of recombinantly produced CSAPK expressed in host cells. Moreover, an anti-CSAPK antibody can be used to detect CSAPK protein (e.g., in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the CSAPK protein. Anti-CSAPK antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, e.g., to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling (i.e., physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

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III. Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding a CSAPK protein (or a portion thereof). As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA

segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adenoassociated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (e.g., in an in vitro transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to includes promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; Gene Expression Technology Methods in Enzymology 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and those which direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the

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host cell to be transformed, the level of expression of protein desired, and the like. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., CSAPK proteins, mutant forms of CSAPK proteins, fusion proteins, and the like).

The recombinant-expression vectors of the invention can be designed for expression of CSAPK proteins in prokaryotic or eukaryotic cells. For example, CSAPK proteins can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith, D.B. and Johnson, K.S. (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Purified fusion proteins can be utilized in CSAPK activity assays, (e.g., direct assays or competitive assays described in detail below), or to generate antibodies specific for CSAPK proteins, for example. In a preferred embodiment, a CSAPK fusion

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protein expressed in a retroviral expression vector of the present invention can be utilized to infect bone marrow cells which are subsequently transplanted into irradiated recipients. The pathology of the subject recipient is then examined after sufficient time has passed (e.g., six (6) weeks).

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann et al., (1988) *Gene* 69:301-315) and pET 11d (Studier et al., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada et al., (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the CSAPK expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S cerivisae* include pYepSec1 (Baldari, et al., (1987) *Embo J.* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz et al., (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and picZ (InVitrogen Corp, San Diego, CA).

Alternatively, CSAPK proteins can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith et al. (1983) *Mol. Cell Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, B. (1987) Nature 329:840) and pMT2PC (Kaufman et al. (1987) EMBO J. 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements.

For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning. A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissuespecific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert et al. (1987) Genes Dev 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) Adv. Immunol. 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) EMBO J. 8:729-733) and immunoglobulins (Banerji et al. (1983) Cell 33:729-740; Queen and Baltimore (1983) Cell 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter, Byrne and Ruddle (1989) Proc. Natl. Acad. Sci. USA 86:5473-5477), pancreas-specific promoters (Edlund et al. (1985) Science 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentallyregulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) Science 249:374-379) and the α-fetoprotein promoter (Campes and Tilghman (1989) Genes Dev. 3:537-546).

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The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to CSAPK mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub, H. et al., Antisense RNA as a molecular tool for genetic analysis, *Reviews - Trends in Genetics*, Vol. 1(1) 1986.

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, a CSAPK protein can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including

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calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding a CSAPK protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (i.e., express) a CSAPK protein. Accordingly, the invention further provides methods for producing a CSAPK protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding a CSAPK protein has been introduced) in a suitable medium such that a CSAPK protein is produced. In another embodiment, the method further comprises isolating a CSAPK protein from the medium or the host cell.

The host cells of the invention can also be used to produce non-human transgenic animals. Transgenic mice expressing the CSAPK-2 nucleic acid sequence have been prepared (see Example 5). For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which CSAPK-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous CSAPK sequences have been introduced into their genome or homologous recombinant animals in which endogenous CSAPK sequences have been altered. Such animals are useful for studying the function and/or activity of a CSAPK

and for identifying and/or evaluating modulators of CSAPK activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous CSAPK gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing a CSAPKencoding nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by 15 microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The CSAPK cDNA sequence of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:13 can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of a human CSAPK gene, such as a mouse or rat CSAPK gene, can be used as a transgene. Alternatively, a CSAPK gene homologue, such as another CSAPK family member, can be isolated based on hybridization to the CSAPK cDNA sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 (described further in subsection I above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to a CSAPK transgene to direct expression of a CSAPK protein to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, 30 particularly animals such as mice, have become conventional in the art and are

described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., Manipulating the Mouse Embryo, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of a CSAPK transgene in its genome and/or expression of CSAPK mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding a CSAPK protein can further be bred to other transgenic animals carrying other

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of a CSAPK gene into which a deletion, addition or substitution has been introduced to thereby alter, e.g., functionally disrupt, the CSAPK gene. The CSAPK gene can be a human gene (e.g., the SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19), but more preferably, is a non-human homologue of a human CSAPK gene (e.g., a cDNA isolated by stringent hybridization with the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19). For example, a mouse CSAPK gene can be used to construct a homologous recombination vector suitable for altering an endogenous CSAPK gene in the mouse genome. In a preferred embodiment, the vector is designed such that, upon homologous recombination, the endogenous CSAPK gene is functionally disrupted (i.e., no longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous CSAPK gene is mutated or otherwise altered but still encodes a functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous CSAPK protein). In the homologous recombination vector, the altered portion of the CSAPK gene is flanked at its 5' and 3' ends by additional nucleic acid sequence of the CSAPK gene to allow for homologous recombination to occur between the exogenous CSAPK gene carried by the vector and an endogenous CSAPK gene in an

embryonic stem cell. The additional flanking CSAPK nucleic acid sequence is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the vector (see e.g., Thomas, K.R. and Capecchi, M. R. (1987) Cell 51:503 for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced CSAPK gene has homologously recombined with the endogenous CSAPK gene are selected (see, e.g., Li, E. et al. (1992) Cell 69:915). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see e.g., Bradley, A. in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E.J. Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing 15 homologous recombination vectors and homologous recombinant animals are described further in Bradley, A. (1991) Current Opinion in Biotechnology 2:823-829 and in PCT International Publication Nos.: WO 90/11354 by Le Mouellec et al.; WO 91/01140 by Smithies et al.; WO 92/0968 by Zijlstra et al.; and WO 93/04169 by Berns et al.

In another embodiment, transgenic non-humans animals can be produced which contain selected systems which allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman et al. (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, e.g., by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

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Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. (1997) *Nature* 385:810-813 and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G_0 phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The recontructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

IV. Pharmaceutical Compositions

The CSAPK nucleic acid molecules, CSAPK proteins, and anti-CSAPK antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents;

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antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor ELTM (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyetheylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a CSAPK protein or anti-CSAPK antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by

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incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

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The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a

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delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (i.e., the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Patent 5,328,470) or by stereotactic injection (see e.g., Chen et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, e.g., retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

V. Uses and Methods of the Invention

The nucleic acid molecules, proteins, protein homologues, and antibodies described herein can be used in one or more of the following methods: a) screening assays; b) predictive medicine (e.g., diagnostic assays, prognostic assays, monitoring

clinical trials, and pharmacogenetics); and c) methods of treatment (e.g., therapeutic and prophylactic). The isolated nucleic acid molecules of the invention can be used, for example, to express CSAPK protein (e.g., via a recombinant expression vector in a host cell in gene therapy applications), to detect CSAPK mRNA (e.g., in a biological sample) or a genetic alteration in a CSAPK gene, and to modulate CSAPK activity, as described further below. The CSAPK proteins can be used to treat disorders characterized by insufficient or excessive production of a CSAPK substrate or production of CSAPK inhibitors. In addition, the CSAPK proteins can be used to screen for naturally occurring CSAPK substrates, to screen for drugs or compounds which modulate CSAPK activity, as well as to treat disorders characterized by insufficient or excessive production of CSAPK protein or production of CSAPK protein forms which have decreased or aberrant activity compared to CSAPK wild type protein. Moreover, the anti-CSAPK antibodies of the invention can be used to detect and isolate CSAPK proteins, regulate the bioavailability of CSAPK proteins, and modulate CSAPK activity.

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A. Screening Assays:

The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, i.e., candidate or test compounds or agents (e.g., peptides, peptidomimetics, small molecules or other drugs) which bind to CSAPK proteins, have a stimulatory or inhibitory effect on, for example, CSAPK expression or CSAPK activity, or have a stimulatory or inhibitory effect on, for example, the expression or activity of a CSAPK substrate.

In one embodiment, the invention provides assays for screening candidate or test compounds which are substrates of a CSAPK protein or polypeptide or biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of a CSAPK protein or polypeptide or biologically active portion thereof, e.g., modulate the ability of CSAPK to interact with its cognate ligand. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring

deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:6909; Erb et al. (1994) Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al. (1994). J. Med Chem. 37:2678; Cho et al. (1993) Science 261:1303; Carrell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al. (1994) Angew. Chem Int. Ed. Engl. 33:2061; and in Gallop et al. (1994) J. Med. Chem. 37:1233.

Libraries of compounds may be presented in solution (e.g., Houghten (1992) Biotechniques 13:412-421), or on beads (Lam (1991) Nature 354:82-84), chips (Fodor (1993) Nature 364:555-556), bacteria (Ladner USP 5,223,409), spores (Ladner USP '409), plasmids (Cull et al. (1992) Proc Natl Acad Sci USA 89:1865-1869) or on phage (Scott and Smith (1990) Science 249:386-390); (Devlin (1990) Science 249:404-406); (Cwirla et al. (1990) Proc. Natl. Acad. Sci. 87:6378-6382); (Felici (1991) J. Mol. Biol. 222:301-310); (Ladner supra).

In another embodiment, an assay is a cell-based assay comprising contacting a cell expressing a CSAPK target molecule (e.g., a CSAPK phosphorylation substrate) with a test compound and determining the ability of the test compound to modulate (e.g. stimulate or inhibit) the activity of the CSAPK target molecule. Determining the ability of the test compound to modulate the activity of a CSAPK target molecule can be accomplished, for example, by determining the ability of the CSAPK protein to bind to or interact with the CSAPK target molecule, or by determining the ability of the CSAPK protein to phosphorylate the CSAPK target molecule.

The ability of the CSAPK protein to phosphorylate a CSAPK target molecule can be determined by, for example, an *in vitro* kinase assay. Briefly, a CSAPK target molecule, e.g., an immunoprecipitated CSAPK target molecule from a cell line expressing such a molecule, can be incubated with the CSAPK protein and radioactive ATP, e.g., $[\gamma^{-32}P]$ ATP, in a buffer containing MgCl₂ and MnCl₂, e.g., 10 mM MgCl₂

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and 5 mM MnCl₂. Following the incubation, the immunoprecipitated CSAPK target molecule can be separated by SDS-polyacrylamide gel electrophoresis under reducing conditions, transferred to a membrane, e.g., a PVDF membrane, and autoradiographed. The appearance of detectable bands on the autoradiograph indicates that the CSAPK substrate has been phosphorylated. Phosphoaminoacid analysis of the phosphorylated substrate can also be performed in order to determine which residues on the CSAPK substrate are phosphorylated. Briefly, the radiophosphorylated protein band can be excised from the SDS gel and subjected to partial acid hydrolysis. The products can then be separated by one-dimensional electrophoresis and analyzed on, for example, a phosphoimager and compared to ninhydrin-stained phosphoaminoacid standards.

Determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished by determining direct binding. Determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished, for example, by coupling the CSAPK protein with a radioisotope or enzymatic label such that binding of the CSAPK protein to a CSAPK target molecule can be determined by detecting the labeled CSAPK protein in a complex. For example, CSAPK molecules, e.g., CSAPK proteins, can be labeled with 125I, 35S, 14C, or 3H, either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, CSAPK molecules can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a compound to modulate the interaction between CSAPK and its target molecule, without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of CSAPK with its target molecule without the labeling of either CSAPK or the target molecule. McConnell, H. M. et al. (1992) *Science* 257:1906-1912. As used herein, a "microphysiometer" (e.g., Cytosensor) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between compound and receptor.

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In a preferred embodiment, determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (e.g., intracellular Ca²⁺, diacylglycerol, IP₃, etc.), detecting catalytic/enzymatic activity of the target an appropriate substrate, detecting the induction of a reporter gene (comprising a target-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, e.g., chloramphenicol acetyl transferase), or detecting a target-regulated cellular response.

In yet another embodiment, an assay of the present invention is a cell-free assay in which a CSAPK protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the CSAPK protein or biologically active portion thereof is determined. Binding of the test compound to the CSAPK protein can be determined either directly or indirectly as described above. In a preferred embodiment, the assay includes contacting the CSAPK protein or biologically active portion thereof with a known compound which binds CSAPK to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a CSAPK protein, wherein determining the ability of the test compound to interact with a CSAPK protein comprises determining the ability of the test compound to preferentially bind to CSAPK or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which a CSAPK protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate (e.g., stimulate or inhibit) the activity of the CSAPK protein or biologically active portion thereof is determined. Determining the ability of the test compound to modulate the activity of a CSAPK protein can be accomplished, for example, by determining the ability of the CSAPK protein to bind to a CSAPK target molecule by one of the methods described above for determining direct binding.

Determining the ability of the CSAPK protein to bind to a CSAPK target molecule can also be accomplished using a technology such as real-time Biomolecular Interaction Analysis (BIA). Sjolander, S. and Urbaniczky, C. (1991) Anal. Chem. 63:2338-2345

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and Szabo et al. (1995) Curr. Opin. Struct. Biol. 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (e.g., BIAcore). Changes in the optical phenomenon of surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

In an alternative embodiment, determining the ability of the test compound to modulate the activity of a CSAPK protein can be accomplished by determining the ability of the CSAPK protein to further modulate the activity of a CSAPK target molecule (e.g., a CSAPK mediated signal transduction pathway component). For example, the activity of the effector molecule on an appropriate target can be determined, or the binding of the effector to an appropriate target can be determined as previously described.

In yet another embodiment, the cell-free assay involves contacting a CSAPK protein or biologically active portion thereof with a known compound which binds the CSAPK protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the CSAPK protein, wherein determining the ability of the test compound to interact with the CSAPK protein comprises determining the ability of the CSAPK protein to preferentially bind to or modulate the activity of a CSAPK target molecule.

The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of proteins (e.g., CSAPK proteins or biologically active portions thereof, or receptors to which CSAPK binds). In the case of cell-free assays in which a membrane-bound form a protein is used (e.g., a cell surface CSAPK receptor) it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the protein is maintained in solution. Examples of such solubilizing agents include nonionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPSO), or N-dodecyl=N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either CSAPK or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to a CSAPK protein, or interaction of a CSAPK protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase/ CSAPK fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or CSAPK protein, and the mixture incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of CSAPK binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either a CSAPK protein or a CSAPK target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated CSAPK protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with CSAPK protein or target molecules but which do not interfere with binding of the CSAPK protein to its target molecule can be derivatized to the wells of the plate, and unbound target or CSAPK protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized

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complexes, include immunodetection of complexes using antibodies reactive with the CSAPK protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the CSAPK protein or target molecule.

In another embodiment, modulators of CSAPK expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of CSAPK mRNA or protein in the cell is determined. The level of expression of CSAPK mRNA or protein in the presence of the candidate compound is compared to the level of expression of CSAPK mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of CSAPK expression based on this comparison. For example, when expression of CSAPK mRNA or protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of CSAPK mRNA or protein expression. Alternatively, when expression of CSAPK mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of CSAPK mRNA or protein expression. The level of CSAPK mRNA or protein expression in the cells can be determined by methods described herein for detecting CSAPK mRNA or protein.

In yet another aspect of the invention, the CSAPK proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al. (1993) *Cell* 72:223-232; Madura et al. (1993) *J. Biol. Chem.* 268:12046-12054; Bartel et al. (1993) *Biotechniques* 14:920-924; Iwabuchi et al. (1993) *Oncogene* 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with CSAPK ("CSAPK-binding proteins" or "CSAPK-bp") and are involved in CSAPK activity. Such CSAPK-binding proteins are also likely to be involved in the propagation of signals by the CSAPK proteins or CSAPK targets as, for example, downstream elements of a CSAPK-mediated signaling pathway. Alternatively, such CSAPK-binding proteins are likely to be CSAPK inhibitors.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for a

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CSAPK protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming a CSAPK-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the CSAPK protein.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (e.g., a CSAPK modulating agent, an antisense CSAPK nucleic acid molecule, a CSAPK-specific antibody, or a CSAPK-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

B. Detection Assays

Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue typing); and (iii) aid in forensic identification of a biological sample. These applications are described in the subsections below.

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1. Chromosome Mapping

Once the sequence (or a portion of the sequence) of a gene has been isolated, this sequence can be used to map the location of the gene on a chromosome. This process is called chromosome mapping. Accordingly, portions or fragments of the CSAPK nucleotide sequences, described herein, can be used to map the location of the CSAPK genes on a chromosome. The mapping of the CSAPK sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease.

Briefly, CSAPK genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the CSAPK nucleotide sequences. Computer analysis of the CSAPK sequences can be used to predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the CSAPK sequences will yield an amplified fragment.

Somatic cell hybrids are prepared by fusing somatic cells from different mammals (e.g., human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of individual genes to specific human chromosomes. (D'Eustachio P. et al. (1983) *Science* 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycler. Using the CSAPK nucleotide sequences to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can similarly be

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used to map a 90, 1p, or 1v sequence to its chromosome include *in situ* hybridization (described in Fan, Y. et al. (1990) *Proc. Natl. Acad. Sci. USA*, 87:6223-27), prescreening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a DNA sequence to a metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a DNA sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good results at a reasonable amount of time. For a review of this technique, see Verma et al., Human Chromosomes: A Manual of Basic Techniques (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are preferred for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, Mendelian Inheritance in Man, available on-line through Johns Hopkins University Welch Medical Library). The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland, J. et al. (1987) Nature, 325:783-787.

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Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the CSAPK gene, can be determined. If a mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

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2. Tissue Typing

The CSAPK sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an alternative technique which determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the CSAPK nucleotide sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The CSAPK nucleotide sequences of the invention uniquely represent

portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals. The noncoding sequences of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers which each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences, such as those in SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9, SEQ ID NO:12, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

If a panel of reagents from CSAPK nucleotide sequences described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

3. Use of Partial CSAPK Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means for positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, e.g., hair or skin, or body fluids, e.g., blood, saliva, or semen found at a crime scene. The amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

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The sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e. another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Sequences targeted to noncoding regions of SEQ ID NO:1, SEQ ID NO:1, SEQ ID NO:16, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are particularly appropriate for this use as greater numbers of polymorphisms occur in the noncoding regions, making it easier to differentiate individuals using this technique. Examples of polynucleotide reagents include the CSAPK nucleotide sequences or portions thereof, e.g., fragments derived from the noncoding regions of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 having a length of at least 20 bases, preferably at least 30 bases.

The CSAPK nucleotide sequences described herein can further be used to provide polynucleotide reagents, e.g., labeled or labelable probes which can be used in, for example, an *in situ* hybridization technique, to identify a specific tissue, e.g., brain tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such CSAPK probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, e.g., CSAPK primers or probes can be used to screen tissue culture for contamination (i.e. screen for the presence of a mixture of different types of cells in a culture).

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C. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining CSAPK protein and/or nucleic acid expression as well as CSAPK activity,

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in the context of a biological sample (e.g., blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant CSAPK expression or activity. The invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with CSAPK protein, nucleic acid expression or activity. For example, mutations in a CSAPK gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby phophylactically treat an individual prior to the onset of a disorder characterized by or associated with CSAPK protein, nucleic acid expression or activity.

Another aspect of the invention pertains to monitoring the influence of agents (e.g., drugs, compounds) on the expression or activity of CSAPK in clinical trials.

These and other agents are described in further detail in the following sections.

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of CSAPK protein or nucleic acid in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting CSAPK protein or nucleic acid (e.g., mRNA, genomic DNA) that encodes CSAPK protein such that the presence of CSAPK protein or nucleic acid is detected in the biological sample. A preferred agent for detecting CSAPK mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to CSAPK mRNA or genomic DNA. The nucleic acid probe can be, for example, a human CSAPK nucleic acid, such as the nucleic acid of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to CSAPK mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

A preferred agent for detecting CSAPK protein is an antibody capable of binding to CSAPK protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof

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(e.g., Fab or F(ab')2) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect CSAPK mRNA, protein, or genomic DNA in a biological sample in vitro as well as in vivo. For example, in vitro techniques for detection of CSAPK mRNA include Northern hybridizations and in situ hybridizations. In vitro techniques for detection of CSAPK protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. In vitro techniques for detection of CSAPK genomic DNA include Southern hybridizations. Furthermore, in vivo techniques for detection of CSAPK protein include introducing into a subject a labeled anti-CSAPK antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a serum sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting CSAPK protein, mRNA, or genomic DNA, such that the presence of CSAPK protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of CSAPK protein, mRNA or genomic DNA in the control sample with the presence of CSAPK protein, mRNA or genomic DNA in the test sample.

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The invention also encompasses kits for detecting the presence of CSAPK in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting CSAPK protein or mRNA in a biological sample; means for determining the amount of CSAPK in the sample; and means for comparing the amount of CSAPK in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect CSAPK protein or nucleic acid.

2. Prognostic Assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant CSAPK expression or activity. For example, the assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing a disorder associated with CSAPK protein, nucleic acid expression or activity. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant CSAPK expression or activity in which a test sample is obtained from a subject and CSAPK protein or nucleic acid (e.g., mRNA, genomic DNA) is detected, wherein the presence of CSAPK protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant CSAPK expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest. For example, a test sample can be a biological fluid (e.g., serum), cell sample, or tissue.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant CSAPK expression or activity. Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant CSAPK expression or activity in which a test sample is obtained and CSAPK protein or nucleic acid expression or activity is detected (e.g., wherein the abundance of CSAPK protein or

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nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant CSAPK expression or activity).

The methods of the invention can also be used to detect genetic alterations in a CSAPK gene, thereby determining if a subject with the altered gene is at risk for a disorder associated with the CSAPK gene. In preferred embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding a CSAPK-protein, or the mis-expression of the CSAPK gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of 1) a deletion of one or more nucleotides from a CSAPK gene; 2) an addition of one or more nucleotides to a CSAPK gene; 3) a substitution of one or more nucleotides of a CSAPK gene, 4) a chromosomal rearrangement of a CSAPK gene; 5) an alteration in the level of a messenger RNA transcript of a CSAPK gene, 6) aberrant modification of a CSAPK gene, such as of the methylation pattern of the genomic DNA, 7) the presence of a non-wild type splicing pattern of a messenger RNA transcript of a CSAPK gene, 8) a non-wild type level of a CSAPK-protein, 9) allelic loss of a CSAPK gene, and 10) inappropriate post-translational modification of a CSAPK-protein. As described herein, there are a large number of assay techniques known in the art which can be used for detecting alterations in a CSAPK gene. A preferred biological sample is a tissue or serum sample isolated by conventional means from a subject, e.g., a cardiac tissue sample.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al. (1988) *Science* 241:1077-1080; and Nakazawa et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:360-364), the latter of which can be particularly useful for detecting point mutations in the CSAPK-gene (see Abravaya et al. (1995) *Nucleic Acids Res* .23:675-682). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to a CSAPK gene under conditions

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such that hybridization and amplification of the CSAPK-gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. et al., (1990) *Proc. Natl. Acad. Sci. USA* 87:1874-1878), transcriptional amplification system (Kwoh, D.Y. et al., (1989) *Proc. Natl. Acad. Sci. USA* 86:1173-1177), Q-Beta Replicase (Lizardi, P.M. et al. (1988) *Bio-Technology* 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in a CSAPK gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Patent No. 5,498,531) can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in CSAPK can be identified by hybridizing a sample and control nucleic acids, e.g., DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin, M.T. et al. (1996) *Human Mutation* 7: 244-255; Kozal, M.J. et al. (1996) *Nature Medicine* 2: 753-759). For example, genetic mutations in CSAPK can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. et al. supra. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential ovelapping probes. This step allows the identification

of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the CSAPK gene and detect mutations by comparing the sequence of the sample CSAPK with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on techniques developed by Maxam and Gilbert ((1977) Proc. Natl. Acad. Sci. USA 74:560) or Sanger ((1977) Proc. Natl. Acad. Sci. USA 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) Biotechniques 19:448), including sequencing by mass spectrometry (see, e.g., PCT International Publication No. WO 94/16101; Cohen et al. (1996) Adv. Chromatogr. 36:127-162; and Griffin et al. (1993) Appl Biochem. Biotechnol. 38:147-159).

Other methods for detecting mutations in the CSAPK gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers et al. (1985) Science 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes formed by hybridizing (labeled) RNA or DNA containing the wild-type CSAPK sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent which cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton et al. (1988) Proc Natl Acad Sci USA 85:4397; Saleeba et al.

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(1992) Methods Enzymol. 217:286-295. In a preferred embodiment, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in CSAPK cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994) *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on a CSAPK sequence, e.g., a wild-type CSAPK sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in CSAPK genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita et al. (1989) Proc Natl. Acad. Sci USA: 86:2766, see also Cotton (1993) Mutat Res 285:125-144; and Hayashi (1992) Genet Anal Tech Appl 9:73-79). Single-stranded DNA fragments of sample and control CSAPK nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In a preferred embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen et al. (1991) Trends Genet 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers et al. (1985) Nature 313:495). When

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DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys Chem* 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki et al. (1986) Nature 324:163); Saiki et al. (1989) Proc. Natl Acad. Sci USA 86:6230). Such allele specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology which depends on selective PCR amplification may be used in conjunction with the instant invention.

Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs et al. (1989) Nucleic Acids Res 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner et al. (1993) Tibtech 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini et al. (1992) Mol. Cell Probes 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) Proc. Natl. Acad. Sci USA 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing prepackaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, e.g., in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving a CSAPK gene.

Furthermore, any cell type or tissue in which CSAPK is expressed may be utilized in the prognostic assays described herein.

3. Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (e.g., drugs or compounds) on the expression or activity of a CSAPK protein can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase CSAPK gene expression, protein levels, or upregulate CSAPK activity, can be monitored in clinical trials of subjects exhibiting decreased CSAPK gene expression, protein levels, or downregulated CSAPK activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease CSAPK gene expression, protein levels, or downregulate CSAPK activity, can be monitored in clinical trials of subjects exhibiting increased CSAPK gene expression, protein levels, or upregulated CSAPK activity. In such clinical trials, the expression or activity of a CSAPK gene, and preferably, other genes that have been implicated in a disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes, including CSAPK, that are modulated in cells by treatment with an agent (e.g., compound, drug or small molecule) which modulates CSAPK activity (e.g., identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on a CSAPK associated disorder, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of CSAPK and other genes implicated in the CSAPK associated disorder, respectively. The levels of gene expression (i.e., a gene expression pattern) can be quantified by Northern blot analysis or RT-PCR, as described herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of CSAPK or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be

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determined before, and at various points during treatment of the individual with the agent.

In a preferred embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) comprising the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of a CSAPK protein, mRNA, or genomic DNA in the pre-administration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the CSAPK protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the CSAPK protein, mRNA, or genomic DNA in the pre-administration sample with the CSAPK protein, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of CSAPK to higher levels than detected, i.e., to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of CSAPK to lower levels than detected, i.e. to decrease the effectiveness of the agent. According to such an embodiment, CSAPK expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

C. Methods of Treatment:

The present invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant CSAPK expression or activity. With regards to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics.

"Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market. More specifically, the term refers the study of

how a patient's genes determine his or her response to a drug (e.g., a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with either the CSAPK molecules of the present invention or CSAPK modulators according to that individual's drug response genotype. Pharmacogenomics allows a clinician or physician to target prophylactic or therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug-related side effects.

1. Prophylactic Methods

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In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with an aberrant CSAPK expression or activity, by administering to the subject a CSAPK or an agent which modulates CSAPK expression or at least one CSAPK activity. Subjects at risk for a disease which is caused or contributed to by aberrant CSAPK expression or activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the CSAPK aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of CSAPK aberrancy, for example, a CSAPK, CSAPK agonist or CSAPK antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating CSAPK expression or activity for therapeutic purposes. Accordingly, in an exemplary embodiment, the modulatory method of the invention involves contacting a cell with a CSAPK or agent that modulates one or more of the activities of CSAPK protein activity associated with the cell. An agent that modulates CSAPK protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of a CSAPK protein (e.g., a CSAPK phosphorylation substrate), a CSAPK

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antibody, a CSAPK agonist or antagonist, a peptidomimetic of a CSAPK agonist or antagonist, or other small molecule. In one embodiment, the agent stimulates one or more CSAPK activities. Examples of such stimulatory agents include active CSAPK protein and a nucleic acid molecule encoding CSAPK that has been introduced into the cell. In another embodiment, the agent inhibits one or more CSAPK activities. Examples of such inhibitory agents include antisense CSAPK nucleic acid molecules, anti-CSAPK antibodies, and CSAPK inhibitors. These modulatory methods can be performed in vitro (e.g., by culturing the cell with the agent) or, alternatively, in vivo (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of a CSAPK protein or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) CSAPK expression or activity. In another embodiment, the method involves administering a CSAPK protein or nucleic acid molecule as therapy to compensate for reduced or aberrant CSAPK expression or activity.

Stimulation of CSAPK activity is desirable in situations in which CSAPK is abnormally downregulated and/or in which increased CSAPK activity is likely to have a beneficial effect. For example, stimulation of CSAPK activity is desirable in situations in which a CSAPK is downregulated and/or in which increased CSAPK activity is likely to have a beneficial effect. Likewise, inhibition of CSAPK activity is desirable in situations in which CSAPK is abnormally upregulated and/or in which decreased CSAPK activity is likely to have a beneficial effect.

25 3. Pharmacogenomics

The CSAPK molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on CSAPK activity (e.g., CSAPK gene expression) as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) disorders (e.g., cardiovascular disorders such as congestive heart failure) associated with aberrant CSAPK activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship

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between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer a CSAPK molecule or CSAPK modulator as well as tailoring the dosage and/or therapeutic regimen of treatment with a CSAPK molecule or CSAPK modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, for example, Eichelbaum, M. et al. (1996) Clin Exp. Pharmacol. Physiol 23(10-11):983-985 and Linder, M.W. et al. (1997) Clin Chem. 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (e.g., a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants.) Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a

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"SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict a drug response. According to this method, if a gene that encodes a drug target is known (e.g., a CSAPK protein or CSAPK receptor of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (e.g., N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

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Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (e.g., a CSAPK molecule or CSAPK modulator of the present invention) can give an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with a CSAPK molecule or CSAPK modulator, such as a modulator identified by one of the exemplary screening assays described herein.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the Figures and the Sequence Listing, are incorporated herein by reference.

EXAMPLES

20 Example 1: Identification and Characterization Human CSAPK cDNA

In this example, the identification and characterization of the genes encoding human CSAPK-1, CSAPK-2, CSAPK-3, CSAPK-4, and CSAPK-5 (also referred to as pjchrb014g05, pjchrb086g01, pJlhba191a08, pjchrb002d04, and pjchrb155a01, respectively) is described. Four splice variants of CSAPK-2 were also identified, referred to herein as CSAPK-2a, CSAK-2b, CSAPK-2c, and CSAPK-2d.

Isolation of the human CSAPK cDNA

The invention is based, at least in part, on the discovery of five human genes encoding members of the CSAPK family. The human CSAPK family members were isolated from cDNA libraries which were prepared from tissue obtained from subjects suffering from congestive heart failure of ischemic and idiopathic origin. Briefly, a

cardiac tissue sample was obtained from a biopsy of a patient suffering from congestive heart failure. mRNA was isolated from the cardiac tissue and a cDNA library was prepared therefrom using art known methods (described in, for example, *Molecular Cloning A Laboratory Manual*, 2nd Ed., ed. by Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory Press: 1989). Positive clones were isolated following comparison to homologs in public protein databases, including a comparison with known kinases and/or examination of the sequence for protein motifs of kinases.

The sequences of the positive clones were determined and found to contain open reading frames. The nucleotide sequence encoding the human CSAPK-1 protein is shown in Figure 1 and is set forth as SEQ ID NO:1. The protein encoded by this nucleic acid comprises about 302 amino acids and has the amino acid sequence shown in Figure 1 and set forth as SEQ ID NO:2. The coding region (open reading frame) of SEQ ID NO:1 is set forth as SEQ ID NO:3. The clone comprising the entire coding region of human CSAPK-1 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203308.

The nucleotide sequence encoding the human CSAPK-2 protein is shown in Figure 2 and is set forth as SEQ ID NO:4. The protein encoded by this nucleic acid comprises about 455 amino acids and has the amino acid sequence shown in Figure 2 and set forth as SEQ ID NO:5. The coding region (open reading frame) of SEQ ID NO:4 is set forth as SEQ ID NO:6. The clone comprising the entire coding region of human CSAPK-2 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209 on October 27, 1998, and assigned Accession No. 203306.

Four splice variants of CSAPK-2 were also identified. These variants are schematically depicted in Figure 7 (for the cDNA) and Figure 8 (for the protein). The start methionines and in-frame stop codons are indicated for each splice variant clone, in relation to the CSAPK-2 clone shown at the top. The dual specificity kinase catalytic domain is also indicated.

The CSAPK-2a (clone 1.1) nucleotide sequence (which includes parts of clones zchr86SA004 and jchrb086g01) is shown in Figure 9 and set forth as SEQ ID NO:16.

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The CSAPK-2b (clone 2) nucleotide sequence (which includes parts of clones jthrc169f03, jthrc089d05, and jchrb086g01) is shown in Figure 10 and set forth as SEQ ID NO:17. This clone was identified and/or is expressed in pulmonary artery smooth muscle.

The CSAPK-2c (clone 3) nucleotide sequence (which includes parts of clones jthvb025c05, jchrb116b08, jthka142dfl, and jchrb086g01) is shown in Figure 11 and set forth as SEQ ID NO:18. This clone was identified and/or is expressed in mammary epithelial cells and in keratinocytes.

The CSAPK-2d (clone 4) nucleotide sequence (which includes parts of clones johnc051a01 and jchrb086g01) is shown in Figure 12 and set forth as SEQ ID NO:19. This clone was identified and/or is expressed in colon to liver metastasis tissues.

The nucleotide sequence encoding the human CSAPK-3 protein is shown in Figure 3 and is set forth as SEQ ID NO:7. The protein encoded by this nucleic acid comprises about 581 amino acids and has the amino acid sequence shown in Figure 3 and set forth as SEQ ID NO:8. The coding region (open reading frame) of SEQ ID NO:7 is set forth as SEQ ID NO:9. The clone comprising the entire coding region of human CSAPK-3 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203309.

The nucleotide sequence encoding the human CSAPK-4 protein is shown in Figure 4 and is set forth as SEQ ID NO:10. The protein encoded by this nucleic acid comprises about 160 amino acids and has the amino acid sequence shown in Figure 4 and set forth as SEQ ID NO:11. The coding region (open reading frame) of SEQ ID NO:10 is set forth as SEQ ID NO:12. The clone comprising the entire coding region of human CSAPK-4 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203307.

The nucleotide sequence encoding the human CSAPK-5 protein is shown in Figure 5 and is set forth as SEQ ID NO:13. The protein encoded by this nucleic acid comprises about 444 amino acids and has the amino acid sequence shown in Figure 5 and set forth as SEQ ID NO:14. The coding region (open reading frame) of SEQ ID

NO:13 is set forth as SEQ ID NO:15. The clone comprising the entire coding region of human CSAPK-5 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203305.

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Analysis of Human CSAPK Molecule

A BLASTN 1.4.9 search, using a score of 100 and a word length of 12 (Altschul et al. (1990) *J. Mol. Biol.* 215:403) of the nucleotide sequence of human CSAPK-1 revealed that CSAPK-1 is similar to the human protein kinase HPK-1 coding sequence (Accession No. V23831). This nucleic acid molecule is approximately 70% identical to CSAPK-1, over nucleotides 388-1214.

CSAPK-2 is similar to the human MST mRNA for serine/threonine kinase (Accession No. Z48615). This nucleic acid molecule is approximately 54% identical to CSAPK-2, over nucleotides 482-805.

CSAPK-3 is similar to the human cDNA clone IMAGE:1257327 (Accession No. AA746653). This nucleic acid molecule is approximately 99% identical to CSAPK-3, over nucleotides 2068-2430.

CSAPK-4 is similar to the Mus musculus cDNA clone 902193 (Accession No. AA516800) This nucleic acid molecule is approximately 86% identical to CSAPK-4, over nucleotides 321-755.

CSAPK-5 is similar to the non-annotated human mRNA for KIAA0551 protein (Accession No. AB011123). This nucleic acid molecule is approximately 100% identical to CSAPK-5, over nucleotides 1-1333.

25 Tissue Distribution of CSAPK mRNA

This Example describes the tissue distribution of CSAPK mRNA, as determined by Northern blot hybridizations.

Northern blot hybridizations with the various RNA samples were performed under standard conditions and washed under stringent conditions, i.e., 0.2 X SSC at 65° C. The DNA probe was radioactively labeled with ³²P-dCTP using the Prime-It kit (Stratagene, La Jolla, CA) according to the instructions of the supplier. Filters

containing human mRNA (MultiTissue Northern I and MultiTissue Northern II from Clontech, Palo Alto, CA) were probed in ExpressHyb hybridization solution (Clontech) and washed at high stringency according to manufacturer's recommendations.

CSAPK-1 message was detected predominantly in the heart, skeletal muscle, or the placenta. CSAPK-2 message was detected predominantly in muscle, e.g., skeletal or cardiac muscle. CSAPK-3 message was detected predominantly in heart, skeletal muscle, brain, placenta, lung, liver, kidney, and pancreas. CSAPK-4 message was detected predominantly in skeletal muscle.

10 Example 2: Expression of Recombinant Protein in Bacterial Cells

In this example, CSAPK is expressed as a recombinant glutathione-S-transferase (GST) fusion polypeptide in *E. coli* and the fusion polypeptide is isolated and characterized. Specifically, CSAPK is fused to GST and this fusion polypeptide is expressed in *E. coli*, e.g., strain PEB199. Expression of the GST-CSAPK fusion protein in PEB199 is induced with IPTG. The recombinant fusion polypeptide is purified from crude bacterial lysates of the induced PEB199 strain by affinity chromatography on glutathione beads. Using polyacrylamide gel electrophoretic analysis of the polypeptide purified from the bacterial lysates, the molecular weight of the resultant fusion polypeptide is determined.

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Example 3: Expression of Recombinant CSAPK Protein in COS Cells

To express the CSAPK gene in COS cells, the pcDNA/Amp vector by Invitrogen Corporation (San Diego, CA) is used. This vector contains an SV40 origin of replication, an ampicillin resistance gene, an *E. coli* replication origin, a CMV promoter followed by a polylinker region, and an SV40 intron and polyadenylation site. A DNA fragment encoding the entire CSAPK protein and an HA tag (Wilson et al. (1984) *Cell* 37:767) or a FLAG tag fused in-frame to its 3' end of the fragment is cloned into the polylinker region of the vector, thereby placing the expression of the recombinant protein under the control of the CMV promoter.

To construct the plasmid, the CSAPK DNA sequence is amplified by PCR using two primers. The 5' primer contains the restriction site of interest followed by

approximately twenty nucleotides of the CSAPK coding sequence starting from the initiation codon; the 3' end sequence contains complementary sequences to the other restriction site of interest, a translation stop codon, the HA tag or FLAG tag and the last 20 nucleotides of the CSAPK coding sequence. The PCR amplified fragment and the pCDNA/Amp vector are digested with the appropriate restriction enzymes and the vector is dephosphorylated using the CIAP enzyme (New England Biolabs, Beverly, MA). Preferably the two restriction sites chosen are different so that the CSAPK gene is inserted in the correct orientation. The ligation mixture is transformed into *E. coli* cells (strains HB101, DH5a, SURE, available from Stratagene Cloning Systems, La Jolla, CA, can be used), the transformed culture is plated on ampicillin media plates, and resistant colonies are selected. Plasmid DNA is isolated from transformants and examined by restriction analysis for the presence of the correct fragment.

COS cells are subsequently transfected with the CSAPK-pcDNA/Amp plasmid DNA using the calcium phosphate or calcium chloride co-precipitation methods, DEAE-dextran-mediated transfection, lipofection, or electroporation. Other suitable methods for transfecting host cells can be found in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory,* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989. The expression of the CSAPK polypeptide is detected by radiolabelling (35S-methionine or 35S-cysteine available from NEN, Boston, MA, can be used) and immunoprecipitation (Harlow, E. and Lane, D. *Antibodies: A Laboratory Manual,* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1988) using an HA specific monoclonal antibody. Briefly, the cells are labelled for 8 hours with 35S-methionine (or 35S-cysteine). The culture media are then collected and the cells are lysed using detergents (RIPA buffer, 150 mM NaCl, 1% NP-40, 0.1% SDS, 0.5% DOC, 50 mM Tris, pH 7.5). Both the cell lysate and the culture media are precipitated with an HA specific monoclonal antibody. Precipitated polypeptides are then analyzed by SDS-PAGE.

Alternatively, DNA containing the CSAPK coding sequence is cloned directly into the polylinker of the pCDNA/Amp vector using the appropriate restriction sites.

The resulting plasmid is transfected into COS cells in the manner described above, and

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the expression of the CSAPK polypeptide is detected by radiolabelling and immunoprecipitation using a CSAPK specific monoclonal antibody.

Example 4: Chromosomal Mapping of CSAPK-2 (MLK)

In order to chromosomally map the CSAPK-2 (MLK) gene, the following primers were designed based on the sequence of jchrb086g01 (CSAPK-2): Forward-5'-GTCCGGTGGAAGTATAATACTTTGTC-3' (SEQ ID NO:20) and Reverse-5'-CTATTTTGAGGAGCTTCTTTACAGCC-3' (SEQ ID NO:21). Using these primers and PCR, a 185 bp product was amplified from human control cell line DNA and from control hamster cell line DNA. These primers were subsequently used used to amplify the 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel.

The PCR reactions were performed by using 5μl of template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR Buffer, 1.2μl Pharmacia dNTP mix (2.5mM), 1.15μl Forward primer 6.6μM, 1.15μl Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq (05U/μl) (Hot Start) and subjecting the reaction mix to 95°C for 10 minutes, followed by 35 Cycles of 94°C for 40 minutes, 55°C for 40 minutes, 72°C for 40 minutes, followed by 72°C for 5 minutes.

The products of the PCR reaction were run on 2% agarose gels, post-stained with SYBR Gold (1:10,000 dilution in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

RH linkage analysis was performed using the Map Manager QTb27 software package.

Clone jchrb086g01 (CSAPK-2 or MLK) was found to map to human

chromosome 2, 16.5 cR₃₀₀₀ telomeric to the Whitehead Institute framework marker WI-5538, and 16.5 cR₃₀₀₀ centromeric of the Whitehead framework marker WI-5555. LOD

scores for linkage were 15.3 for both WI-5538 and WI-5555. This region corresponds to the cytogenetic location 2q23-31.

Example 5: Generation of CSAPK-2 (MLK) Transgenic Animals

An N-terminal His-tagged MLK (CSAPK-2) cDNA was ligated to the cardiacspecific α-myosin heavy chain promoter and the human growth hormone polyadenylation signal sequence using art known techniques (see Figure 6). This construct was microinjected into the male pronucleus of fertilized mouse eggs, which were subsequently implanted into a pseudopregnant foster mother. Transgenic animals were identified by Southern blot analysis and founder mice were bred with nontransgenic cohorts for propagation of individual lines.

Equivalents

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Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

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What is claimed:

- 1. An isolated nucleic acid molecule selected from the group consisting of:
- (a) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:1;
 - (b) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:3;
 - (c) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:4;
- (d) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:6;
 - (e) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:7;
 - (f) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:9;
 - (g) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:10;
 - (h) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:12;
- 20 (i) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:13;
 - (j) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:15;
- (k) a nucleic acid molecule comprising the nucleotide sequence set forthin SEQ ID NO:16;
 - (l) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:17;
 - (m) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:18; and
- 30 (n) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:19.

- An isolated nucleic acid molecule selected from the group consisting of:
 (a) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2;
 - (b) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:5;
 - (c) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:8;
 - (d) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:11; and
- (e) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:14.
- An isolated nucleic acid molecule selected from the group consisting of:

 (a) a nucleic acid molecule comprising the nucleotide sequence contained

 n the plasmid deposited with ATCC® as Accession Number 203305;
 (b) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203306;
- (c) a nucleic acid molecule comprising the nucleotide sequence

 contained in the plasmid deposited with ATCC® as Accession Number

 203307;
 - (d) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203308; and
- 25 (e) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203309.

- An isolated nucleic acid molecule selected from the group consisting of:

 (a) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:1 or 3 under stringent conditions;
 (b) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:5, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:4, 6, 16, 17, 18, or 19 under stringent
- 10 conditions;

- (c) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:8, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:7 or 9 under stringent conditions;
 (d) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:10 or 12 under stringent conditions; and
- (e) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:14, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:13 or 15 under stringent conditions.

5. An isolated nucleic acid molecule selected from the group consisting of: a) a nucleic acid molecule comprising a nucleotide sequence which is at least 60% homologous to the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 15, 16, 17, 18, or 19, or a complement thereof; 5 b) a nucleic acid molecule comprising a fragment of at least 200 nucleotides of a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19, or a complement thereof; c) a nucleic acid molecule which encodes a polypeptide comprising an amino acid sequence at least about 60% homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14; and 10 d) a nucleic acid molecule which encodes a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14, wherein the fragment comprises at least 15 contiguous amino acid residues of the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.

- 6. An isolated nucleic acid molecule which hybridizes to the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 under stringent conditions.
- 7. An isolated nucleic acid molecule comprising a nucleotide sequence which is complementary to the nucleotide sequence of the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5.
- 8. An isolated nucleic acid molecule comprising the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5, and a nucleotide sequence encoding a heterologous
 25 polypeptide.
 - A vector comprising the nucleic acid molecule of any one of claims 1, 2, 3, 4, or
- 30 10. The vector of claim 9, which is an expression vector.

- 11. A host cell transfected with the vector of claim 9.
- 12. A method of producing a polypeptide comprising culturing a host cell transfected with the vector of claim 9 in an appropriate culture medium to, thereby, produce the polypeptide.
- 13. An isolated polypeptide selected from the group consisting of:
 a) a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14, wherein the fragment comprises at least 15 contiguous
 amino acids of SEQ ID NO:2, 5, 8, 11, or 14;
 - b) a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14, wherein the polypeptide is encoded by a nucleic acid molecule which hybridizes to a nucleic acid molecule comprising SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19 under stringent conditions;
 - c) a polypeptide which is encoded by a nucleic acid molecule comprising a nucleotide sequence which is at least 60 % homologous to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19;
- d) a polypeptide comprising an amino acid sequence which is at least 60% homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.
 - 14. The isolated polypeptide of claim 13 comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.
 - 15. The polypeptide of claim 13, further comprising heterologous amino acid sequences.
 - 16. An antibody which selectively binds to a polypeptide of claim 13.

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- 17. A method for detecting the presence of a polypeptide of claim 13 in a sample comprising:
 - a) contacting the sample with a compound which selectively binds to the polypeptide; and
- b) determining whether the compound binds to the polypeptide in the sample to thereby detect the presence of a polypeptide of claim 13 in the sample.
 - 18. The method of claim 17, wherein the compound which binds to the polypeptide is an antibody.

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- 19. A kit comprising a compound which selectively binds to a polypeptide of claim 13 and instructions for use.
- 20. A method for detecting the presence of a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 in a sample comprising:
 - a) contacting the sample with a nucleic acid probe or primer which selectively hybridizes to the nucleic acid molecule; and
 - b) determining whether the nucleic acid probe or primer binds to a nucleic acid molecule in the sample to thereby detect the presence of a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 in the sample.
 - 21. The method of claim 20, wherein the sample comprises mRNA molecules and is contacted with a nucleic acid probe.
- 25 22. A kit comprising a compound which selectively hybridizes to a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 and instructions for use.

- 23. A method for identifying a compound which binds to a polypeptide of claim 13 comprising:
 - a) contacting the polypeptide, or a cell expressing the polypeptide with a test compound; and
- b) determining whether the polypeptide binds to the test compound.
 - 24. The method of claim 23, wherein the binding of the test compound to the polypeptide is detected by a method selected from the group consisting of:
 - a) detection of binding by direct detection of test compound/polypeptide binding;
 - b) detection of binding using a competition binding assay; and
 - c) detection of binding using an assay for CSAPK activity.
 - 25. A method for modulating the activity of a polypeptide of claim 13 comprising contacting the polypeptide or a cell expressing the polypeptide with a compound which binds to the polypeptide in a sufficient concentration to modulate the activity of the polypeptide.
 - 26. A method for identifying a compound which modulates the activity of a polypeptide of claim 13 comprising:
- a) contacting a polypeptide of claim 13 with a test compound; and
 - b) determining the effect of the test compound on the activity of the polypeptide to thereby identify a compound which modulates the activity of the polypeptide.

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- TCGACCCACGCGTCCGGGAGGATCGGGGAGTCGCGGAGGATGGGCCGCCTAGGCTCGCACTCCGGACGCGCCTCGC

- TGGATGAGCAATCACAAGGAATGCAAGGGCCACCTGTTCCTCAGTTCCAACCACAGAAGGCCTTACGA CCGGATATGG
- AAGCACGTGCTGATTGCATCAAAGAAATAGATCTTCTTAAGCAACTCAACCATCCAAATGTAATAAAA TATTATGCAT
- CATTCATTGAAGATAATGAACTAAACATAGTTTTGGAACTAGCAGATGCTGGCGACCTATCCA GAATGATCAAGCATTTTAAGAAGCAAAAGAGGCTAATTCCTGAAAGAACTGTTTGGAAGTATTTTGTT CAGCTTTGCA
- GTGCATTGGAACACATGCATTCTCGAAGAGTCATGCATAGAGATATAAAACCAGCTAATGTGTTCATTACAGCCACTG
- GGGTGGTAAAACTTGGAGATCTTGGGCTTGGCCGGTTTTTCAGCTCAAAAACCACAGCTGCAC ATTCTTTAGTTGGTACGCCTTATTACATGTCTCCAGAGAGAATACATGAAAATGGATACAACTTCAAA TCTGACATCT
- GGTCTCTTGGCTGTCTACTATATGAGATGGCTGCATTACAAAGTCCTTTCTATGGTGACAAAATGAAT
 TTATACTCAC
- AGAGGATGCATGCACTGCAAGCAGCTAAACATGCAAGATCATGAAGAGTGTAACCAAAGTAATT GAAAGTATTT
- TGTGCAAAGTCGTACCTSCCCATTTATGTCTGGGTGTTAAGATTAATATTTCAGAGCTAGTGT GCTCTGAATCCTTAACCAGTTTTCATATAAGCTTCATTTTGTACCAGTCACCTAAATCACCTCCTTGC AACCCCCAA
- TGACTTTGGAATAACTGAATTGCATGTTAGGAGAGAAAATGAAACATGATGGTTTTGAATGGCTAAAG GTTTATAGAA
- TTTATGGACATTGAGACACAATTGTGAACTTTTGTGAAGATTTTATTTTTAAACGTTTGAAGTA
 CTAGTTTTAG
- TTCTTAGCAGAGTAGTTTTCAAATATGATTCTTATGATAAATGTAGACACAAACTATTTGAGA AACATTTAGAACTCTTAGCTTATACATTCAAAATGTAACTATTAAATGTGAAGATTTGGGGACAAAAT GTGAGTCAGA
- CACTGAAGAGTTTTTTGTTTTGATATTTTTTGATATTCTCTTTGCATTGAAATGGTATAAATGA
 ATCCATTTAA
- AAAGTGGTTAAGGATTTGTTTAGCTGGTGTGATAATAATTTTTTAAAGTTGCACATTGCCCAAG GCTTTTTTTGTGTGTTTTTATTGTTGTTACATTTGAAAAATATTCTTTGAATAACCTTGCAGTAC TATATTTCAA

FIGURE 1

TTTCTTTATAAATTTAAGTGCATTTTAACTCATAATTGTACACTATAATATAAGCCTAAGTTTTTATTCATAAGTTTT

ATTGAAGTTCTGATCGGTCCCCTTCAGAAATTTTTTTATATTATTCTTCAAGTTACTTTCTTA TTTATATTGTATGTGCATTTATCCATTAATGTTTCATACTTTCTGAGAGTATAATACCCTTTTAAAA GATATTTGGT

ATACCAATACTTTTĆCTGGATTGAAAACTTTTTTTAAACTTTTTAAAATTTGGGCCACTCTGTATGCA TATGTTTGGT

CTTGTTAAAGAGGAAGAAGGATGTGTTATACTGTACCTGTGAATGTTGATACAGTTACAA TTTATTTGACAAGGTTGTAATTCTAGAATATGCTTAATAAAATGAAAACTGGCCATGACTACAGCCAG AACTGTTATG

AGATTAACATTTCTATTGAGAAGCTTTTGAGTAAAGTACTGTATTTGTTCATGAAGATGACTGAGATGGTAACACTTC

GTGTAGCTTAAGGAAATGGGCAGAATTTCGTAAATGCTGTTGTGCAGATGTTGTTTCCCTGAA TGCTTTCGTATTAGTGGCGACCAGTTTCTCACAGAATTGTGAAGCCTGAAGGCCAAGAGGAAGTCACT GTTAAAGGAC

TCTGTGCCATCTTACAACCTTGGATGAATTATCCTGCCAACGTGAAAACCTCATGTTCAAAGAACACT

CGATGTAACTGCTGGTTTTGTTTTTCATATGTGTTTTTCTTACACTCATTTGAATGCTTTCAA GCATTTGTAAACTTAAAAAAAANWAWAAAGGGCAAAAAGTCTGAACCCTTGTTTTCTGAAATCTAATC AGTTATGTAT

GGTTTCTGAAGGGTAATTTTATTTTGGAATAGGTAAAGCGAAACCTGTTTTGTCWTGTTTTTCCTGAG GGCTAGATGC

ATTTTTTTCTCACACTCTTAATGACTTTTAACATTTATACTGAGCATCCATAGATATATTCC
TAGAAGTATGAGAAGAATTATTCTTATTGACCATTAATGTCATGTTCATTTTAATGTAATATATTGA
GATGAAATGT

TCTCTGGTTGGAACAGATACTCTCTTTTTTTTTTTTTTCTTGCAATCTTTAAGAATACATAGATCTAAAAATTC ATTAGCTTGA

CCCCTCAAAGTAACTTTTAAGTAAAGATTAAAGCTTTTCTCTCAGTGAATATATCTGCTAGA AGGAAATAGCTGGGAAGAATTTAATGATCAGGGAAATTCATTATTTCTATATGTGGAAACTTTTTGCT TCGAATATTG

TATCTTTTAAATCTAAATGTTCATATTTTTCCTGAAGAAACCACTGTGTAAAAATCAAATTTTAATT TTGAATGGAA

TAATTTCAAAGAACTATGAAGATGATTTGAAGCTCTAATTTATATAGTCACCTATAAAATGTT
CTTTATATGTGTTCATAAGTAAATTTTATATTGATTAAGTTAAACTTTTGAATTGAATTGAGGAGCAG
TAAAATGAAA

GCTATATCTATINCTAAACCYTATTTAGACATTGGKACCAGTTACCCAGGTGAAAATAKGGAGTAACT TTGTTTTGTA

TGGTAAGGTTTAGGAATGGNGGATGAAGGGTATCTCTATATAAATAAAGTGCTCAACAATGTG CAATGATTGTAAATTTAGTAAGATATTACAGCCATTTCATGAATGCTTTACCATTCAACATAGTATCT ATTACAAAAC

CATTAAATGTTTATTCTTTAAAATGAATGTATTATGTTTTTAACCCACAAATGCATACTTACC CTGTGCCTCATATTTCAATAGTACTGTAATATGGACATCTTTTGTGAAATACTTTTATTTTGTTATGC TTTAAATATA

CATACAAAAGATTTCTGTTATTAGCTTTGAAAATTGTATAATATCCTAATATAAACAAAAATATAAA AATAAAAATG

FIGURE 1 (cont'd)

 ${\tt MDEQSQGMQGPPVPQFQPQKALRPDMGYNTLANFRIEKKIGRGQFSEVYRAACLLDGVPVALKKVQIFDLMDAKARAD}$

CIKEIDLLKQLNHPNVIKYYASFIEDNELNIVLELADAGDLSRMIKHFKKQKRLIPERTVWKYFVQLC SALEHMHSRR

VMHRDIKPANVFITÄTGVVKLGDLGLGRFFSSKTTAAHSLVGTPYYMSPERIHENGYNFKSDI WSLGCLLYEMAALQSPFYGDKMNLYSLCKKIEQCDYPPLPSDHYSEELRQLVNMCINPDPEKRPDVTY VYDVAKRMHA CTASS

FIGURE 1 (cont'd)

4/18 GTCGACCCACGCGTCCGGTGGAAGTATAATACTTTGTCATTATGAGATGTCGTCTCTCGG TGCCTCCTTTGTGCAAATTAAATTTGATGACTTGCAGTTTTTTGAAAACTGCGGTGGAGG **AAGTTTTGGGAGTGTTTATCGAGCCAAATGGATATCACAGGACAAGGAGGTGGCTGTAAA** GAAGCTCCTCAAAATAGAGAAAGAGGCAGAAATACTCAGTGTCCTCAGTCACAGAAACAT CATCCAGTTTTATGGAGTAATTCTTGAACCTCCCAACTATGGCATTGTCACAGAATATGC TTCTCTGGGATCACTCTATGATTACATTAACAGTAACAGAAGTGAGGAGATGGATATGGA TCACATTATGACCTGGGCCACTGATGTAGCCAAAGGAATGCATTATTTACATATGGAGGC TCCTGTCAAGGTGATTCACAGAGACCTCAAGTCAAGAAACGTTGTTATAGCTGCTGATGG AGTACTGAAGATCTGTGACTTTGGTGCCTCTCGGTTCCATAA CCATACAACACACATGTC CTTGGTTGGAACTTTCCCATGGATGGCTCCAGAAGTTATCCAGAGTCTCCCTGTGTCAGA **AACTTGTGACACATATTCCTATGGTGTGGTTCTCTGGGAGATGCTAACAAGGGAGGTCCC** CTTTAAAGGTTTGGAAGGATTACAAGTAGCTTGGCTTGTAGTGGAAAAAAACGAGAGATT AACCATTCCAAGCAGTTGCCCCAGAAGTTTTGCTGAACTGTTACATCAGTGTTGGGAAGC TGATGCCAAGAAACGGCCATCATTCAAGCAAATCATTTCAATCCTGGAGTCCATGTCAAA TGACACGAGCCTTCCTGACAAGTGTAACTCATTCCTACACAACAAGGCGGAGTGGAGGTG CGAAATTGAGGCAACTCTTGAGAGGCTAAAGAAACTAGAGCGTGATCTCAGCTTTAAGGA GCAGGAGCTTAAAGAACGAGAAAGACGTTTAAAGATGTGGGAGCAAAAGCTGACAGAGCA GTCCAACACCCGCTTCTCTTGCCTCTTGCTGCAAGAATGTCTGAGGAGTCTTACTTTGA **ATCTAAAACAGAGGAGTCAAACAGTGCAGAGATGTCATGTCAGATCACAGCAACAAGTAA** CGGGGAGGCCATGCATGAACCCAAGTCTGCAGGCCATGATGCTGATGGGCTTTGGGGA TATCTTCTCAATGAACAAGCAGGAGCTGTGATGCATTCTGGGATGCAGATAAACATGCA AGCCAAGCAGAATTCTTCCAAAACCACATCTAAGAGAAGGGGGAAGAAAGTCAACATGGC TCTGGGGTTCAGTGATTTTGACTTGTCAGAAGGTGACGATGATGATGATGATGACGGTGA GGAGGAGGATAATGACATGGATAATAGTGAATGAAAGCAGAAAGCAAAGTAATAAAATCA CAAATGTTTGGAAAACACAAAAGTAACTTGTTTATCTCAGTCTGTACAAAAACAGTAAGG AGGCAGAAAGCCAAGCACTGCATTTTTAGGCCAATCACATTTACATGACCGTAATTTCTT ATCAATTCTACTTTTATTTTTGCTTACAGAAAAACGGGGGGAGAATTAAGCCAAAGAAGT **ATATTTATGAATCAGCAAATGTGGTGCCTGATTATAGAAATTTGTGATCCTATATACAAT** TAGCAATACACTGTAATATCAGAAATGGTTGGCCTGAGCAACCTAGTAAGACCTCGTCTC TACTAATAATTAAAAAACTAGCTGGCATGGTAGCACACCCTGTAGTCCCAGATACTTGG GATGCACCACTGCACTCTAGCCTGGGCAAGAGAACAAGATCCTGTCTCAAAAAAACAAAA AAAAAAAAAAGGGCGGCCG

MSSLGASFYQIKPOOLOFFERCOGGSFGSYTRAKHISODKSYAVK
KLLKIEKEAEILSVLSHRNIIQFTGVILEPPNYGIVTEYASLGSLYDTIMSRESENDMO
HUMTMATDVAKGRHYLINEAPVKVIHROLKSRNVVIAADGVLKICOFGASRFFRHETTEMS
LVGTFMGAJFVIQSLJVSETCDTTSTGVVLMEHLARSVFFKILLSILGVAMLAVEKNERL
TTPSSCFRSFAELLHQCMEADAKKRPSFKQIEILESMSKOTSLFDECNSFLENKAENGC
EIEATLERLAKLERDLSFKPQELEERERLKMMPQKLITEQSNTFILLFLAKMSEESTFE
SKTEESNSAEMSCQITATSNGEGHGRNFSLQANNIMGFGDIFSNKKAGVKHSGRQINRQ
AKQNSSKTTSKRGKKVNGALGFSDPDLSEGDDDDDDDGREEDNDMSSE

CGGTGGTGGCGGCAGCGGCGGCTGCGGGGCACCATGGCCGTGC GACAGGCGCTGGGCCGCGCCTGCAGCTGGGTCGAGCGCTGCTGCTGCGCTTCACGGGCA AGCGTCCAGGCTGGGCCGCAGGACCGGGCGCGGGGCCTCGCAGGGTCGGGCTTC CTAACCGTOTCCGCCTTCTTCCGCCAGTCGGTGGCCGGGCTGGCGGCGGGTTGCAGCGGC AGTTCGTGGTGCGGGCCTGGGGCTGCGCGGGCCCTTGCGGCCGGGCAGTCTTTCTGGCCT TCGGGCTAGGGCTCGTCATCGAGGAAAAACAGGCGGAGAGCCGGCGGCGGTCTCGG **ACACGAGACGCTTGCAGGGCTTTCGGCTGGAGGAGTATCTGATAGGGCAGTCCATTGGTA** AGGGCTGCAGTGCTGTGTATGAAGCCACCATGCCTACATTGCCCCAGAACCTGGAGG TGACAAAGAGCACCGGGTTGCTTCCAGGGAGAGGCCCAGGTACCAGTGCACCAGGAGAAG GGCAGGAGCGAGCTCCGGGGGCCCCTGCCTTCCCCTTGGCCATCAAGATGATGTGGAACA TCTCGGCAGGTTCCTCCAGCGAAGCCATCTTGAACACAATGAGCCAGGAGCTGGTCCCAG CGAGCCGAGTGGCCTTGGCTGGGGAGTATGGAGCAGTCACTTACAGAAAATCCAAGAGAG GTCCCAAGCAACTAGCCCCTCACCCCAACATCATCCGGGTTCTCCGCGCCTTCACCTCTT CCGTGCCGCTGCCAGGGGCCCTGGTCGACTACCCTGATGTGCTGCCCTCACGCCTCC ACCCTGAAGGCCTGGGCCATGGCCGGACGCTGTTCCTCGTTATGAAGAACTATCCCTGTA CCCTGCGCCAGTACCTTTGTGTGAACACACCCAGCCCCCGCCTCGCCGCCATGATGCTGC TGCAGCTGCTGGAAGGCGTGGACCATCTGGTTCAACAGGGCATCGCGCACAGAGACCTGA ATTTTGGCTGCTGCTGATGAGAGCATCGGCCTGCAGTTGCCCTTCAGCAGCTGGT ACGTGGATCGGGCGGAAACGGCTGTCTGATGGCCCCAGAGGTGTCCACGGCCCGTCCTG GCCCCAGGCAGTGATTGACTACAGCAAGGCTGATGCCTGGGCAGTGGGAGCCATCGCCT ATGAAATCTTCGGGCTTGTCAATCCCTTCTACGGCCAGGGCAAGGCCCACCTTGAAAGCC GCAGCTACCAAGAGGCTCAGCTGCCTGCACTGCCCGAGTCAGTGCCTCCAGACGTGAGAC AGTTGGTGAGGGCACTGCTCCAGCGAGAGGCCAGCAAGAGACCATCTGCCCGAGTAGCCG CAAATGTGCTTCATCTAAGCCTCTGGGGTGAACATATTCTAGCCCTGAAGAATCTGAAGT TAGACAAGATGGTTGGCTGGCTCCCAACAATCGGCCGCCACTTTGTTGGCCAACAGGC TCACAGAGAAGTGTTGTGTGGAAAAAAAAGATGCTCTTTCTGGCTAACCTGGAGT GTGAAACGCTCTGCCAGGCAGCCCTCCTCTCTCTCTCATGGAGGGCAGCCCTGTGATGTC CCTGCATGGAGCTGGTGAATTACTAAAAGAACTTGGCATCCTCTGTGTCGTGATGGTCTG AAGGCCTCGGGCTTGGCAAATGGAAGAACTTGAGTGAGAGTTCAGTCTGCAGTCCTCTGC GGGTAGGCCTGCATCCACAGAGAGGGATCCAGGCCAAGGCACTGGCTGTCAGTGGCAGAG TTTGGCTGTGACCTTTGCCCCTAACACGAGGAACTCGTTTGAAGGGGGCAGCGTAGCATG TCTGATTTGCCACCTGGATGAAGGCAGACATCAACATGGGTCAGCACGTTCAGTTACGGG AGTGGGAAATTACATGAGGCCTGGGCCTCTGCGTTCCCAAGCTGTGCGTTCTGGACCAGC GGGATTTAAACTTGAGGGTTTCCCTCCTGACTAGCCTCTCTTACAGGAATTGTGAAATAT

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Met Ala

Val Arg Gln Ala Leu Gly Arg Gly Leu Gln Leu Gly Arg Ala Leu Leu
5 10 15

Leu Arg Phe Thr Gly Lys Pro Gly Arg Ala Tyr Gly Leu Gly Arg Pro 20 25 30

Gly Pro Ala Ala Gly Cys Val Arg Gly Glu Arg Pro Gly Trp Ala Ala 35 40 45 50

Gly Pro Gly Ala Glu Pro Arg Arg Val Gly Leu Gly Leu Pro Asn Arg
55 60 65

Leu Arg Phe Phe Arg Gln Ser Val Ala Gly Leu Ala Ala Arg Leu Gln 70 75 80

Arg Gln Phe Val Val Arg Ala Trp Gly Cys Ala Gly Pro Cys Gly Arg 85 90 95

Ala Val Phe Leu Ala Phe Gly Leu Gly Leu Gly Leu Ile Glu Glu Lys 100 105 110

Gln Ala Glu Ser Arg Arg Ala Val Ser Ala Cys Gln Glu Ile Gln Ala 115 120 125 130

Ile Phe Thr Gln Lys Ser Lys Pro Gly Pro Asp Pro Leu Asp Thr Arg
135 140 145

Arg Leu Gln Gly Phe Arg Leu Glu Glu Tyr Leu Ile Gly Gln Ser Ile 150 155 160

Gly Lys Gly Cys Ser Ala Ala Val Tyr Glu Ala Thr Met Pro Thr Leu 165 170 175

Pro Gln Asn Leu Glu Val Thr Lys Ser Thr Gly Leu Leu Pro Gly Arg 180 185 190

Gly Pro Gly Thr Ser Ala Pro Gly Glu Gly Gln Glu Arg Ala Pro Gly 195 200 205 210

Ala Pro Ala Phe Pro Leu Ala Ile Lys Met Met Trp Asn Ile Ser Ala 215 220 225

FIGURE 3 (cont'd)

Gly Ser Ser Ser Glu Ala Ile Leu Asn Thr Met Ser Gln Glu Leu Val 230 235 240

Pro Ala Ser Arg Val Ala Leu Ala Gly Glu Tyr Gly Ala Val Thr Tyr 245 250 255

Arg Lys Ser Lys Arg Gly Pro Lys Gln Leu Ala Pro His Pro Asn Ile 260 265 270

Ile Arg Val Leu Arg Ala Phe Thr Ser Ser Val Pro Leu Leu Pro Gly 275 280 285 295

Ala Leu Val Asp Tyr Pro Asp Val Leu Pro Ser Arg Leu His Pro Glu 295 300 305

Gly Leu Gly His Gly Arg Thr Leu Phe Leu Val Met Lys Asn Tyr Pro 310 315 320

Cys Thr Leu Arg Gln Tyr Leu Cys Val Asn Thr Pro Ser Pro Arg Leu 325 330 335

Ala Ala Met Met Leu Leu Gln Leu Leu Glu Gly Val Asp His Leu Val 340 345 350

Gln Gln Gly Ile Ala His Arg Asp Leu Lys Ser Asp Asn Ile Leu Val 355 360 365 370

Glu Leu Asp Pro Asp Gly Cys Pro Trp Leu Val Ile Ala Asp Phe Gly 375 380 385

Cys Cys Leu Ala Asp Glu Ser Ile Gly Leu Gln Leu Pro Phe Ser Ser 390 395 400

Trp Tyr Val Asp Arg Gly Gly Asn Gly Cys Leu Met Ala Pro Glu Val 405 410 415

Ser Thr Ala Arg Pro Gly Pro Arg Ala Val Ile Asp Tyr Ser Lys Ala 420 425 430

FIGURE 3 (cont'd)

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Asp Ala Trp Ala Val Gly Ala Ile Ala Tyr Glu Ile Phe Gly Leu Val
435 440 445 456

Asn Pro Phe Tyr Gly Gln Gly Lys Ala His Leu Glu Ser Arg Ser Tyr.
455 460 465

Gln Glu Ala Gln Leu Pro Ala Leu Pro Glu Ser Val Pro Pro Asp Val
470 475 480

Arg Gln Leu Val Arg Ala Leu Leu Gln Arg Glu Ala Ser Lys Arg Pro 485 490 495

Ser Ala Arg Val Ala Ala Asn Val Leu His Leu Ser Leu Trp Gly Glu 500 505 510

His Ile Leu Ala Leu Lys Asn Leu Lys Leu Asp Lys Met Val Gly Trp 515 520 530

Leu Leu Gln Gln Ser Ala Ala Thr Leu Leu Ala Asn Arg Leu Thr Glu
535 540 545

Lys Cys Cys Val Glu Thr Lys Met Lys Met Leu Phe Leu Ala Asn Leu 550 555 560

Glu Cys Glu Thr Leu Cys Gln Ala Ala Leu Leu Leu Cys Ser Trp Arg 565 570 575

Ala Ala Leu

GTCGACCCACGCGTCCGCCCACGCGTTCCGGAGACATGTCTCTGTGTTTC TCTCCCCTCCGCTTTTGAGTCCGTTGAAGACACAATTTCTCTCTGTCGGGT GCTTAGGAGGAGCTCCATGAACATGTATTGAATTGGACTTAGCTGAACAG GCTGCTGGTTGGCTGCCCAGAGGGGGCAGGCTGTGTTGCTGGGAGCCTTC CAGCTCCCTGCAGCAGTCATGGGGCAGGGTTCCCCGAGTCCGTAATCCCC TTAAGTGGAGCATCCCCTTTCCTGGGAGACACGAAGCAGGAAACACTGGC **AAATATCACAGCAGTGAGTTACGACTTTGATGAGGAATTCTTCAGCCAGA** CGAGCGAGCTGGCCAAGGACTTTATTCGGAAGCTTCTGGTTAAAGAGACC CGGAAACGCTCACAATCCAAGAGGCTCTCAGACACCCCTGGATCACGCC GGTGGACAACCAGCAAGCCATGGTGCGCAGGGAGTCTGTGGTCAATCTGG AGAACTTCAGGAAGCAGTATGTCCGCAGGCGGTGGAAGCTTTCCTTCAGC ATCGTGTCCCTGTGCAACCACCTCACCGCTCGCTGATGAAGAAGGTGCA CCTGAGGCCGGATGAGGACCTGAGGAACTGTGAGAGTGACACTGAGGAG GACATCGCCAGGAGGAAAGCCCTCCACCCACGGAGGAGGAGCAGCACCT CCTAACTGGCCTGACCTGCAGTGGCCGCCAGGGAGGTCTGGGCCCAGCGG GGCTCCCTTCTGTGCAGACTTTTGGACCCAGCTCAGCACCAGCACCCGGGC GTCCTGAGCACTTTGCAAGAGAGATGGGCCCAAGGAATTCAGAAGAGCTT GCAGGCAAGCCAGGAGACCCTGGGAGCTGTGGCTGTCTTCTGTGGAGGAG GCTCCAGCATTCCCAAAGCTCTTAATTCTCCATAAAATGGGCTTTCCTCTG TCTGCCATCCTCAGAGTCTGGGGTGGGAGTGTGGACTTAGGAAAACAATA TAAAGGACATCCTCATCATCACGGGGTGAAGGTCAGACTAAGGCAGCCTT CTTCACAGGCTGAGGGGGTTCAGAACCAGCCTGGCCAAAAATTACACCAG AGAGACAGAGTCCTCCCCATTGGGAACAGGGTGATTGAGGAAAGTGAACC TTGGGTGTGAGGGACCAATCCTGTGACCTCCCAGAACCATGGAAGCCAGG **ACGTCAGGCTGACCAACACCTCAGACCTTCTGAAGCAGCCCATTGCTGGC** CCGCCATGTTGTAATTTTGCTCATTTTTATTAAACTTCTGGTTTACCTGATG CTTGGCTTCTTTTAGGGCTACCCCCATCTCATTTCCTTTAGCCCGTGTGCCT GTAACTCTGAGGGGGGGCACCCAGTGGGGTGCTGAGTGGGCAGAATCTCA CTGGGAGTGGTGGCTCACGCCTATAATCCCTGCGCTTTGGCAGTCCGAGG CAGGGGAATTGCTTGAACTCAGGAGTTGGAGACCAGCCTGGGCAACATGG CAAAACGCAGTCTGTACAAAAAATACAAAAAATTAGCCAGGTGTAGGGGT AGGCACCTGGCATCCCAGCTACTCCAGGGGCTGAGGTGACAGCATTGCTT **AAGCCAGAAGGTCGAGGCTGCAGTGAGCTGAGATCACGCCACTGCACTC** CGGCCGC

LFDSLSVVLSLSGASPFLGDTKQETLANITAVSYDFDEEFFSQTSELAKDFIRKL LVKETRKRLTIQEALRHPWITPVDNQQAMVRRESVVNLENFRKQYVRRRWK LSFSIVSLCNHLTRSLMKKVHLRPDEDLRNCESDTEEDIARRKALHPRRRSSTS

KELRELRI EET 19 G ACG OCA TEX COC-AAA GAA CEA AGA GAA CEC COG AST GAA GAA ACA AAC COC CCA AEG K K V T D Y S S S B B S B S S S ANG ANG GTG ACT GAT TAC TOO TOO TOO ACT GAG GAG TOA GAA ACT ACC GAG GAA GAG GAG H D G T V R $\mathbf{D} \cdot \mathbf{I}$ 59 GAA GAT GGA GAG AGC GAG-ACC CAT GAT GGG ACA GTG GCT GTC AGC GAC ATA CCC AGA CTC 177 I P T G A P G S N E Q Y N V G K V G 79 ATA COA ACA GGA GCT CCA GGC AGC AAC GRG CAG TAC AAT GTG GGA ATG GTG GGG ACG CAT 237 в н P S G D R R 99 OGG CTG GAG ACC TCT CAT GCG GAC ACT TTC AGC GCC AGT ATT TCA AGA GAA GGA ACC TTG 297 I R E T S G B K K R S G H S D 8 N Q 119 ATG ATT AGA GAG ACG TOT GGA GAG AAG AAG CGA TOT GGC CAC AGT GAC AGC AAT GGC TIT 357 V Q Q L H 139 COT COC CAC ATC AAC CTC CCT. GAC CTG GTG CAG CAG CAG CAT TCT CCA CCT CCA ACC CCC 417 TEGLGRVSTHSQEMDSG.T 159 NET CAG CEA CTG CCC CEC CEC TEA NEC CAT TEC CAG GRG ATG GAC TET CCC ACT GAA TAT SSTK SFTP? V D P R V 179 GOOD ARTO GOOD AGO AGO AGO AAA GOOD TOO TITO AGO COOD TITT GITG GAO COOD AGA GITA THE CAG 537 T D E R E 8 S מ E D R 199 AGG TOT COO ACT GAT GAA GAT GAA GAG GAT GAG GAA TOA TOA GOO GOA GOT CTG TTT ACT 597 L R Q B Q λ K L N B R ĸ T λ 219 NGC GRA CTT CTT NGG CRA GRA CNG GCC NAA CTC NAT GRA GCA NGA NAG ATT TGG GTG GTA 657 N I ₽ H 8 D T P BI R ĸ 239 ART GEN AND DEA AND ART COG DOT ORT NOO DRO ACA DOA GRA ATC AGA ARA TRO AND 717 K R P N S E I L C A A L W G V N L L 259 MAA OGA TIC AAC TCA GAA ATA CIT TOT OCA OCT CIG TOG GGT GTA AAC CIT CIG GIG GGG 777 E N G L M L L D R E G Q G K v 279 ACT GAA AAT GGC CTG ATG CTT. TTG GAC GGA AGT GGG CAA GGC AAA GTC TAT AAT CTG ATC 637 P Q ONDVLE G 299 ANC COO AGO COA TIT CHG CAGATO CAT COO CIA CHG CGA CTG AAT GTC CIT GTC ACA ACT 897 319 TCA GGA ANG ANG ANT ANG CTA COR GTT THE THE CTT TCA TGG TTA AGA ANG AGA ATA CTA 957 H N D P E V E K R Q G W I T V G D L CAT ANT CAC CCA CAA GTA CAA:AAG AAA CAA COC TOO ATC ACT OTT COO GAC TTO CAA CCC 1017 VKYBRIKF CIHYKV L V I 359 TOT ATA CHE TAT ANA GIT GIT! ANA THE GAA AGO AIC ANA TIT TIG GIV AT GOO TIA ANG 1077 e i Y À W ĸ H A P Y ĸ P 379 ANT OCT GTG GAA ATA ERT OCT TOG GCT GCT MAA CCG TAT CAT AAA TTC ATG GCA TIT ANG 1137 PADLQHKPLLV D LTV E 2 399 TET TIT GEA GAT CIC CAG CAC ANG CET CES CEA OTT GAT CTC ACG GTA GAA GAA GGT CAA 1197 H V a D G 419

FIGURE 5

11/18

AGA	TTA	MG	GTT	ATT	TTT	CCT	TCA	CAC	ACT	OCT	TIC	CAT	QEY	ATT	GAT	GTT	GAT	TCA	GGA	1257
N N	s TCT	Y TAT	D CAT	I ATC	Y TAC	I ATA	œ X	S TCT	H CAT	I ATT	care ő	G GGC	n aat	I ATC	T ACT	P CCT	H CAT	A GCT	I Att	439 1317
	I ATC																	,		444 1332

FIGURE 5 (cont'd)

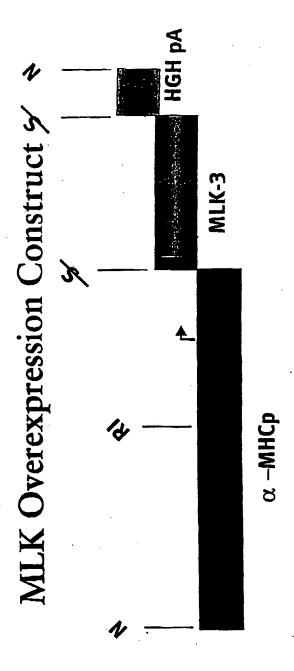
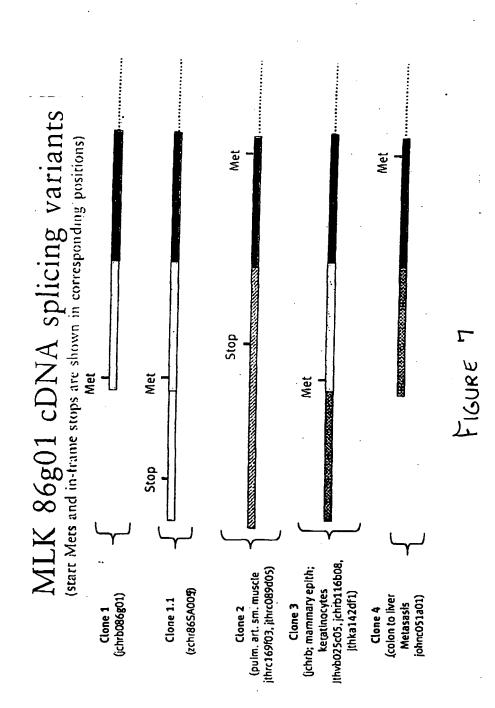
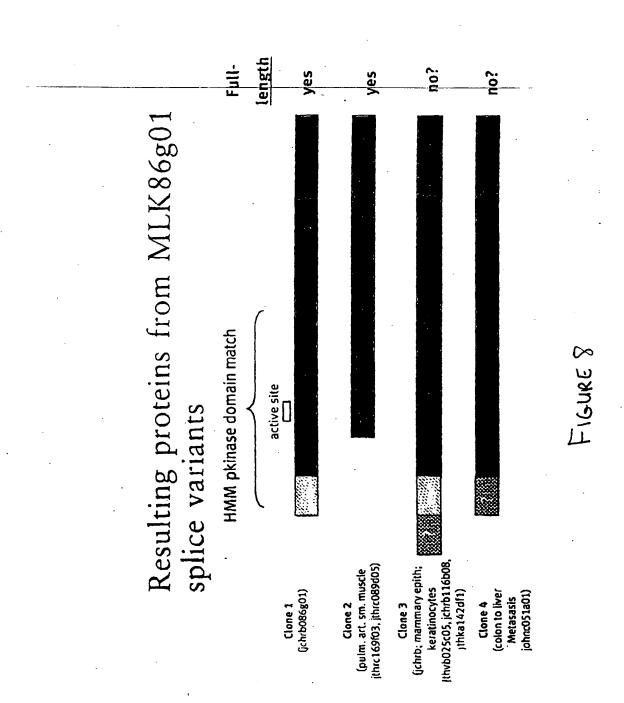


Figure 6



BNSDOCID: <WO__0014212A1_J_>

14/18



BNSDOCID: <WO__0014212A1_I_>

15/18

Clone 1.1 (includes parts of zchr86SA004, jchrb086g01)

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16/18
Clone 2 (includes parts of jthrc169f03, jthrc089d05, jchrb086g01)

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17/18

Clone 3 (includes parts of jthvb025c05, jchrb116b08, jthka142df1, jchrb086g01)

CGCGTCCGCCACGCGTCCGGCAGAGAGGCGGAATGTTCAACTCCTAACT GCGGCGAAACGTGGGAGCCGCGCGGGCCGCTGTCGTCCCAACCCCCGCC GCCCGGGAGCCAGATTTTGTGGAAGTATAATACTTTGTCATTATGAGATGT CGTCTCTCGGTGCCTCCTTTGTGCAAATTAAATTTGATGACTTGCAGTTTTT TGAAAACTGCGGTGGAGGAAGTTTTGGGAGTGTTTATCGAGCCAAATGGA TATCACAGGACAAGGAGGTGGCTGTAAAGAAGCTCCTCAAAATAGAGAA AGAGGCAGAAATACTCAGTGTCCTCAGTCACAGAAACATCATCCAGTTTT ATGGAGTAATTCTTGAACCTCCCAACTATGGCATTGTCACAGAATATGCTT CTCTGGGATCACTCTATGATTACATTAACAGTAACMGAAKTGAGGAGATG GATATGGATCACATTATGACCTGGGCCACTGATGTAGCCAAAGGAATGCA TTATTTACATATGGAGGCTCCTGTCAAGGTGATTCACAGAGACCTCAAGTC AAGAAACGTTGTTATAGCTGCTGATGGAGTACTGAAGATCTGTGACTTTG TCCCATGGATGGCTCCAGAAGTTATCCAGAGTCTCCCTGTGTCAGAAACTT GTGACACATATTCCTATGGTGTGGTTCTCTGGGAGATGCTAACAAGGGAG GTCCCCTTTAAAGGTTTGGAAGGATTACAAGTAGCTTGGCTTGTAGTGGA AAAAACGAGAGATTAACCATTCCAAGCAGTTGCCCCAGAAGTTTTGCTG AACTGTTACATCAGTGTTGGGAAGCTGATGCCAAGAAACGGCCATCATTC AAGCAAATCATTTCAATCCTGGAGTCCATGTCAAATGACACGAGCCTTCCT GACAAGTGTAACTCATTCCTACACAACAAGGCGGAGTGGAGGTGCGAAAT TGAGGCAACTCTTGAGAGGCTAAAGAAACTAGAGCGTGATCTCAGCTTTA AGGAGCAGGAGCTTAAAGAACGAGAAAGACGTTTAAAGATGTGGGAGCA AAAGCTGACAGAGCAGTCCAACACCCGCTTCTCTTGCCTCTTGCTGCAAG **AATGTCTGAGGAGTCTTACTTTGAATCTAAAACAGAGGAGTCAAACAGTG** CAGAGATGTCATGTCAGATCACAGCAACAAGTAACGGGGAGGGCCATGG CATGAACCCAAGTCTGCAGGCCATGATGCTGATGGGCTTTGGGGATATCT TCTCAATGAACAAAGCAGGAGCTGTGATGCATTCTGGGATGCAGATAAAC ATGCAAGCCAAGCAGAATTCTTCCAAAACCACATCTAAGAGAAGGGGGA AGAAAGTCAACATGGCTCTGGGGTTCAGTGATTTTGACTTGTCAGAAGGT GACGATGATGATGATGACGGTGAGGAGGAGGATAATGACATGGATA ATAGTGAATGAAAGCAGAAGCAAAGTAATAAAATCACAAATGTTTGGA AAACACAAAAGTAACTTGTTTATCTCAGTCTGTACAAAAACAGTAAGGAG GCAGAAAGCCAAGCACTGCATTTTTAGGCCAATCACATTTACATGACCGT AATTTCTTATCAATTCTACTTTTATTTTTGCTTACAGAAAAACGGGGGGAG AATTAAGCCAAAGAAGTATATTTATGAATCAGCAAATGTGGTGCCTGATT ATAGAAATTTGTGATCCTATATACAATATAGGACTTTTAAAGTTGTGACAT TCTGGCTTTTTCTTTTAATGAATACTTTTTAGTTTGTATTTGACTTTATTTCC TAGCAATACACTGTAATATCAGAAATGGTTGGCCTGAGCAACCTAGTAAG ACCTCGTCTCTACTAATAATTAAAAAAACTAGCTGGCATGGTAGCACACAC CTGTAGTCCCAGATACTTGGGAGGCCAAGGCAGGAGGATTGCTTGAGACC TAGCAATCAGTCAGGCTGCAGTGAGCCATGATGGCACCACTGCACTCTA AAGGGCGGCCGC

18/18

Clone 4 (includes parts of johnc051a01 and jchrb086g01)

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SEQUENCE LISTING

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gaa Glu	ata Ile	gat Asp	ctt Leu 85	ctt Leu	aag Lys	caa Gln	ctc Leu	aac Asn 90	cat His	cca Pro	aat Asn	gta Val	ata Ile 95	aaa Lys	tat Tyr	587
tat Tyr	gca Ala	tca Ser 100	ttc Phe	att Ile	gaa Glu	gat Asp	aat Asn 105	gaa Glu	cta Leu	aac Asn	ata Ile	gtt Val 110	ttg Leu	gaa Glu	cta Leu	635
gca Ala	gat Asp 115	gct Ala	ggc Gly	gac Asp	cta Leu	tcc Ser 120	aga Arg	atg Met	atc Ile	aag Lys	cat His 125	ttt Phe	aag Lys	aag Lys	caa Gln	683
aag Lys 130	agg Arg	cta Leu	att Ile	cct Pro	gaa Glu 135	aga Arg	act Thr	gtt Val	tgg Trp	aag Lys 140	tat Tyr	ttt Phe	gtt Val	cag Gln	ctt Leu 145	731
tgc Cys	agt Ser	gca Ala	ttg Leu	gaa Glu 150	cac His	atg Met	cat His	tct Ser	cga Arg 155	aga Arg	gt <i>c</i> Val	atg Met	cat His	aga Arg 160	gat Asp	779
ata Ile	aaa Lys	cca Pro	gct Ala 165	aat Asn	gtg Val	ttc Phe	att	aca Thr 170	gcc Ala	act Thr	ggg Gly	gtg Val	gta Val 175	aaa Lys	ctt Leu	827
gga Gly	gat Asp	ctt Leu 180	ggg Gly	ctt Leu	ggc Gly	cgg Arg	ttt Phe 185	ttc Phe	agc Ser	tca Ser	aaa Lys	acc Thr 190	aca Thr	gct Ala	gca Ala	875
cat His	tct Ser 195	tta Leu	gtt Val	ggt Gly	acg Thr	cct Pro 200	tat Tyr	tac Tyr	atg Met	tct Ser	cca Pro 205	gag Glu	aga Arg	ata Ile	cat His	923
gaa Glu 210	aat Asn	gga Gly	tac Tyr	aac Asn	ttc Phe 215	aaa Lys	tct Ser	gac Asp	atc Ile	tgg Trp 220	tct Ser	ctt Leu	ggc Gly	tġt Cys	cta Leu 225	971
cta Leu	tat Tyr	gag Glu	atg Met	gct Ala 230	gca Ala	tta Leu	caa Gln	agt Ser	cct Pro 235	ttc Phe	tat Tyr	ggt Gly	gac Asp	aaa Lys 240	atg Met	1019
aat Asn	tta Leu	tac Tyr	tca Ser 245	ctg Leu	tgt Cys	aag Lys	aag Ly s	ata Ile 250	gaa Glu	cag Ģln	tgt Cys	gac Asp	tac Tyr 255	cca Pro	cct Pro	1067
ctt Leu	cct Pro	tca Ser 260	gat Asp	cac His	tat Tyr	tca Ser	gaa Glu 265	g aa Glu	ctc Leu	cga Arg	cag Gln	tta Leu 270	gtt Val	aat Asn	atg Met	1115
tgc Cys	atc Ile 275	aac Asn	cca Pro	gat Asp	cca Pro	gag Glu 280	aag Lys	cga Arg	cca Pro	gac Asp	gtc Val 285	acc Thr	tat Tyr	gtt Val	tat Tyr	1163
gac	gta	gca	aag	agg	atg	cat	gca	tgc	act	gca	agc	agc	taaa	cato	ıca	1212

-3-

Asp Val Ala Lys Arg Met His Ala Cys Thr Ala Ser Ser 290 295 300

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- 5 -

PCT/US99/20631

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		tct Ser 195													ata Ile.	624
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-7-

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	aat Asn															768
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gtc		gcc	tcc	ttt	gtg	caa	att	aaa	ttt	gat	gac	ttg	let S 1 cag	Ser S	er ttt	55 103
ctc Leu	gacco ggt Gly	gcc Ala tgc	tcc Ser	ttt Phe gga	gt <i>g</i> Val gga	caa Gln 10 agt	att Ile ttt	aaa Lys ggg	ttt Phe agt	gat Asp gtt	gac Asp 15	ttg Leu cga	cag Gln	ttt Phe	ttt Phe tgg	ì
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	atg Met															1255
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	aaa Lys															1351
	gac Asp														atg Met	1399
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- 11 -

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Glu 305	Gln	Glu	Leu	Lys	Glu 310		Glu	Arg	Arg	Leu 315		Met	Trp	Glu	Gln 320	
Lys	Leu	Thr	Glu	Gln 325		Asn	Thr	Pro	Leu 330	Leu	Leu	Pro	Leu	Ala 335	Ala	
Arg	Met	Ser	Glu 340	Glu	Ser	Tyr	Phe	Glu 345	Ser	Lys	Thr	Glu	Glu 350	Ser	Asn	
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His	Gly 370	Met	Asn	Pro	Ser	Leu 375	Gln	Ala	Met	Met	Leu 380	Met	Gly	Phe	Gly	
Asp 385		Phe	Ser	Met	Asn 390	Lys	Ala	Gly	Ala	Va1 395	Met	His	Ser	Gly	Met 400	
Gln	Ile	Asn	Met	Gln 405	Ala	Lys	Gln	Asn	Ser 410	Ser	Lys	Thr	Thr	Ser 415	Lys	
Arg	Arg	Gly	Lys 420	Lys	Val	Asn	Met	Ala 425	Leu	Gly	Phe	Ser	Asp 430	Phe	Asp	
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- 12 -

		35					40					45				
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		_				Val			_		ccc Pro					240
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Leu	_				Val	-			-		gaa Glu 220				-	672
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- 13 -

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			Phe												gag Glu	864
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	•		ctt Leu		-	-	-	-			_	_				960
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_	_		gag Glu 340					_								1056
			atg Met													1104
		-	aac Asn		-	_	_	-	-	_	-	-				1152
			tca Ser													1200
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-			aag Lys 420		-		_	_	-			Ser	-		-	1296
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										cgt Arg 45						200
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	_					_			-	gcg Ala			-			344
_	-		-	-						ggc Gly				-		392
										tgt Cys 125						440
								Gly		gac Asp						488
										ctg Leu						536
	•		_	-	-	-			-	gcc Ala		_			_	584
	-		-				_	-		ggg Gly	_				_	632
										cag Gln 205						680
gcc	cct	gcc	ttc	ccc	ttg	gcc	atc	aag	atg	atg	tgg	aac	atc	tcg	gca	728

Ala	Pro	Ala	Phe	Pro 215	Leu	Ala	Ile	Lys	Met 220		Trp	Asn	Ile	Ser 225	Ala	
				Glu	gcc Ala											776
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- 17 -

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Gly	Arg	Ala	Val 100	Phe	Leu	Ala	Phe	Gly 105	Leu	Gly	Leu	Gly	Leu 110	Ile	G1
Glu	Lys	Gln 115	Ala	Glu	Ser	Arg	Arg 120	Ala	Val	Ser	Ala	Cys 125	Gln	Glu	I1
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Pro	Gly 210	Ala	Pro	Ala	Phe	Pro 215	Leu	Ala	Ile	Lys	Met 220	Met	Trp	Asn	Ile
Ser 225	Ala	Gly	Ser.	Ser	Ser 230	Glu	Ala	Ile	Leu	Asn 235	Thr	Met	Ser	Gln	Ğ11 240
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Thr	Tyr	Arg	Lys 260	Ser	Lys	Arg	Gly	Pro 265	Lys	Gln	Leu	Ala	Pro 270	His	Pro

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Pro	Gly 290	Ala	Leu	Val	Asp	Tyr 295	Pro	Asp	Val	Leu	Pro 300	Ser	Arg	Leu	Hi
Pro 305	Glu	Gly	Leu	Gly	His 310	Gly	Arg	Thr	Leu	Phe 315	Leu	Val	Met	Lys	As:
Tyr	Pro	Cys	Thr	Leu 325	Arg	Gln	Tyr	Leu	Cys 330	Val	Asn	Thr	Pro	Ser 335	Pro
Arg	Leu	Ala	Ala 340	Met	Met	Leu	Leu	Gln 345	Leu	Leu	Glu	Gly	Val 350	Asp	His
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Phe 385	Gly	Cys	Cys	Leu	Ala 390	Asp	Glu	Ser	Ile	Gly 395	Leu	Gln	Leu	Pro	Phe 400
Ser	Ser	Trp	Tyr	Val 405	Asp	Arg	Gly	Gly	Asn 410	Gly	Cys	Leu	Met	Ala 415	Pro
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	450				-	455				Ala	460				
165	_		, ,		470					Pro 475					480
	-			485					490	Gln			٠	495	
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_	530					535				Leu	540				
545			_		550			ē		Lys 555					560
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ggg Gly	aga Arg	ggc Gly 195	cca Pro	ggt Gly	acc Thr	agt Ser	gca Ala 200	cca Pro	gga Gly	gaa Glu	ggg Gly	cag Gln 205	gag Glu	cga 'Arg	gct Ala	624
ccg Pro	ggg Gly 210	gcc Ala	cct Pro	gcc Ala	ttc Phe	ccc Pro 215	ttg Leu	gcc Ala	atc Ile	aag Lys	atg Met 220	atg Met	tgg Trp	aac Asn	atc Ile	672
tcg Ser 225	gca Ala	ggt Gly	tcc Ser	tcc Ser	agc Ser 230	gaa Glu	gcc Ala	atc Ile	ttg Leu	aac Asn 235	aca Thr	atg Met	agc Ser	cag Gln	gag Glu 240	720
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cct Pro 305	gaa Glu	ggc Gly	ctg Leu	ggc Gly	cat His 310	ggc Gly	cgg Arg	acg Thr	ctg Leu	ttc Phe 315	ctc Leu	gtt Val	atg Met	aag Lys	aac Asn 320	960
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cac His	ccc Pro	tgg Trp	atc Ile 75	acg Thr	ccg Pro	gtg Val	gac Asp	aac Asn 80	cag Gln	caa Gln	gcc Ala	atg Met	gtg Val 85	cgc Arg	agg Arg	535
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Lys Asp Phe Ile Arg Lys Leu Leu Val Lys Glu Thr Arg Lys Arg Leu 50 60

Thr Ile Gln Glu Ala Leu Arg His Pro Trp Ile Thr Pro Val Asp Asn 65 70 75 80

Gln Gln Ala Met Val Arg Arg Glu Ser Val Val Asn Leu Glu Asn Phe 85 90 95

Arg Lys Gln Tyr Val Arg Arg Arg Trp Lys Leu Ser Phe Ser Ile Val 100 105 110

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Ser	Leu	Cys 115		His	Leu	Thr	Arg 120		Leu	Met	Lys	Lys 125		His	Leu	
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					aag Lys											192
				_	ctc Leu 70	-					_	-		-		240
					cgc Arg											288
	-	_		_	cgc Arg				_				-			336
					ctc Leu											384
					ctg Leu											432
atc	gcc	agg	agg	aaa	gcc	ctc	cac	cca	cgg	agg	agg	agc	agc	acc	tcc	480

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	agt Ser															145
	aca Thr 50															193
	ggc Gly															241
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	acc Thr															337
	agt Ser															385
	cag Gln 130															433
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	Ilė									atc Ile 235						721
										ctg Leu						769
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										cga Arg						865
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Arg	Phe	Asn	Ser	Glu 245	Ile	Leu	Суз	Ala	Ala 250	Leu	Trp	Gly	Val	Asn 255	Leu
Leu	Val	Gly	Thr 260	Glu	Asn	Gly	Leu	Met 265	Leu	Leu	Asp	Arg	Ser 270	Gly	Gln
Gly	Lys	Val 275	Tyr	Asn	Leu	Ile	Asn 280	Arg	Àrg	Arg	Phe	Gln 285	Gln	Met	Asp
Val	Leu 290	Glu	Gly	Leu	Asn	Val 295	Leu	V al	Thr	Ile	Ser 300	Gly	Lys	Lys	Asn
Lys 305	Leu	Arg	Val	Tyr	Tyr 310	Leu	Ser	Trp	Leu	Arg 315	Asn	Arg	Ile	Leu	His 320
Asn	Asp	Pro	Glu	Val 325	Glu	Lys	Lys	Gln	Gly 330	Trp	Ile	Thr	Val	Gly 335	Asp
Leu	Glu	Gly	Cys 340	Ile	His	Tyr	Lys	Val 345	Val	Lys	Tyr	Glu	Arg 350	Ile	Lys
Phe	Leu	Val 355	Ile	Ala	Leu	Lys	Asn 360	Ala	Val	Glu	Ile	Tyr 365	Ala	Trp	Ala
Pro	Lys 370	Pro	Tyr	His	Lys	Phe 375	Met	Ala	Phe	Lys	Ser 380	Phe	Ala	Asp	Leu
Gln 385	His	Lys	Pro	Leu	Leu 390	Val	Asp	Leu	Thr	Val 395	Glu	Glu	Gly	Gln	Arg 400
Leu	Lys	Val	Ile	Phe 405	Gly	Ser	His	Thr	Gly 410	Phe	His	Val	Ile	Asp 415	Val
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		Val					Ile					Pro			gct Ala	192
	Gly					Tyr					Val				ggg Gly 80	240
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US99/20631

			. 0.7.687772003	
IPC(6) US CL	SSIFICATION OF SUBJECT MATTER Please See Extra Sheet. 435/194, 6, 15, 7.1, 320.1, 325, 252.3; 530/387.1; 53 International Patent Classification (IPC) or to both		and IPC	·
B. FIEL	DS SEARCHED			
Minimum d	ocumentation searched (classification system followed	by classification syn	nbols)	
U.S. :	435/194, 6, 15, 7.1, 320.1, 325, 252.3; 530/387.1; 536	5/23.2		
Documentat	ion searched other than minimum documentation to the	extent that such docu	ments are included	in the fields searched
•	lata base consulted during the international search (na T. MEDLINE, EMBASE, BIOSIS, SCISEARCH, BIO			•
C. DOC	UMENTS CONSIDERED TO BE RELEVANT			
Category*	Citation of document, with indication, where ap	propriate, of the relev	ant passages	Relevant to claim No.
X	WO 9811234 A2 (INCYTE PHARMAC 1998, see abstract and their SEQ ID I acid sequence with 67% homology to comprising a fragment encoding at lead of SEQ ID NO:2 of this invention.	NO:1 for encodi claimed SEQ I	ing an amino D NO:2 and	5-26
· -				
	<u> </u>		·	
Furt	ner documents are listed in the continuation of Box C	. See pate	at family annex.	
•	necial categories of cited documents:	date and not		ternational filing date or priority
	be of particular relevance riser document published on or after the international filling date	"X" document of considered no	particular relevance; the	ne claimed invention cannot be sred to involve an inventive step
cit sp "O" do	soument which may throw doubts on priority claim(s) or which is ted to establish the publication data of another citation or other soint reason (as specified) soument referring to an oral disclosure, use, exhibition or other seaso	°Y° document of considered to combined with	o, involve en inventiv	se claimed invention cannot be a step when the document is the documents, such combination the art
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	actual completion of the international search	Date of mailing of		-
	EMBER 1999		03 FEB 2	
Commission Box PCT	mailing address of the ISA/US mer of Patents and Trademarks e., D.C. 20231	Authorized officer MARYAM MO	SULLENSHIPOURI	Ta
Facsimile N		Telephone No.	(703) 308-0196	•

INTERNATIONAL SEARCH REPORT

International application No PCT/US99/20631

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims No.: because they relate to subject matter not required to be searched by this Authority, namely:
Claims Nos.: 1-2, 4-26 partially because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
These claims refer to SEQ ID NO:s but the case does not comply with sequence rules. Only SEQ ID NOs: 1-3 which were elected and searched in the parent U.S. application could be searched in the instant case.
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee
As only some of the required additional search tees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.

	PCT/US99/20631
A. CLASSIFICATION OF SUBJECT MATTER: IPC (6):	
C12N 15/00, 5/00, 1/20, 9/12; C07K 16/00; C07H 21/04; G01N 33/53	
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Form PCT/ISA/210 (extra sheet)(July 1992)*



CORRECTED VERSION

(19) World Intellectual Property Organization International Bureau





(43) International Publication Date 16 March 2000 (16.03.2000)

(10) International Publication Number WO 00/14212 A1

(51) International Patent Classification6: C12N 15/00, 5/00, 1/20, 9/12, C07K 16/00, C07H 21/04, G01N 33/53

(75) Inventor/Applicant (for US only): ACTON, Susan [US/US]; 42 Munroe Road, Lexington, MA 02421 (US).

(21) International Application Number: PCT/US99/20631

(74) Agents: MANDRAGOURAS, Amy, E.; Lahive & Cockfield, LLP, 28 State Street, Boston, MA 02109 et al. (US).

(22) International Filing Date:

9 September 1999 (09.09.1999)

(81) Designated States (national): AU, CA, JP, US.

(48) Date of publication of this corrected version:

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(54) Title: NOVEL PROTEIN KINASE MOLECULES AND USES THEREFOR

(57) Abstract: The invention provides isolated nucleic acids molecules, designated CSAPK nucleic acid molecules, which encode novel protein kinases. The invention also provides antisense nucleic acid molecules, recombinant expression vectors containing CSAPK nucleic acid molecules, host cells into which the expression vectors have been introduced, and nonhuman transgenic animals in which a CSAPK gene has been introduced or disrupted. The invention still further provides isolated CSAPK proteins, fusion proteins, antigenic peptides and anti-CSAPK antibodies. Diagnostic, screening, and therapeutic methods utilizing compositions of the invention are also provided.

NOVEL PROTEIN KINASE MOLECULES AND USES THEREFOR

Background of the Invention

Phosphate tightly associated with protein has been known since the late

nineteenth century. Since then, a variety of covalent linkages of phosphate to proteins have been found. The most common involve esterification of phosphate to serine, threonine, and tyrosine with smaller amounts being linked to lysine, arginine, histidine, aspartic acid, glutamic acid, and cysteine. The occurrence of phosphorylated proteins implies the existence of one or more protein kinases capable of phosphorylating amino acid residues on proteins, and also of protein phosphatases capable of hydrolyzing phosphorylated amino acid residues on proteins.

Protein kinases play critical roles in the regulation of biochemical and morphological changes associated with cellular growth and division (D'Urso, G. et al. (1990) Science 250: 786-791; Birchmeier. C. et al. (1993) Bioessays 15: 185-189). They serve as growth factor receptors and signal transducers and have been implicated in cellular transformation and malignancy (Hunter. T. et al. (1992) Cell 70: 375-387; Posada, J. et al. (1992) Mol. Biol. Cell 3: 583-592; Hunter, T. et al. (1994) Cell 79: 573-582). For example, protein kinases have been shown to participate in the transmission of signals from growth-factor receptors (Sturgill, T. W. et al. (1988) Nature 344: 715-718; Gomez, N. et al. (1991) Nature 353: 170-173), control of entry of cells into mitosis 20 (Nurse, P. (1990) Nature 344: 503-508; Maller, J. L. (1991) Curr. Opin. Cell Biol. 3: 269-275) and regulation of actin bundling (Husain-Chishti, A. et al. (1988) Nature 334: 718-721). Protein kinases can be divided into two main groups based on either amino acid sequence similarity or specificity for either serine/threonine or tyrosine residues. A small number of dual-specificity kinases are structurally like the serine/threoninespecific group. Within the broad classification, kinases can be further sub-divided into families whose members share a higher degree of catalytic domain amino acid sequence identity and also have similar biochemical properties. Most protein kinase family members also share structural features outside the kinase domain that reflect their particular cellular roles. These include regulatory domains that control kinase activity or interaction with other proteins (Hanks, S.K. et al. (1988) Science 241: 42-52).

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least in part, on the discovery of novel nucleic uch nucleic acid molecules, referred to herein tein Kinase" ("CSAPK") proteins. The s of the present invention are useful as f cellular processes, e.g., cardiac cellular invention provides isolated nucleic acid logically active portions thereof, as well as r hybridization probes for the detection of

c acid molecule of the invention is at least , 95%, 98% or more homologous to a of the nucleotide sequence) including SEQ reof. In another embodiment, a CSAPK 6, 65%, 70%, 75%, 80%, 85%, 86%, 90%,

....... sequence including SEQ ID NO:4, SEQ ID

NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a complement thereof. In yet another embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 97%, 98% homologous to a nucleotide sequence including SEQ ID NO:7, SEQ ID NO:9, or a complement thereof. In yet another embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 73%, 75%, 80%, 85%, 86%, 90%, 95%, 98% homologous to a nucleotide sequence including SEQ ID NO:10, SEQ ID NO:12, or a complement thereof. In a further embodiment, a CSAPK nucleic acid molecule is 60%, 65%, 70%, 75%, 78%, 80%, 85%, 90%, 95%, 98% homologous to a nucleotide sequence including SEQ ID NO:13, SEQ ID NO:15, or a complement thereof.

In a preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:1 or 3, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:3 and nucleotides 1-296 of SEQ ID NO:1. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:3 and nucleotides 1202-4137 of SEQ ID NO:1. In another preferred embodiment,

the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:1 or 3. In another preferred embodiment, the nucleic acid molecule comprises a fragment of at least 509 nucleotides of the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, or a complement thereof.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:4, 6, 16, 17, 18, 19, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:6 and nucleotides 1-46 of SEQ ID NO:4. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:6 and nucleotides 1411-2120 of SEQ ID NO:4. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:4, 6, 16, 17, 18, 19.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:7 or 9, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:9 and nucleotides 1-50 of SEQ ID NO:7. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:9 and nucleotides 1793-2454 of SEQ ID NO:7. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:7 or 9.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:10 or 12, or a complement thereof. In another embodiment, the nucleic acid molecule includes SEQ ID NO:12 and nucleotides 1-274 of SEQ ID NO:10. In yet another embodiment, the nucleic acid molecule includes SEQ ID NO:12 and nucleotides 755-1864 of SEQ ID NO:10. In another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:10 or 12.

In another preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown in SEQ ID NO:13 or 15, or a complement thereof. In yet another preferred embodiment, the nucleic acid molecule has the nucleotide sequence shown in SEQ ID NO:13 or 15.

In another embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or

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SEO ID NO:14. In a preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:2 (e.g., the entire amino acid sequence of SEQ ID NO:2). In another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 42%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:5 (e.g., the entire amino acid sequence of SEQ ID NO:5). In yet another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an 10 amino acid sequence at least 41%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:8 (e.g., the entire amino acid sequence of SEQ ID NO:8). In a further preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 59%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:11 (e.g., the entire amino acid sequence of SEQ ID NO:11). In another preferred embodiment, a CSAPK nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:14).

In another preferred embodiment, an isolated nucleic acid molecule encodes the amino acid sequence of a human CSAPK. In yet another preferred embodiment, the nucleic acid molecule includes a nucleotide sequence encoding a protein which includes the amino acid sequence of SEQ ID NO: 2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14. In yet another preferred embodiment, the nucleic acid molecule includes a nucleotide sequence encoding a protein having the amino acid sequence of SEQ ID NO: 2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14.

Another embodiment of the invention features nucleic acid molecules, preferably

CSAPK nucleic acid molecules, which specifically detect CSAPK nucleic acid

molecules relative to nucleic acid molecules encoding non-CSAPK proteins. For

example, in one embodiment, such a nucleic acid molecule is at least 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, or 800 nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, 19, or a complement thereof.

In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEO ID NO:2, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:1 or SEQ ID NO:3 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:5, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:8, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:7 or SEQ ID NO:9 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:10 or SEQ ID NO:12 under stringent conditions. In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide which includes the amino acid sequence of SEQ ID NO:14, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule which includes SEQ ID NO:13 or SEQ ID NO:15 under stringent conditions.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to a CSAPK nucleic acid molecule, e.g., the coding strand of a CSAPK nucleic acid molecule.

Another aspect of the invention provides a vector comprising a CSAPK nucleic acid molecule. In certain embodiments, the vector is a recombinant expression vector. In another embodiment, the invention provides a host cell containing a vector of the invention. The invention also provides a method for producing a protein, preferably a CSAPK protein, by culturing in a suitable medium, a host cell, e.g., a mammalian host cell such as a non-human mammalian cell, of the invention containing a recombinant expression vector, such that the protein is produced.

Another aspect of this invention features isolated or recombinant CSAPK proteins and polypeptides. In one embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATPbinding site. In another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 51%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 99% or more homologous to an amino acid sequence including SEQ ID NO:2. In yet another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATPbinding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-1 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, at least one leucine zipper- basic region, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity

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kinase catalytic domain and at least one ATP-binding site and has an amino acid sequence which is at least 42%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:5. In yet another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, and at least one ATPbinding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, and at least one ATPbinding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-2 protein, includes at least one dual specificity kinase catalytic domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

In yet another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 41%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:8. In yet another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-3 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which

hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:7 or SEQ ID NO:9.

In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 59%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:11. In yet another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kings domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated with cell cycle regulation. In another embodiment, the isolated protein, preferably a CSAPK-4 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEO ID NO:10 or SEO ID NO:12.

In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site. In yet another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and has an amino acid sequence which is at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:14. In yet another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is expressed and/or functions in cells of the cardiovascular system. In an even further embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and plays a role in signalling pathways associated with cellular growth, e.g., signalling pathways associated

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with cell cycle regulation. In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is able to interact with Nck (described in Lehman et al. (1990) Nucleic Acids Res. 18:1048). In another embodiment, the isolated protein, preferably a human CSAPK-5 protein, includes at least one Ser/Thr kinase domain, and at least one ATP-binding site and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:13 or SEQ ID NO:15.

In another embodiment, the isolated protein, preferably a CSAPK protein, has an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14. In a preferred embodiment, the protein, preferably a CSAPK protein, has an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to an amino acid sequence including SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11. or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14). In another embodiment, the invention features fragments of the proteins having the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID 8, SEQ ID NO:11, or SEQ ID NO:14, wherein the fragment comprises at least 15 amino acids (e.g., contiguous amino acids) of the amino acid sequence of SEQ ID NO:2, SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, or SEQ ID NO:11, or SEQ ID NO:14, respectively. In another embodiment, the protein, preferably a CSAPK protein, has the amino acid sequence of SEQ ID NO:2, SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:5, SEQ ID NO:14.

Another embodiment of the invention features an isolated protein, preferably a CSAPK protein, which is encoded by a nucleic acid molecule having a nucleotide sequence at least about 50%, 54%, 55%, 60%, 62%, 65%, 70%, 75%, 78%, 80%, 85%, 86%, 90%, 95%, 97%, 98% or more homologous to a nucleotide sequence (e.g., to the entire length of the nucleotide sequence) including SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, or a complement thereof. This invention further features an isolated protein,

preferably a CSAPK protein, which is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a complement thereof.

The proteins of the present invention or biologically active portions thereof, can be operatively linked to a non-CSAPK polypeptide (e.g., heterologous amino acid sequences) to form fusion proteins. The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind proteins of the invention, preferably CSAPK proteins. In addition, the CSAPK proteins or biologically active portions thereof can be incorporated into pharmaceutical compositions, which optionally include pharmaceutically acceptable carriers.

In another aspect, the present invention provides a method for detecting the presence of a CSAPK nucleic acid molecule, protein or polypeptide in a biological sample by contacting the biological sample with an agent capable of detecting a CSAPK nucleic acid molecule, protein or polypeptide such that the presence of a CSAPK nucleic acid molecule, protein or polypeptide is detected in the biological sample.

In another aspect, the present invention provides a method for detecting the presence of CSAPK activity in a biological sample by contacting the biological sample with an agent capable of detecting an indicator of CSAPK activity such that the presence of CSAPK activity is detected in the biological sample.

In another aspect, the invention provides a method for modulating CSAPK activity comprising contacting a cell capable of expressing CSAPK with an agent that modulates CSAPK activity such that CSAPK activity in the cell is modulated. In one embodiment, the agent inhibits CSAPK activity. In another embodiment, the agent stimulates CSAPK activity. In one embodiment, the agent is an antibody that specifically binds to a CSAPK protein. In another embodiment, the agent modulates expression of CSAPK by modulating transcription of a CSAPK gene or translation of a CSAPK mRNA. In yet another embodiment, the agent is a nucleic acid molecule having

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a nucleotide sequence that is antisense to the coding strand of a CSAPK mRNA or a CSAPK gene.

In one embodiment, the methods of the present invention are used to treat a subject having a disorder characterized by aberrant CSAPK protein or nucleic acid expression or activity by administering an agent which is a CSAPK modulator to the subject. In one embodiment, the CSAPK modulator is a CSAPK protein. In another embodiment the CSAPK modulator is a CSAPK nucleic acid molecule. In yet another embodiment, the CSAPK modulator is a peptide, peptidomimetic, or other small molecule. In a preferred embodiment, the disorder characterized by aberrant CSAPK protein or nucleic acid expression is a cellular growth related disorder, e.g., a cardiovascular disorder.

The present invention also provides a diagnostic assay for identifying the presence or absence of a genetic alteration characterized by at least one of (i) aberrant modification or mutation of a gene encoding a CSAPK protein; (ii) mis-regulation of the gene; and (iii) aberrant post-translational modification of a CSAPK protein, wherein a wild-type form of the gene encodes a protein with a CSAPK activity.

In another aspect the invention provides a method for identifying a compound that binds to or modulates the activity of a CSAPK protein, by providing an indicator composition comprising a CSAPK protein having CSAPK activity, contacting the indicator composition with a test compound, and determining the effect of the test compound on CSAPK activity in the indicator composition to identify a compound that modulates the activity of a CSAPK protein.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

Brief Description of the Drawings

Figure 1 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-1. The nucleotide sequence corresponds to nucleic acids 1 to 4137 of SEQ ID NO:1. The amino acid sequence corresponds to amino acids 1 to 302 of SEQ ID NO:2. The coding region without the 5' and 3' untranslated regions of the human CSAPK-1 gene is shown in SEQ ID NO:3.

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Figure 2 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-2. The nucleotide sequence corresponds to nucleic acids 1 to 2120 of SEQ ID NO:4. The amino acid sequence corresponds to amino acids 1 to 455 of SEQ ID NO:5. The coding region without the 5' and 3' untranslated regions of the human CSAPK-2 gene is shown in SEQ ID NO:6.

Figure 3 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-3. The nucleotide sequence corresponds to nucleic acids 1 to 2454 of SEQ ID NO:7. The amino acid sequence corresponds to amino acids 1 to 581 of SEQ ID NO:8. The coding region without the 5' and 3' untranslated regions of the human CSAPK-3 gene is shown in SEQ ID NO:9.

Figure 4 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-4. The nucleotide sequence corresponds to nucleic acids 1 to 1864 of SEQ ID NO:10. The amino acid sequence corresponds to amino acids 1 to 160 of SEQ ID NO:11. The coding region without the 5' and 3' untranslated regions of the human CSAPK-4 gene is shown in SEQ ID NO:12.

Figure 5 depicts the cDNA sequence and predicted amino acid sequence of human CSAPK-5. The nucleotide sequence corresponds to nucleic acids 1 to 1333 of SEQ ID NO:13. The amino acid sequence corresponds to amino acids 1 to 444 of SEQ ID NO:14. The coding region without the 5' and 3' untranslated regions of the human CSAPK-5 gene is shown in SEQ ID NO:15.

Figure 6 depicts a schematic diagram of the MLK (CSAPK-2) overexpression construct used to create the MLK (CSAPK-2) transgenic animals.

Figure 7 is a diagramatic depiction of the MLK (CSAPK-2) cDNA splice variants.

Figure 8 is a diagramatic depiction of the resulting proteins from the MLK (CSAPK-2) splice variants.

Figure 9 depicts the cDNA sequence of the human splice variant CSAPK-2a. The nucleotide sequence corresponds to nucleic acids 1 to 2194 of SEQ ID NO:16.

Figure 10 depicts the cDNA sequence of the human splice variant CSAPK-2b.

30 The nucleotide sequence corresponds to nucleic acids 1 to 2254 of SEQ ID NO:17.

Figure 11 depicts the cDNA sequence of the human splice variant CSAPK-2c. The nucleotide sequence corresponds to nucleic acids 1 to 2272 of SEQ ID NO:18.

Figure 12 depicts the cDNA sequence of the human splice variant CSAPK-2d. The nucleotide sequence corresponds to nucleic acids 1 to 2069 of SEQ ID NO:19.

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Detailed Description of the Invention

The present invention is based, at least in part, on the discovery of novel molecules, referred to herein as "Cardiovascular System Associated Protein Kinase" or "CSAPK" nucleic acid and polypeptide molecules, which play a role in or function in signalling pathways associated with cellular growth. In one embodiment, the CSAPK molecules modulate the activity of one or more proteins involved in cellular growth or differentiation, e.g., cardiac cell growth or differentiation. In another embodiment, the CSAPK molecules of the present invention are capable of modulating the phosphorylation state of a CSAPK molecule or one or more proteins involved in cellular growth or differentiation, e.g., cardiac cell growth or differentiation.

In a preferred embodiment, the CSAPK molecules are protein kinases which are expressed and/or function in cells of the cardiovascular system, e.g., cells of the heart, the blood vessels, and/or the blood.

As used herein, the term "protein kinase" includes a protein or polypeptide which is capable of modulating its own phosphorylation state or the phosphorylation state of another protein or polypeptide. Protein kinases can have a specificity for (i.e., a specificity to phosphorylate) serine/threonine residues, tyrosine residues, or both serine/threonine and tyrosine residues, e.g., the dual specificity kinases. As referred to herein, protein kinases preferably include a catalytic domain of about 200-400 amino acid residues in length, preferably about 200-300 amino acid residues in length, or more preferably about 250-300 amino acid residues in length, which includes preferably 5-20, more preferably 5-15, or preferably 11 highly conserved motifs or subdomains separated by sequences of amino acids with reduced or minimal conservation.

Specificity of a protein kinase for phosphorylation of either tyrosine or serine/threonine can be predicted by the sequence of two of the subdomains (VIb and VIII) in which different residues are conserved in each class (as described in, for example, Hanks et al.

(1988) Science 241:42-52) the contents of which are incorporated herein by reference). These subdomains are also described in further detail herein.

Protein kinases play a role in signalling pathways associated with cellular growth. For example, protein kinases are involved in the regulation of signal transmission from cellular receptors, e.g., growth-factor receptors; entry of cells into mitosis; and the regulation of cytoskeleton function, e.g., actin bundling. Thus, the CSAPK molecules of the present invention may be involved in: 1) the regulation of transmission of signals from cellular receptors, e.g., cardiac cell growth factor receptors; 2) the modulation of the entry of cells, e.g., cardiac precursor cells, into mitosis; 3) the modulation of cellular differentiation; 4) the modulation of cell death; and 5) the regulation of cytoskeleton function, e.g., actin bundling.

Inhibition or over stimulation of the activity of protein kinases involved in signaling pathways associated with cellular growth can lead to perturbed cellular growth, which can in turn lead to cellular growth related disorders. As used herein, a "cellular growth related disorder" includes a disorder, disease, or condition characterized by a deregulation, e.g., an upregulation or a downregulation, of cellular growth. Cellular growth deregulation may be due to a deregulation of cellular proliferation, cell cycle progression, cellular differentiation and/or cellular hypertrophy. Examples of cellular growth related disorders include cardiovascular disorders such as heart failure, hypertension, atrial fibrillation, dilated cardiomyopathy, idiopathic cardiomyopathy, or angina; proliferative disorders or differentiative disorders such as cancer, e.g., melanoma, prostate cancer, cervical cancer, breast cancer, colon cancer, or sarcoma.

The present invention is based, at least in part, on the discovery of novel molecules, referred to herein as CSAPK protein and nucleic acid molecules, which comprise a family of molecules having certain conserved structural and functional features. The term "family" when referring to the protein and nucleic acid molecules of the invention is intended to mean two or more proteins or nucleic acid molecules having a common structural domain or motif and having sufficient amino acid or nucleotide sequence homology as defined herein. Such family members can be naturally or non-naturally occurring and can be from either the same or different species. For example, a family can contain a first protein of human origin, as well as other, distinct proteins of

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human origin or alternatively, can contain homologues of non-human origin. Members of a family may also have common functional characteristics.

One embodiment of the invention features CSAPK nucleic acid molecules, preferably human CSAPK molecules, e.g., CSAPK-1, CSAPK-2, CSAPK-2a, CSAPK-2b, CSAPK-2d, CSAPK-3, CSAPK-4, and CSAPK-5, which were identified from cDNA libraries made from hearts of patients with congestive heart failure (CHF) of ischemic and idiopathic origin. The CSAPK nucleic acid and protein molecules of the invention are described in further detail in the following subsections.

10 A. The CSAPK-1 Nucleic Acid and Protein Molecules

In one embodiment, the isolated proteins of the present invention, preferably CSAPK-1 proteins, are identified based on the presence of at least one "Ser/Thr kinase site" and at least one "ATP-binding region." As used herein, the term "Ser/Thr kinase site" includes an amino acid sequence of about 200-400 amino acid residues in length, preferably 200-300 amino acid residues in length, and more preferably 250-300 amino acid residues in length, which is conserved in kinases which phosphorylate serine and threonine residues and found in the catalytic domain of Ser/Thr kinases. Preferably, the Ser/Thr kinase site includes the following amino acid consensus sequence X9-g-X-G-X₄-V-X₁₂-K-X-₍₁₀₋₁₉₎-E-X₆₆-h-X₈-h-r-D-X-K-X₂-N-X₁₇-K-X₂-D-f-g-X₂₁-p-X₁₃w-X3-g-X55-R-X14-h-X3 (SEQ ID NO:22) (where invariant residues are indicated by upper case letters and nearly invariant residues are indicated by lower case letters). The nearly invariant residues are usually found in most Ser/Thr kinase sites, but can be replaced by other amino acids which, preferably, have similar characteristics. For example, a nearly invariant hydrophobic amino acid in the above amino acid consensus sequence would most likely be replaced by another hydrophobic amino acid. Ser/Thr kinase domains are described in, for example, Levin D.E. et al. (1990) Proc. Natl. Acad. Sci. USA 87:8272-76, the contents of which are incorporated herein by reference.

As used herein, the term "ATP-binding region" includes an amino acid sequence of about 20-40, preferably 20-30, and more preferably 25-30 amino acid residues in length, present in enzymes which activate their substrates by phosphorylation, and involved in binding adenosine triphosphate (ATP). ATP-binding regions preferably

include the following amino acid consensus sequence: G-X-G-X-X-G-X(15-23)-K(SEQ ID NO:23). ATP-binding regions are described in, for example, Samuel K.P. et al. (1987) *FEBS Let.* 218(1): 81-86, the contents of which are incorporated herein by reference. Amino acid residues 40 to 63 of CSAPK-1 comprise an ATP-binding region.

Isolated proteins of the present invention, preferably CSAPK-1 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:1 or SEQ ID NO:3. As used herein, the term "sufficiently homologous" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain) amino acid residues or nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences share common structural domains or motifs and/or a common functional activity. For example, amino acid or nucleotide sequences which share common structural domains have at least 30%, 40%, or 50% homology, preferably 60% homology, more preferably 70%-80%, and even more preferably 90-95% homology across the amino acid sequences of the domains and contain at least one and preferably two structural domains or motifs, are defined herein as sufficiently homologous. Furthermore, amino acid or nucleotide sequences which share at least 30%, 40%, or 50%, preferably 60%, more preferably 70-80%, or 90-95% homology and share a common functional activity are defined herein as sufficiently homologous.

As used interchangeably herein a "CSAPK-1 activity", "biological activity of CSAPK-1" or "functional activity of CSAPK-1", refers to an activity exerted by a CSAPK-1 protein, polypeptide or nucleic acid molecule on a CSAPK-1 responsive cell or a CSAPK-1 protein substrate, as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-1 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-1 proteins and polypeptides having a CSAPK-1 activity. Preferred proteins are CSAPK-1 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region and, preferably, a CSAPK-1 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent

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hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

The nucleotide sequence of the isolated human CSAPK-1 cDNA and the predicted amino acid sequence of the human CSAPK-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-1 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203308. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-1 gene, which is approximately 4137 nucleotides in length, encodes a protein having a molecular weight of approximately 34.7 kD and which is approximately 302 amino acid residues in length. The CSAPK-1 gene is expressed predominantly in heart, skeletal muscle, or the placenta.

B. The CSAPK-2 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-2 proteins, are identified based on the presence of at least one dual specificity kinase catalytic domain and at least one ATP-binding region. In yet a further embodiment, the isolated proteins of the present invention, preferably CSAPK-2 proteins, are identified based on the presence of at least one dual specificity kinase catalytic domain, at least one ATP-binding region, and at least one leucine zipper region.

As used herein, the term "dual specificity kinase catalytic domain" includes an amino acid sequence of 200-400 amino acid residues in length, preferably 200-300 amino acid residues in length, and more preferably 250-300 amino acid residues in length, which includes a kinase catalytic domain whose primary sequence is a hybrid between a serine/threonine kinase catalytic domain and a tyrosine kinase catalytic domain. Kinases containing the dual specificity kinase catalytic domain are capable of phosphorylating both serine/threonine and tyrosine residues. Preferably, a dual

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specificity kinase catalytic domain includes the following amino acid consensus sequence X9-G-X-G-X2-G-X-V-X12-K-X-(10-19)-E-X34-G-X40-H-R-D-X-K-X2-N-X17-K-X2-D-F-G-X19-W-X-A-P-E-X13-W-X7-E-X6-P-X36-C-W-X6-R-P-X-F-X14 (SEQ ID NO:24). Dual specificity kinase catalytic domains are described in, for example, Holzman L. B. et al. (1994) *J. Biol. Chem.* 269:30808-817, the contents of which are incorporated herein by reference. Amino acid residues 31-277 of the CSAPK-2 protein comprise a dual specificity kinase catalytic domain.

As used herein, the term "leucine zipper region" includes a protein domain which contains either leucine or isoleucine residues in every seventh position over a stretch of at least 20-30 amino acid residues, more preferably at least 20-25 amino acid residues. and most preferably at least 22 amino acid residues. Typically, the leucine zipper domain is rich in charged residues and forms a helical structure with a hydrophobic ridge of leucines or isoleucines down one face of the helix. The leucine zipper promotes dimerization through hydrophobic interactions between the leucine residues. Leucine zipper domains are described in, for example, Landschultz et al. (1988) Science 240: 1759-1764, the contents of which are incorporated herein by reference. Amino acid residues 294 to 322 of the CSAPK-2 protein comprise a leucine zipper region. Optionally, the leucine zipper region is found adjacent to a basic region. As used herein, the term "basic region" includes an amino acid sequence of 5-30, preferably 5-20, more preferably 5-15 amino acid residues in length, which is composed of a series of basic and hydrophobic amino acid residues, at either the N-terminus or the C-terminus of the leucine zipper region. For example, a 15 amino acid basic domain can contain at least 6, 8, or 10 basic residues and/or at least 1, 2, or 4 hydrophobic residues. Amino acid residues 407-421 of the CSAPK-2 protein comprise a basic region.

Isolated proteins of the present invention, preferably CSAPK-2 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:5 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:4 or SEQ ID NO:6.

As used interchangeably herein a "CSAPK-2 activity", "biological activity of CSAPK-2" or "functional activity of CSAPK-2", refers to an activity exerted by a CSAPK-2 protein, polypeptide or nucleic acid molecule on a CSAPK-2 responsive cell

or a CSAPK-2 protein substrate, as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-2 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-2 proteins and polypeptides having a CSAPK-2 activity. Preferred proteins are CSAPK-2 proteins having at least one dual specificity kinase catalytic domain and at least one ATP-binding region. Yet further preferred CSAPK-2 proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region, and at least one leucine zipper region. Preferred proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region and, preferably, a CSAPK-2 activity. Additional preferred proteins have at least one dual specificity kinase catalytic domain, at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

The nucleotide sequence of the isolated human CSAPK-2 cDNA and the predicted amino acid sequence of the human CSAPK-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-2 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203306. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-2 gene, which is approximately 2120 nucleotides in length, encodes a protein having a molecular weight of approximately 52.3 kD and which is approximately 455 amino acid residues in length. The CSAPK-2 gene is expressed predominantly in muscle, e.g., skeletal or cardiac muscle. Four splice variants of CSAPK-2 were also identified (see Figures 7-12).

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C. The CSAPK-3 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-3 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. The Ser/Thr kinase site and the ATP-binding region are described herein. Amino acid residues 5-164 of the CSAPK-3 protein comprise a Ser/Thr kinase domain.

Isolated proteins of the present invention, preferably CSAPK-3 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:8 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:7 or SEQ ID NO:9.

As used interchangeably herein a "CSAPK-3 activity", "biological activity of CSAPK-3" or "functional activity of CSAPK-3", refers to an activity exerted by a CSAPK-3 protein, polypeptide or nucleic acid molecule on a CSAPK-3 responsive cell or a CSAPK-3 protein substrate as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-3 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-3 proteins and polypeptides having a CSAPK-3 activity. Preferred proteins are CSAPK-3 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-3 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:7 or SEQ ID NO:9.

The nucleotide sequence of the isolated human CSAPK-3 cDNA and the predicted amino acid sequence of the human CSAPK-3 polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-3 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203309. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of

Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The CSAPK-3 gene, which is approximately 2454 nucleotides in length, encodes a protein having a molecular weight of approximately 66.8 kD and which is approximately 581 amino acid residues in length. The CSAPK-3 gene is expressed predominantly in heart, skeletal muscle, brain, placenta, lung, liver, kidney, and pancreas.

10 D. The CSAPK-4 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-4 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. The Ser/Thr kinase site and the ATP-binding region are described herein. Amino acid residues 11 to 75 of the CSAPK-4 protein comprise a Ser/Thr kinase domain.

Isolated proteins of the present invention, preferably CSAPK-4 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:11 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:10 or SEQ ID NO:12.

As used interchangeably herein a "CSAPK-4 activity", "biological activity of CSAPK-4" or "functional activity of CSAPK-4", refers to an activity exerted by a CSAPK-4 protein, polypeptide or nucleic acid molecule on a CSAPK-4 responsive cell or a CSAPK-4 protein substrate as determined *in vivo*, or *in vitro*, according to standard techniques. The biological activity of CSAPK-4 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-4 proteins and polypeptides having a CSAPK-4 activity. Preferred proteins are CSAPK-4 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-4 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes

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under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:10 or SEQ ID NO:12.

The nucleotide sequence of the isolated human CSAPK-4 cDNA and the predicted amino acid sequence of the human CSAPK-4 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-4 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203307. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The human CSAPK-4 gene, which is approximately 1864 nucleotides in length, encodes a protein having a molecular weight of approximately 18.4 kD and which is approximately 160 amino acid residues in length. The CSAPK-4 gene is expressed predominantly in skeletal muscle.

E. The CSAPK-5 Nucleic Acid and Protein Molecules

In another embodiment, the isolated proteins of the present invention, preferably CSAPK-5 proteins, are identified based on the presence of at least one Ser/Thr kinase site and at least one ATP-binding region. In yet another embodiment, the isolated proteins of the present invention, preferably CSAPK-5 proteins, are identified based on the presence of at least one Ser/Thr kinase site, and at least one ATP-binding region.

Isolated proteins of the present invention, preferably CSAPK-5 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:14 or are encoded by a nucleotide sequence sufficiently homologous to SEQ ID NO:13 or SEQ ID NO:15.

As used interchangeably herein a "CSAPK-5 activity", "biological activity of CSAPK-5" or "functional activity of CSAPK-5", refers to an activity exerted by a CSAPK-5 protein, polypeptide or nucleic acid molecule on a CSAPK-5 responsive cell

or a CSAPK-5 protein substrate as determined in vivo, or in vitro, according to standard techniques. The biological activity of CSAPK-5 is described herein.

Accordingly, another embodiment of the invention features isolated CSAPK-5 proteins and polypeptides having a CSAPK-5 activity. Preferred proteins are CSAPK-5 proteins having at least one Ser/Thr kinase site and at least one ATP-binding region. Additional preferred proteins have at least one Ser/Thr kinase site, at least one ATP-binding region, and, preferably a CSAPK-5 activity. Additional preferred proteins have at least one Ser/Thr kinase site and at least one ATP-binding region and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:13 or SEQ ID NO:15.

The nucleotide sequence of the isolated human CSAPK-5 cDNA and the predicted amino acid sequence of the human CSAPK-5 polypeptide are shown in Figure 5 and in SEQ ID NOs:13 and 14, respectively. A plasmid containing the nucleotide sequence encoding human CSAPK-1 was deposited with American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on 10/27/98 and assigned Accession Number 203305. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The human CSAPK-5 gene, which is approximately 1333 nucleotides in length, encodes a protein having a molecular weight of approximately 51 kD and which is approximately 444 amino acid residues in length.

Various aspects of the invention are described in further detail in the following subsections:

I. Isolated Nucleic Acid Molecules

One aspect of the invention pertains to isolated nucleic acid molecules that

encode CSAPK proteins or biologically active portions thereof, as well as nucleic acid
fragments sufficient for use as hybridization probes to identify CSAPK-encoding nucleic

acids (e.g., CSAPK mRNA) and fragments for use as PCR primers for the amplification or mutation of CSAPK nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (e.g., cDNA or genomic DNA) and RNA molecules (e.g., mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. For example, with regards to genomic DNA, the term "isolated" includes nucleic acid molecules which are separated from the chromosome with which the genomic DNA is naturally associated. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated CSAPK nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion thereof, can be isolated using standard molecular biology techniques and the sequence information provided herein. For example, using all or portion of the nucleic acid sequence of SEQ ID NO:1, or the nucleotide sequence of SEQ ID NO:3, as a hybridization probe, CSAPK nucleic acid molecules can be isolated using standard hybridization and cloning techniques (e.g., as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring*

Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

Moreover, a nucleic acid molecule encompassing all or a portion of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to CSAPK nucleotide sequences can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:1. The sequence of SEQ ID NO:1 corresponds to the partial human CSAPK-1 cDNA. This cDNA comprises sequences encoding the human CSAPK-1 protein (i.e., "the coding region", from nucleotides 297-1202), as well as 5' untranslated sequences (nucleotides 1-296) and 3' untranslated sequences (nucleotides 1203-4137). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:1 (e.g., nucleotides 297-1202, corresponding to SEQ ID NO:3).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:4. The sequence of SEQ ID NO:4 corresponds to the partial human CSAPK-2 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-2 protein (i.e., "the coding region", from nucleotides 47-1411), as well as 5' untranslated sequences (nucleotides 1-46) and 3' untranslated sequences (nucleotides 1412-2120). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:4 (e.g., nucleotides 47-

1411, corresponding to SEQ ID NO:6). In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:7. The sequence of SEQ ID NO:7 corresponds to the partial human CSAPK-3 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-3 protein (i.e., "the coding region", from nucleotides 51-1793), as well as 5' untranslated sequences (nucleotides 1-50) and 3' untranslated sequences (nucleotides 1794-2454). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:7 (e.g., nucleotides 51-1793, corresponding to SEQ ID NO:9).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:10. The sequence of SEQ ID NO:10 corresponds to the partial human CSAPK-4 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-4 protein (i.e., "the coding region", from nucleotides 275-754), as well as 5' untranslated sequences (nucleotides 1-274) and 3' untranslated sequences (nucleotides 755-1864). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:10 (e.g., nucleotides 275-754, corresponding to SEQ ID NO:12).

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:13. The sequence of SEQ ID NO:13 corresponds to the partial human CSAPK-5 cDNA. This cDNA comprises sequences encoding the partial human CSAPK-5 protein (i.e., "the coding region", from nucleotides 2-1333).

In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion of any of these nucleotide sequences. A nucleic acid molecule which is complementary to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ

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ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 is one which is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively, such that it can hybridize to the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, respectively, thereby forming a stable duplex.

In still another preferred embodiment, an isolated nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least about 50%, 54%, 55%, 60%, 62%, 65%, 70%, 75%, 78%, 80%, 85%, 86%, 90%, 95%, 97%, 98% or more homologous to the nucleotide sequence (e.g., to the entire length of the nucleotide sequence) shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, or a portion of any of these nucleotide sequences.

Moreover, the nucleic acid molecule of the invention can comprise only a portion
of the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID
NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13,
SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 for
example a fragment which can be used as a probe or primer or a fragment encoding a
biologically active portion of a CSAPK protein. The nucleotide sequence determined
from the cloning of the CSAPK gene allows for the generation of probes and primers
designed for use in identifying and/or cloning other CSAPK family members, as well as
CSAPK homologues from other species. The probe/primer typically comprises
substantially purified oligonucleotide. The oligonucleotide typically comprises a region
of nucleotide sequence that hybridizes under stringent conditions to at least about 12 or
15, preferably about 20 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, or 75
consecutive nucleotides of a sense sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID

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NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 of an anti-sense sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 or of a naturally occurring allelic variant or mutant of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In an exemplary embodiment, a nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least 350, 400, 450, 500, 550, 600, 650, 700, 750, or 800 nucleotides in length and hybridizes under stringent hybridization conditions to a nucleic acid molecule of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19.

Probes based on the CSAPK nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments, the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissues which misexpress a CSAPK protein, such as by measuring a level of a CSAPK-encoding nucleic acid in a sample of cells from a subject e.g., detecting CSAPK mRNA levels or determining whether a genomic CSAPK gene has been mutated or deleted.

A nucleic acid fragment encoding a "biologically active portion of a CSAPK protein" can be prepared by isolating a portion of the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, which encodes a polypeptide having a CSAPK biological activity (the biological activities of the CSAPK proteins are described herein), expressing the encoded portion of the CSAPK protein (e.g., by

recombinant expression in vitro) and assessing the activity of the encoded portion of the CSAPK protein.

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID 5 NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEO ID NO:15, SEO ID NO:16, SEO ID NO:17, SEO ID NO:18, or SEO ID NO:19 due to the degeneracy of the genetic code and, thus, encode the same CSAPK proteins as those encoded by the nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEO ID NO:6, SEO ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEO ID NO:13, SEO ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14.

In addition to the CSAPK nucleotide sequences shown in SEQ ID NO:1, SEQ ID 15 NO:3, SEO ID NO:4, SEO ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEO ID NO:19 it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the CSAPK proteins may exist within a population (e.g., the human population). Such genetic polymorphism in the CSAPK genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which include an open reading frame encoding an CSAPK protein, preferably a mammalian CSAPK protein, and can further include non-coding regulatory sequences, and introns. Such natural allelic variations include both functional and non-functional CSAPK proteins and can typically result in 1-5% variance in the nucleotide sequence of a CSAPK gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in CSAPK genes that are the result of natural allelic variation and that do not alter the functional activity of a CSAPK protein are intended to be within the scope of the invention.

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Moreover, nucleic acid molecules encoding other CSAPK family members and, thus, which have a nucleotide sequence which differs from the CSAPK sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are intended to be within the scope of the invention. For example, another CSAPK cDNA can be identified based on the nucleotide sequence of human CSAPK. Moreover, nucleic acid molecules encoding CSAPK proteins from different species, and thus which have a nucleotide sequence which differs from the CSAPK sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are intended to be within the scope of the invention. For example, a mouse CSAPK cDNA can be identified based on the nucleotide sequence of a human CSAPK.

Nucleic acid molecules corresponding to natural allelic variants and homologues of the CSAPK cDNAs of the invention can be isolated based on their homology to the CSAPK nucleic acids disclosed herein using the cDNAs disclosed herein, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15, 20, 25, 30 or more nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. In other embodiment, the nucleic acid is at least 30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, or 600 nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 30%, 40%, 50%, or 60% homologous to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 80%, even more preferably at least about 85%

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or 90% homologous to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50°C, 55°C, 60°C, or 65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In addition to naturally-occurring allelic variants of the CSAPK sequences that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, thereby leading to changes in the amino acid sequence of the encoded CSAPK proteins, without altering the functional ability of the CSAPK proteins. For example, nucleotide substitutions leading to amino acid substitutions at "nonessential" amino acid residues can be made in the sequence oSEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of CSAPK (e.g., the sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14) without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the CSAPK proteins of the present invention, are predicted to be particularly unamenable to alteration. Furthermore, additional amino acid residues that are conserved between the

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CSAPK proteins of the present invention and other CSAPK family members are not likely to be amenable to alteration.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding CSAPK proteins that contain changes in amino acid residues that are not essential for activity. Such CSAPK proteins differ in amino acid sequence from SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, or SEQ ID NO:14).

An isolated nucleic acid molecule encoding a CSAPK protein homologous to the protein of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15, respectively. such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15 by standard techniques, such as site-directed mutagenesis and PCRmediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine,

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phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in a CSAPK protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a CSAPK coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for CSAPK biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:15, the encoded protein can be expressed recombinantly and the activity of the protein can be determined.

In a preferred embodiment, a mutant CSAPK protein can be assayed for the ability to: 1) regulate trasmission of signals from cellular receptors, e.g., cardiac cell growth factor receptors; 2) control entry of cells, e.g., cardiac cells, into mitosis; 3) modulate cellular differentiation; 4) modulate cell death; or 5) regulate cytoskeleton function, e.g., actin bundling.

In addition to the nucleic acid molecules encoding CSAPK proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire CSAPK coding strand, or only to a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding CSAPK. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (e.g., the coding region of human CSAPK-1 corresponds to SEQ ID NO:3). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding CSAPK. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding CSAPK disclosed herein (e.g., SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9, SEQ ID NO:12, or SEQ ID NO:15), antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of CSAPK mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding or noncoding region of CSAPK mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of CSAPK mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-Dgalactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5- oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (i.e., RNA transcribed from

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the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated in situ such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding a CSAPK protein to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention include direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an α-anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β-units, the strands run parallel to each other (Gaultier et al. (1987) *Nucleic Acids. Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to

catalytically cleave CSAPK mRNA transcripts to thereby inhibit translation of CSAPK mRNA. A ribozyme having specificity for a CSAPK-encoding nucleic acid can be designed based upon the nucleotide sequence of a CSAPK cDNA disclosed herein (i.e., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19). For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a CSAPK-encoding mRNA. See, e.g., Cech et al. U.S. Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, CSAPK mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, e.g., Bartel, D. and Szostak, J.W. (1993) *Science* 261:1411-1418.

Alternatively, CSAPK gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the CSAPK (e.g., the CSAPK promoter and/or enhancers) to form triple helical structures that prevent transcription of the CSAPK gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. et al. (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher, L.J. (1992) *Bioassays* 14(12):807-15.

In yet another embodiment, the CSAPK nucleic acid molecules of the present invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acid molecules can be modified to generate peptide nucleic acids (see Hyrup B. et al. (1996) *Bioorganic & Medicinal Chemistry* 4 (1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup B. et al. (1996) *supra*; Perry-O'Keefe et al. Proc. Natl. Acad. Sci. 93: 14670-675.

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PNAs of CSAPK nucleic acid molecules can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, for example, inducing transcription or translation arrest or inhibiting replication. PNAs of CSAPK nucleic acid molecules can also be used in the analysis of single base pair mutations in a gene, (e.g., by PNA-directed PCR clamping); as 'artificial restriction enzymes' when used in combination with other enzymes, (e.g., S1 nucleases (Hyrup B. (1996) supra)); or as probes or primers for DNA sequencing or hybridization (Hyrup B. et al. (1996) supra; Perry-O'Keefe supra).

In another embodiment, PNAs of CSAPK can be modified, (e.g., to enhance 10 their stability or cellular uptake), by attaching lipophilic or other helper groups to PNA. by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of CSAPK nucleic acid molecules can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, (e.g., RNAse H and DNA polymerases), to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup B. (1996) supra). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup B. (1996) supra and Finn 20 P.J. et al. (1996) Nucleic Acids Res. 24 (17): 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs, e.g., 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used as a between the PNA and the 5' end of DNA (Mag, M. et al. (1989) Nucleic Acid Res. 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn P.J. et al. (1996) supra). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser, K.H. et al. (1975) Bioorganic Med. Chem. Lett. 5: 1119-11124).

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In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) *Proc. Natl. Acad. Sci. US.* 86:6553-6556; Lemaitre et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (See, e.g., Krol et al. (1988) *Bio-Techniques* 6:958-976) or intercalating agents. (See, e.g., Zon (1988) *Pharm. Res.* 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, (e.g., a peptide, hybridization triggered cross-linking agent, transport agent, or hybridization-triggered cleavage agent).

II. Isolated CSAPK Proteins and Anti-CSAPK Antibodies

One aspect of the invention pertains to isolated CSAPK proteins, and biologically active portions thereof, as well as polypeptide fragments suitable for use as immunogens to raise anti-CSAPK antibodies. In one embodiment, native CSAPK proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, CSAPK proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, a CSAPK protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the CSAPK protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of CSAPK protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of CSAPK protein having less than about 30% (by dry weight) of non-CSAPK protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-CSAPK protein, still more preferably less than about 10% of non-CSAPK protein, and most preferably less than

about 5% non-CSAPK protein. When the CSAPK protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, i.e., culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of CSAPK protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of CSAPK protein having less than about 30% (by dry weight) of chemical precursors or non-CSAPK chemicals, more preferably less than about 20% chemical precursors or non-CSAPK chemicals, still more preferably less than about 10% chemical precursors or non-CSAPK chemicals, and most preferably less than about 5% chemical precursors or non-CSAPK chemicals.

Biologically active portions of a CSAPK protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the CSAPK protein, e.g., the amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, which include less amino acids than the full length CSAPK proteins, and exhibit at least one activity of a CSAPK protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the CSAPK protein. A biologically active portion of a CSAPK protein can be a polypeptide which is, for example, at least 10, 25, 50, 100 or more amino acids in length.

In a preferred embodiment, the CSAPK protein has an amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14. In other embodiments, the CSAPK protein is substantially homologous to SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, and retains the functional activity of the protein of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail in subsection I above.

Accordingly, in another embodiment, the CSAPK protein is a protein which comprises

an amino acid sequence at least about 41%, 42%, 45%, 50%, 55%, 59%, 60%, 65%, 70%, 75%, 80%, 81%, 85%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 (e.g., the entire amino acid sequence of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14).

To determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a preferred embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, or 90% of the length of the reference sequence (e.g., when aligning a second sequence to the CSAPK amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14 having 177 amino acid residues, at least 80, preferably at least 100, more preferably at least 120, even more preferably at least 140, and even more preferably at least 150, 160 or 170 amino acid residues are aligned). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a preferred embodiment, the percent identity between two amino acid sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using either a Blossom 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another preferred

embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6.

The nucleic acid and protein sequences of the present invention can further be used as a "query sequence" to perform a search against public databases to, for example, identify other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (1990) *J. Mol. Biol.* 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences homologous to CSAPK nucleic acid molecules of the invention. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength = 3 to obtain amino acid sequences homologous to CSAPK protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used. See http://www.ncbi.nlm.nih.gov.

The invention also provides CSAPK chimeric or fusion proteins. As used herein, a CSAPK "chimeric protein" or "fusion protein" comprises a CSAPK polypeptide operatively linked to a non-CSAPK polypeptide. An "CSAPK polypeptide" refers to a polypeptide having an amino acid sequence corresponding to CSAPK, whereas a "non-CSAPK polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the CSAPK protein, e.g., a protein which is different from the CSAPK protein and which is derived from the same or a different organism. Within a CSAPK fusion protein the CSAPK polypeptide can correspond to all or a portion of a CSAPK protein. In a preferred embodiment, a CSAPK fusion protein comprises at least one biologically active portion of a CSAPK protein. In another preferred embodiment, a CSAPK fusion protein comprises at least two biologically active portions of a CSAPK protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the CSAPK polypeptide and the

non-CSAPK polypeptide are fused in-frame to each other. The non-CSAPK polypeptide can be fused to the N-terminus or C-terminus of the CSAPK polypeptide.

For example, in one embodiment, the fusion protein is a GST-CSAPK fusion protein in which the CSAPK sequences are fused to the C-terminus of the GST sequences. Such fusion proteins can facilitate the purification of recombinant CSAPK.

In another embodiment, the fusion protein is a CSAPK protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of CSAPK can be increased through use of a heterologous signal sequence.

The CSAPK fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject *in vivo*. The CSAPK fusion proteins can be used to affect the bioavailability of a CSAPK substrate. Use of CSAPK fusion proteins may be useful therapeutically for the treatment of cellular growth related disorders, e.g., cardiovascular disorers. Moreover, the CSAPK-fusion proteins of the invention can be used as immunogens to produce anti-CSAPK antibodies in a subject, to purify CSAPK ligands and in screening assays to identify molecules which inhibit the interaction of CSAPK with a CSAPK substrate.

Preferably, a CSAPK chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression vectors are commercially

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available that already encode a fusion moiety (e.g., a GST polypeptide). A CSAPK-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the CSAPK protein.

The present invention also pertains to variants of the CSAPK proteins which function as either CSAPK agonists (mimetics) or as CSAPK antagonists. Variants of the CSAPK proteins can be generated by mutagenesis, e.g., discrete point mutation or truncation of a CSAPK protein. An agonist of the CSAPK proteins can retain substantially the same, or a subset, of the biological activities of the naturally occurring form of a CSAPK protein. An antagonist of a CSAPK protein can inhibit one or more of the activities of the naturally occurring form of the CSAPK protein by, for example, competitively modulating a cardiovascular system activity of a CSAPK protein. Thus, specific biological effects can be elicited by treatment with a variant of limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the protein has fewer side effects in a subject relative to treatment with the naturally occurring form of the CSAPK protein.

In one embodiment, variants of a CSAPK protein which function as either CSAPK agonists (mimetics) or as CSAPK antagonists can be identified by screening combinatorial libraries of mutants, e.g., truncation mutants, of a CSAPK protein for CSAPK protein agonist or antagonist activity. In one embodiment, a variegated library of CSAPK variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of CSAPK variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential CSAPK sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage display) containing the set of CSAPK sequences therein. There are a variety of methods which can be used to produce libraries of potential CSAPK variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding

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the desired set of potential CSAPK sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, e.g., Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura et al. (1984) *Annu. Rev. Biochem.* 53:323; Itakura et al. (1984) *Science* 198:1056; Ike et al. (1983) *Nucleic Acid Res.* 11:477.

In addition, libraries of fragments of a CSAPK protein coding sequence can be used to generate a variegated population of CSAPK fragments for screening and subsequent selection of variants of a CSAPK protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of a CSAPK coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the CSAPK protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of CSAPK proteins. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recrusive ensemble mutagenesis (REM), a new technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify CSAPK variants (Arkin and Yourvan (1992) *Proc. Natl. Acad. Sci. USA 89*:7811-7815; Delgrave et al. (1993) *Protein Engineering* 6(3):327-331).

In one embodiment, cell based assays can be exploited to analyze a variegated CSAPK library. For example, a library of expression vectors can be transfected into a cell line which ordinarily synthesizes and secretes CSAPK. The transfected cells are then cultured such that CSAPK and a particular mutant CSAPK are secreted and the effect of expression of the mutant on CSAPK activity in cell supernatants can be detected, e.g., by any of a number of enzymatic assays. Plasmid DNA can then be recovered from the cells which score for inhibition, or alternatively, potentiation of CSAPK activity, and the individual clones further characterized.

An isolated CSAPK protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind CSAPK using standard techniques for polyclonal and monoclonal antibody preparation. A full-length CSAPK protein can be used or, alternatively, the invention provides antigenic peptide fragments of CSAPK for use as immunogens. The antigenic peptide of CSAPK comprises at least 8 amino acid residues of the amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 and encompasses an epitope of CSAPK such that an antibody raised against the peptide forms a specific immune complex with CSAPK. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues.

Preferred epitopes encompassed by the antigenic peptide are regions of CSAPK that are located on the surface of the protein, e.g., hydrophilic regions.

A CSAPK immunogen typically is used to prepare antibodies by immunizing a suitable subject, (e.g., rabbit, goat, mouse or other mammal) with the immunogen. An appropriate immunogenic preparation can contain, for example, recombinantly expressed CSAPK protein or a chemically synthesized CSAPK polypeptide. The preparation can further include an adjuvant, such as Freund's complete or incomplete adjuvant, or similar immunostimulatory agent. Immunization of a suitable subject with an immunogenic CSAPK preparation induces a polyclonal anti-CSAPK antibody response.

Accordingly, another aspect of the invention pertains to anti-CSAPK antibodies. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antigen binding site which specifically binds (immunoreacts with) an antigen, such as CSAPK. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind CSAPK. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of CSAPK. A monoclonal antibody composition thus typically displays a single binding affinity for a particular CSAPK protein with which it immunoreacts.

Polyclonal anti-CSAPK antibodies can be prepared as described above by immunizing a suitable subject with a CSAPK immunogen. The anti-CSAPK antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized CSAPK. If desired, the antibody molecules directed against CSAPK can be isolated from the mammal (e.g., from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction. At an appropriate time after immunization, e.g., when the anti-CSAPK antibody titers are highest, antibodyproducing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) Nature 256:495-497) (see also, Brown et al. (1981) J. Immunol. 127:539-46; Brown et al. (1980) J. Biol. Chem .255:4980-83; Yeh et al. (1976) Proc. Natl. Acad. Sci. USA 76:2927-31; and Yeh et al. (1982) Int. J. Cancer 29:269-75), the more recent human B cell hybridoma technique (Kozbor et al. (1983) Immunol Today 4:72), the EBV-hybridoma technique (Cole et al. (1985), Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing monoclonal antibody hybridomas is well known (see generally R. H. Kenneth, in Monoclonal Antibodies: A New Dimension In Biological

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Analyses, Plenum Publishing Corp., New York, New York (1980); E. A. Lerner (1981) Yale J. Biol. Med., 54:387-402; M. L. Gefter et al. (1977) Somatic Cell Genet.

3:231-36). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with a CSAPK immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds CSAPK.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating an anti-CSAPK monoclonal antibody (see, e.g., G. Galfre et al. (1977) Nature 266:55052; Gefter et al. Somatic Cell Genet., cited supra; Lerner, Yale J. Biol. Med., cited supra; Kenneth, Monoclonal Antibodies, cited supra). Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods which also would be useful. Typically, the immortal cell line (e.g., a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line. Preferred immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a number of myeloma cell lines can be used as a fusion partner according to standard techniques, e.g., the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind CSAPK, e.g., using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal anti-CSAPK antibody can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with CSAPK to thereby isolate immunoglobulin library members that bind

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CSAPK. Kits for generating and screening phage display libraries are commercially available (e.g., the Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; and the Stratagene SurfZAPTM Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, Ladner et al. U.S. Patent No. 5,223,409; Kang et al. PCT International Publication No. WO 92/18619; Dower et al. PCT International Publication No. WO 91/17271; Winter et al. PCT International Publication WO 92/20791; Markland et al. PCT International Publication No. WO 92/15679; Breitling et al. PCT International Publication WO 93/01288; McCafferty et al. PCT International Publication No. WO 92/01047; Garrard et al. PCT International Publication No. WO 92/09690; Ladner et al. PCT International Publication No. WO 90/02809; Fuchs et al. (1991) Bio/Technology 9:1370-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; Griffiths et al. (1993) EMBO J 12:725-734; Hawkins et al. (1992) J. Mol. Biol. 226:889-896; Clarkson et al. (1991) Nature 352:624-628; Gram et al. (1992) Proc., Natl. 15 Acad. Sci. USA 89:3576-3580; Garrad et al. (1991) Bio/Technology 9:1373-1377; Hoogenboom et al. (1991) Nuc. Acid Res. 19:4133-4137; Barbas et al. (1991) Proc. Natl. Acad. Sci. USA 88:7978-7982; and McCafferty et al. Nature (1990) 348:552-554.

Additionally, recombinant anti-CSAPK antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson et al. International Application No. PCT/US86/02269; Akira, et al. European Patent Application 184,187; Taniguchi, M., European Patent Application 171,496; Morrison et al. European Patent Application 173,494; Neuberger et al. PCT International Publication No. WO 86/01533; Cabilly et al. U.S. Patent No. 4,816,567; Cabilly et al. European Patent Application 125,023; Better et al. (1988) Science 240:1041-1043; Liu et al. (1987) Proc. Natl. Acad. Sci. USA 84:3439-3443; Liu et al. (1987) J. Immunol. 139:3521-3526; Sun et al. (1987) Proc. Natl. Acad. Sci. USA 84:214-218; Nishimura et 30 al. (1987) Canc. Res. 47:999-1005; Wood et al. (1985) Nature 314:446-449; and Shaw

et al. (1988) J. Natl. Cancer Inst. 80:1553-1559); Morrison, S. L. (1985) Science 229:1202-1207; Oi et al. (1986) BioTechniques 4:214; Winter U.S. Patent 5,225,539; Jones et al. (1986) Nature 321:552-525; Verhoeyan et al. (1988) Science 239:1534; and Beidler et al. (1988) J. Immunol. 141:4053-4060.

An anti-CSAPK antibody (e.g., monoclonal antibody) can be used to isolate 5 CSAPK by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-CSAPK antibody can facilitate the purification of natural CSAPK from cells and of recombinantly produced CSAPK expressed in host cells. Moreover, an anti-CSAPK antibody can be used to detect CSAPK protein (e.g., in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the CSAPK protein. Anti-CSAPK antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, e.g., to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling (i.e., physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, B-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

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III. Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding a CSAPK protein (or a portion thereof). As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA

segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adenoassociated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (e.g., in an in vitro transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to includes promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and those which direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the

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host cell to be transformed, the level of expression of protein desired, and the like. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., CSAPK proteins, mutant forms of CSAPK proteins, fusion proteins, and the like).

The recombinant expression vectors of the invention can be designed for expression of CSAPK proteins in prokaryotic or eukaryotic cells. For example, CSAPK proteins can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in E. coli with vectors containing constitutive or inducible promoters directing the expression of either 15 fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc. Smith, D.B. 25 and Johnson, K.S. (1988) Gene 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Purified fusion proteins can be utilized in CSAPK activity assays, (e.g., direct assays or competitive assays described in detail below), or to generate antibodies specific for CSAPK proteins, for example. In a preferred embodiment, a CSAPK fusion

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protein expressed in a retroviral expression vector of the present invention can be utilized to infect bone marrow cells which are subsequently transplanted into irradiated recipients. The pathology of the subject recipient is then examined after sufficient time has passed (e.g., six (6) weeks).

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann et al., (1988) *Gene* 69:301-315) and pET 11d (Studier et al., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada et al., (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the CSAPK expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerivisae* include pYepSec1 (Baldari, et al., (1987) *Embo J.* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz et al., (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and picZ (InVitrogen Corp., San Diego, CA).

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Alternatively, CSAPK proteins can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith et al. (1983) *Mol. Cell Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, B. (1987) *Nature* 329:840) and pMT2PC (Kaufman et al. (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert et al. (1987) *Genes Dev.* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv. Immunol.* 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) *EMBO J.* 8:729-733) and immunoglobulins (Banerji et al. (1983) *Cell* 33:729-740; Queen and Baltimore (1983) *Cell* 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) *Proc. Natl. Acad. Sci. USA* 86:5473-5477), pancreas-specific promoters (Edlund et al. (1985) *Science* 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally-regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) *Science* 249:374-379) and the α-fetoprotein promoter

(Campes and Tilghman (1989) Genes Dev. 3:537-546).

The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to CSAPK mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or 10 attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub, H. et al., Antisense RNA as a molecular tool for genetic analysis, Reviews - Trends in Genetics, Vol. 1(1) 1986. 15

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, a CSAPK protein can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including

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calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding a CSAPK protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (i.e., express) a CSAPK protein. Accordingly, the invention further provides methods for producing a CSAPK protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding a CSAPK protein has been introduced) in a suitable medium such that a CSAPK protein is produced. In another embodiment, the method further comprises isolating a CSAPK protein from the medium or the host cell.

The host cells of the invention can also be used to produce non-human transgenic animals. Transgenic mice expressing the CSAPK-2 nucleic acid sequence have been prepared (see Example 5). For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which CSAPK-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous CSAPK sequences have been introduced into their genome or homologous recombinant animals in which endogenous CSAPK sequences have been altered. Such animals are useful for studying the function and/or activity of a CSAPK

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and for identifying and/or evaluating modulators of CSAPK activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous CSAPK gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing a CSAPKencoding nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The CSAPK cDNA sequence of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:13 can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of a human CSAPK gene, such as a mouse or rat CSAPK gene, can be used as a transgene. Alternatively, a CSAPK gene homologue, such as another CSAPK family member, can be isolated based on hybridization to the CSAPK cDNA sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 (described further in subsection I above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to a CSAPK transgene to direct expression of a CSAPK protein to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are

described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., Manipulating the Mouse Embryo, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of a CSAPK transgene in its genome and/or expression of CSAPK mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding a CSAPK protein can further be bred to other transgenic animals carrying other

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of a CSAPK gene into which a deletion, addition or substitution has been introduced to thereby alter, e.g., functionally disrupt, the CSAPK gene. The CSAPK gene can be a human gene (e.g., the SEQ ID NO:1, SEQ ID NO:4, SEQ ID 15 NO:7, SEO ID NO:10, SEO ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19), but more preferably, is a non-human homologue of a human CSAPK gene (e.g., a cDNA isolated by stringent hybridization with the nucleotide sequence of SEO ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19). For example, a mouse CSAPK gene can be used to construct a homologous recombination 20 vector suitable for altering an endogenous CSAPK gene in the mouse genome. In a preferred embodiment, the vector is designed such that, upon homologous recombination, the endogenous CSAPK gene is functionally disrupted (i.e., no longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous 25 CSAPK gene is mutated or otherwise altered but still encodes a functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous CSAPK protein). In the homologous recombination vector, the altered portion of the CSAPK gene is flanked at its 5' and 3' ends by additional nucleic acid sequence of the CSAPK gene to allow for homologous recombination to occur between the exogenous CSAPK gene carried by the vector and an endogenous CSAPK gene in an

embryonic stem cell. The additional flanking CSAPK nucleic acid sequence is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the vector (see e.g., Thomas, K.R. and Capecchi, M. R. (1987) Cell 51:503 for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced CSAPK gene has homologously recombined with the endogenous CSAPK gene are selected (see, e.g., Li, E. et al. (1992) Cell 69:915). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see e.g., Bradley, A. in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E.J. Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination vectors and homologous recombinant animals are described further in Bradley, A. (1991) Current Opinion in Biotechnology 2:823-829 and in PCT International Publication Nos.: WO 90/11354 by Le Mouellec et al.; WO 91/01140 by Smithies et al.; WO 92/0968 by Zijlstra et al.; and WO 93/04169 by Berns et al.

In another embodiment, transgenic non-humans animals can be produced which contain selected systems which allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman et al. (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, e.g., by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

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Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. (1997) *Nature* 385:810-813 and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G_0 phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The recontructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

IV. Pharmaceutical Compositions

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The CSAPK nucleic acid molecules, CSAPK proteins, and anti-CSAPK antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents;

antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor ELTM (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as a bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyetheylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a CSAPK protein or anti-CSAPK antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by

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incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid. Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a

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delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (i.e., the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Patent 5,328,470) or by stereotactic injection (see e.g., Chen et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, e.g., retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

V. Uses and Methods of the Invention

The nucleic acid molecules, proteins, protein homologues, and antibodies

described herein can be used in one or more of the following methods: a) screening
assays; b) predictive medicine (e.g., diagnostic assays, prognostic assays, monitoring

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clinical trials, and pharmacogenetics); and c) methods of treatment (e.g., therapeutic and prophylactic). The isolated nucleic acid molecules of the invention can be used, for example, to express CSAPK protein (e.g., via a recombinant expression vector in a host cell in gene therapy applications), to detect CSAPK mRNA (e.g., in a biological sample) or a genetic alteration in a CSAPK gene, and to modulate CSAPK activity, as described further below. The CSAPK proteins can be used to treat disorders characterized by insufficient or excessive production of a CSAPK substrate or production of CSAPK inhibitors. In addition, the CSAPK proteins can be used to screen for naturally occurring CSAPK substrates, to screen for drugs or compounds which modulate CSAPK activity, as well as to treat disorders characterized by insufficient or excessive production of CSAPK protein or production of CSAPK protein forms which have decreased or aberrant activity compared to CSAPK wild type protein. Moreover, the anti-CSAPK antibodies of the invention can be used to detect and isolate CSAPK proteins, regulate the bioavailability of CSAPK proteins, and modulate CSAPK activity.

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A. Screening Assays:

The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, i.e., candidate or test compounds or agents (e.g., peptides, peptidomimetics, small molecules or other drugs) which bind to CSAPK proteins, have a stimulatory or inhibitory effect on, for example, CSAPK expression or CSAPK activity, or have a stimulatory or inhibitory effect on, for example, the expression or activity of a CSAPK substrate.

In one embodiment, the invention provides assays for screening candidate or test compounds which are substrates of a CSAPK protein or polypeptide or biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of a CSAPK protein or polypeptide or biologically active portion thereof, e.g., modulate the ability of CSAPK to interact with its cognate ligand. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring

deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:6909; Erb et al. (1994) Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al. (1994). J. Med. Chem. 37:2678; Cho et al. (1993) Science 261:1303; Carrell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2061; and in Gallop et al. (1994) J. Med. Chem. 37:1233.

Libraries of compounds may be presented in solution (e.g., Houghten (1992) Biotechniques 13:412-421), or on beads (Lam (1991) Nature 354:82-84), chips (Fodor (1993) Nature 364:555-556), bacteria (Ladner USP 5,223,409), spores (Ladner USP '409), plasmids (Cull et al. (1992) Proc Natl Acad Sci USA 89:1865-1869) or on phage (Scott and Smith (1990) Science 249:386-390); (Devlin (1990) Science 249:404-406); (Cwirla et al. (1990) Proc. Natl. Acad. Sci. 87:6378-6382); (Felici (1991) J. Mol. Biol. 222:301-310); (Ladner supra.).

In another embodiment, an assay is a cell-based assay comprising contacting a cell expressing a CSAPK target molecule (e.g., a CSAPK phosphorylation substrate) with a test compound and determining the ability of the test compound to modulate (e.g. stimulate or inhibit) the activity of the CSAPK target molecule. Determining the ability of the test compound to modulate the activity of a CSAPK target molecule can be accomplished, for example, by determining the ability of the CSAPK protein to bind to or interact with the CSAPK target molecule, or by determining the ability of the CSAPK protein to phosphorylate the CSAPK target molecule.

The ability of the CSAPK protein to phosphorylate a CSAPK target molecule can be determined by, for example, an *in vitro* kinase assay. Briefly, a CSAPK target molecule, e.g., an immunoprecipitated CSAPK target molecule from a cell line expressing such a molecule, can be incubated with the CSAPK protein and radioactive ATP, e.g., [γ-32P] ATP, in a buffer containing MgCl₂ and MnCl₂, e.g., 10 mM MgCl₂

and 5 mM MnCl₂. Following the incubation, the immunoprecipitated CSAPK target molecule can be separated by SDS-polyacrylamide gel electrophoresis under reducing conditions, transferred to a membrane, e.g., a PVDF membrane, and autoradiographed. The appearance of detectable bands on the autoradiograph indicates that the CSAPK substrate has been phosphorylated. Phosphoaminoacid analysis of the phosphorylated substrate can also be performed in order to determine which residues on the CSAPK substrate are phosphorylated. Briefly, the radiophosphorylated protein band can be excised from the SDS gel and subjected to partial acid hydrolysis. The products can then be separated by one-dimensional electrophoresis and analyzed on, for example, a phosphoimager and compared to ninhydrin-stained phosphoaminoacid standards.

Determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished by determining direct binding. Determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished, for example, by coupling the CSAPK protein with a radioisotope or enzymatic label such that binding of the CSAPK protein to a CSAPK target molecule can be determined by detecting the labeled CSAPK protein in a complex. For example, CSAPK molecules, e.g., CSAPK proteins, can be labeled with 125I, 35S, 14C, or 3H, either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, CSAPK molecules can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a compound to modulate the interaction between CSAPK and its target molecule, without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of CSAPK with its target molecule without the labeling of either CSAPK or the target molecule. McConnell, H. M. et al. (1992) *Science* 257:1906-1912. As used herein, a "microphysiometer" (e.g., Cytosensor) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between compound and receptor.

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In a preferred embodiment, determining the ability of the CSAPK protein to bind to or interact with a CSAPK target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (e.g., intracellular Ca²⁺, diacylglycerol, IP₃, etc.), detecting catalytic/enzymatic activity of the target an appropriate substrate, detecting the induction of a reporter gene (comprising a target-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, e.g., chloramphenicol acetyl transferase), or detecting a target-regulated cellular response.

In yet another embodiment, an assay of the present invention is a cell-free assay in which a CSAPK protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the CSAPK protein or biologically active portion thereof is determined. Binding of the test compound to the CSAPK protein can be determined either directly or indirectly as described above. In a preferred embodiment, the assay includes contacting the CSAPK protein or biologically active portion thereof with a known compound which binds CSAPK to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a CSAPK protein, wherein determining the ability of the test compound to interact with a CSAPK protein comprises determining the ability of the test compound to preferentially bind to CSAPK or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which a CSAPK protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate (e.g., stimulate or inhibit) the activity of the CSAPK protein or biologically active portion thereof is determined. Determining the ability of the test compound to modulate the activity of a CSAPK protein can be accomplished, for example, by determining the ability of the CSAPK protein to bind to a CSAPK target molecule by one of the methods described above for determining direct binding. Determining the ability of the CSAPK protein to bind to a CSAPK target molecule can also be accomplished using a technology such as real-time Biomolecular Interaction Analysis (BIA). Sjolander, S. and Urbaniczky, C. (1991) *Anal. Chem.* 63:2338-2345

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and Szabo et al. (1995) Curr. Opin. Struct. Biol. 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (e.g., BIAcore). Changes in the optical phenomenon of surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

In an alternative embodiment, determining the ability of the test compound to modulate the activity of a CSAPK protein can be accomplished by determining the ability of the CSAPK protein to further modulate the activity of a CSAPK target molecule (e.g., a CSAPK mediated signal transduction pathway component). For example, the activity of the effector molecule on an appropriate target can be determined, or the binding of the effector to an appropriate target can be determined as previously described.

In yet another embodiment, the cell-free assay involves contacting a CSAPK protein or biologically active portion thereof with a known compound which binds the CSAPK protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the CSAPK protein, wherein determining the ability of the test compound to interact with the CSAPK protein comprises determining the ability of the CSAPK protein to preferentially bind to or modulate the activity of a CSAPK target molecule.

The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of proteins (e.g., CSAPK proteins or biologically active portions thereof, or receptors to which CSAPK binds). In the case of cell-free assays in which a membrane-bound form a protein is used (e.g., a cell surface CSAPK receptor) it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPS), 3-[(3-cholamidopropyl)dimethylamminio]-2-hydroxy-1-propane sulfonate (CHAPSO), or N-dodecyl=N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either CSAPK or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to a CSAPK protein, or interaction of a CSAPK protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase/ CSAPK fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or CSAPK protein, and the mixture incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of CSAPK binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either a CSAPK protein or a CSAPK target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated CSAPK protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with CSAPK protein or target molecules but which do not interfere with binding of the CSAPK protein to its target molecule can be derivatized to the wells of the plate, and unbound target or CSAPK protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized

complexes, include immunodetection of complexes using antibodies reactive with the CSAPK protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the CSAPK protein or target molecule.

In another embodiment, modulators of CSAPK expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of CSAPK mRNA or protein in the cell is determined. The level of expression of CSAPK mRNA or protein in the presence of the candidate compound is compared to the level of expression of CSAPK mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of CSAPK expression based on this comparison. For example, when expression of CSAPK mRNA or protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of CSAPK mRNA or protein expression. Alternatively, when expression of CSAPK mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of CSAPK mRNA or protein expression. The level of CSAPK mRNA or protein expression in the cells can be determined by methods described herein for detecting CSAPK mRNA or protein.

In yet another aspect of the invention, the CSAPK proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al. (1993) *Cell* 72:223-232; Madura et al. (1993) *J. Biol. Chem.* 268:12046-12054; Bartel et al. (1993) *Biotechniques* 14:920-924; Iwabuchi et al. (1993) *Oncogene* 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with CSAPK ("CSAPK-binding proteins" or "CSAPK-bp") and are involved in CSAPK activity. Such CSAPK-binding proteins are also likely to be involved in the propagation of signals by the CSAPK proteins or CSAPK targets as, for example, downstream elements of a CSAPK-mediated signaling pathway.

Alternatively, such CSAPK-binding proteins are likely to be CSAPK inhibitors.

The two-hybrid system is based on the modular nature of most transcription
factors, which consist of separable DNA-binding and activation domains. Briefly, the
assay utilizes two different DNA constructs. In one construct, the gene that codes for a

CSAPK protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming a CSAPK-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the CSAPK protein.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (e.g., a CSAPK modulating agent, an antisense CSAPK nucleic acid molecule, a CSAPK-specific antibody, or a CSAPK-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

B. Detection Assays

Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue typing); and (iii) aid in forensic identification of a biological sample. These applications are described in the subsections below.

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1. Chromosome Mapping

Once the sequence (or a portion of the sequence) of a gene has been isolated, this sequence can be used to map the location of the gene on a chromosome. This process is called chromosome mapping. Accordingly, portions or fragments of the CSAPK nucleotide sequences, described herein, can be used to map the location of the CSAPK genes on a chromosome. The mapping of the CSAPK sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease.

Briefly, CSAPK genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the CSAPK nucleotide sequences. Computer analysis of the CSAPK sequences can be used to predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the CSAPK sequences will yield an amplified fragment.

Somatic cell hybrids are prepared by fusing somatic cells from different mammals (e.g., human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of individual genes to specific human chromosomes. (D'Eustachio P. et al. (1983) *Science* 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycler. Using the CSAPK nucleotide sequences to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can similarly be

used to map a 90, 1p, or 1v sequence to its chromosome include *in situ* hybridization (described in Fan, Y. et al. (1990) *Proc. Natl. Acad. Sci. USA*, 87:6223-27), prescreening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a DNA sequence to a metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a DNA sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good results at a reasonable amount of time. For a review of this technique, see Verma et al., Human Chromosomes: A Manual of Basic Techniques (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are preferred for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, Mendelian Inheritance in Man, available on-line through Johns Hopkins University Welch Medical Library). The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland, J. et al. (1987) *Nature*, 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the CSAPK gene, can be determined. If a mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease.

Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

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2. Tissue Typing

The CSAPK sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an alternative technique which determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the CSAPK nucleotide sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The CSAPK nucleotide sequences of the invention uniquely represent

portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals. The noncoding sequences of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19, can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers which each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences, such as those in SEQ ID NO:3, SEQ ID NO:6, SEQ ID NO:9, SEQ ID NO:12, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

If a panel of reagents from CSAPK nucleotide sequences described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

3. Use of Partial CSAPK Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means for positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, e.g., hair or skin, or body fluids, e.g., blood, saliva, or semen found at a crime scene. The amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

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The sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e. another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Sequences targeted to noncoding regions of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 are particularly appropriate for this use as greater numbers of polymorphisms occur in the noncoding regions, making it easier to differentiate individuals using this technique. Examples of polynucleotide reagents include the CSAPK nucleotide sequences or portions thereof, e.g., fragments derived from the noncoding regions of SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, or SEQ ID NO:19 having a length of at least 20 bases, preferably at least 30 bases.

The CSAPK nucleotide sequences described herein can further be used to provide polynucleotide reagents, e.g., labeled or labelable probes which can be used in, for example, an *in situ* hybridization technique, to identify a specific tissue, e.g., brain tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such CSAPK probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, e.g., CSAPK primers or probes can be used to screen tissue culture for contamination (i.e. screen for the presence of a mixture of different types of cells in a culture).

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C. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining CSAPK protein and/or nucleic acid expression as well as CSAPK activity,

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in the context of a biological sample (e.g., blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant CSAPK expression or activity. The invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with CSAPK protein, nucleic acid expression or activity. For example, mutations in a CSAPK gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby phophylactically treat an individual prior to the onset of a disorder characterized by or associated with CSAPK protein, nucleic acid expression or activity.

Another aspect of the invention pertains to monitoring the influence of agents (e.g., drugs, compounds) on the expression or activity of CSAPK in clinical trials.

These and other agents are described in further detail in the following sections.

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of CSAPK protein or nucleic acid in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting CSAPK protein or nucleic acid (e.g., mRNA, genomic DNA) that encodes CSAPK protein such that the presence of CSAPK protein or nucleic acid is detected in the biological sample. A preferred agent for detecting CSAPK mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to CSAPK mRNA or genomic DNA. The nucleic acid probe can be, for example, a human CSAPK nucleic acid, such as the nucleic acid of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19 or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to CSAPK mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

A preferred agent for detecting CSAPK protein is an antibody capable of binding to CSAPK protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof

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(e.g., Fab or F(ab')2) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect CSAPK mRNA, protein, or genomic DNA in a biological sample in vitro as well as in vivo. For example, in vitro techniques for detection of CSAPK mRNA include Northern hybridizations and in situ hybridizations. In vitro techniques for detection of CSAPK protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. In vitro techniques for detection of CSAPK genomic DNA include Southern hybridizations. Furthermore, in vivo techniques for detection of CSAPK protein include introducing into a subject a labeled anti-CSAPK antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a serum sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control
biological sample from a control subject, contacting the control sample with a
compound or agent capable of detecting CSAPK protein, mRNA, or genomic DNA,
such that the presence of CSAPK protein, mRNA or genomic DNA is detected in the
biological sample, and comparing the presence of CSAPK protein, mRNA or genomic
DNA in the control sample with the presence of CSAPK protein, mRNA or genomic

DNA in the test sample.

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The invention also encompasses kits for detecting the presence of CSAPK in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting CSAPK protein or mRNA in a biological sample; means for determining the amount of CSAPK in the sample; and means for comparing the amount of CSAPK in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect CSAPK protein or nucleic acid.

2. Prognostic Assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant CSAPK expression or activity. For example, the assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing a disorder associated with CSAPK protein, nucleic acid expression or activity. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant CSAPK expression or activity in which a test sample is obtained from a subject and CSAPK protein or nucleic acid (e.g., mRNA, genomic DNA) is detected, wherein the presence of CSAPK protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant CSAPK expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest. For example, a test sample can be a biological fluid (e.g., serum), cell sample, or tissue.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant CSAPK expression or activity. Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant CSAPK expression or activity in which a test sample is obtained and CSAPK protein or nucleic acid expression or activity is detected (e.g., wherein the abundance of CSAPK protein or

nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant CSAPK expression or activity).

The methods of the invention can also be used to detect genetic alterations in a CSAPK gene, thereby determining if a subject with the altered gene is at risk for a disorder associated with the CSAPK gene. In preferred embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding a CSAPK-protein, or the mis-expression of the CSAPK gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of 1) a deletion of one or more nucleotides from a CSAPK gene; 2) an addition of one or more nucleotides to a CSAPK gene; 3) a substitution of one or more nucleotides of a CSAPK gene, 4) a chromosomal rearrangement of a CSAPK gene; 5) an alteration in the level of a messenger RNA transcript of a CSAPK gene, 6) aberrant modification of a CSAPK gene, such as of the methylation pattern of the genomic DNA, 7) the presence of a non-wild type splicing pattern of a messenger RNA transcript of a CSAPK gene, 8) a non-wild type level of a CSAPK-protein, 9) allelic loss of a CSAPK gene, and 10) inappropriate post-translational modification of a CSAPK-protein. As described herein, there are a large number of assay techniques known in the art which can be used for detecting alterations in a CSAPK gene. A preferred biological sample is a tissue or serum sample isolated by conventional means from a subject, e.g., a cardiac tissue sample.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al. (1988) *Science* 241:1077-1080; and Nakazawa et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:360-364), the latter of which can be particularly useful for detecting point mutations in the CSAPK-gene (see Abravaya et al. (1995) *Nucleic Acids Res* .23:675-682). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to a CSAPK gene under conditions

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such that hybridization and amplification of the CSAPK-gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. et al., (1990) *Proc. Natl. Acad. Sci. USA* 87:1874-1878), transcriptional amplification system (Kwoh, D.Y. et al., (1989) *Proc. Natl. Acad. Sci. USA* 86:1173-1177), Q-Beta Replicase (Lizardi, P.M. et al. (1988) *Bio-Technology* 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in a CSAPK gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Patent No. 5.498.531) can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in CSAPK can be identified by hybridizing a sample and control nucleic acids, e.g., DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin, M.T. et al. (1996) *Human Mutation* 7: 244-255; Kozal, M.J. et al. (1996) *Nature Medicine* 2: 753-759). For example, genetic mutations in CSAPK can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. et al. supra. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential ovelapping probes. This step allows the identification

of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the CSAPK gene and detect mutations by comparing the sequence of the sample CSAPK with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on techniques developed by Maxam and Gilbert ((1977) *Proc. Natl. Acad. Sci. USA* 74:560) or Sanger ((1977) *Proc. Natl. Acad. Sci. USA* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, e.g., PCT International Publication No. WO 94/16101; Cohen et al. (1996) *Adv. Chromatogr.* 36:127-162; and Griffin et al. (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

Other methods for detecting mutations in the CSAPK gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers et al. (1985) Science 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes formed by hybridizing (labeled) RNA or DNA containing the wild-type CSAPK sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent which cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton et al. (1988) Proc. Natl Acad Sci USA 85:4397; Saleeba et al.

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(1992) *Methods Enzymol.* 217:286-295. In a preferred embodiment, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in CSAPK cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994) *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on a CSAPK sequence, e.g., a wild-type CSAPK sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in CSAPK genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita et al. (1989) Proc Natl. Acad. Sci USA: 86:2766, see also Cotton (1993) Mutat Res 285:125-144; and Hayashi (1992) Genet Anal Tech Appl 9:73-79). Single-stranded DNA fragments of sample and control CSAPK nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In a preferred embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen et al. (1991) Trends Genet 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers et al. (1985) Nature 313:495). When

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DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys Chem* 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki et al. (1986) Nature 324:163); Saiki et al. (1989) Proc. Natl Acad. Sci USA 86:6230). Such allele specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology which depends on selective PCR amplification may be used in conjunction with the instant invention.

Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs et al. (1989) Nucleic Acids Res. 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner et al. (1993) Tibtech 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini et al. (1992) Mol. Cell Probes 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) Proc. Natl. Acad. Sci USA 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing prepackaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, e.g., in clinical settings to diagnose

patients exhibiting symptoms or family history of a disease or illness involving a CSAPK gene.

Furthermore, any cell type or tissue in which CSAPK is expressed may be utilized in the prognostic assays described herein.

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3. Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (e.g., drugs or compounds) on the expression or activity of a CSAPK protein can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase CSAPK gene expression, protein levels, or upregulate CSAPK activity, can be monitored in clinical trials of subjects exhibiting decreased CSAPK gene expression, protein levels, or downregulated CSAPK activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease CSAPK gene expression, protein levels, or downregulate CSAPK activity, can be monitored in clinical trials of subjects exhibiting increased CSAPK gene expression, protein levels, or upregulated CSAPK activity. In such clinical trials, the expression or activity of a CSAPK gene, and preferably, other genes that have been implicated in a disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes. including CSAPK, that are modulated in cells by treatment with an agent (e.g., compound, drug or small molecule) which modulates CSAPK activity (e.g., identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on a CSAPK associated disorder, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of CSAPK and other genes implicated in the CSAPK associated disorder, respectively. The levels of gene expression (i.e., a gene expression pattern) can be quantified by Northern blot analysis or RT-PCR, as described herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of CSAPK or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be

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determined before, and at various points during treatment of the individual with the agent.

In a preferred embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) comprising the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of a CSAPK protein, mRNA, or genomic DNA in the pre-administration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the CSAPK protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the CSAPK protein, mRNA, or genomic DNA in the pre-administration sample with the CSAPK protein, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of CSAPK to higher levels than detected, i.e., to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of CSAPK to lower levels than detected, i.e. to decrease the effectiveness of the agent. According to such an embodiment, CSAPK expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

C. Methods of Treatment:

The present invention provides for both prophylactic and therapeutic methods of
treating a subject at risk of (or susceptible to) a disorder or having a disorder associated
with aberrant CSAPK expression or activity. With regards to both prophylactic and
therapeutic methods of treatment, such treatments may be specifically tailored or
modified, based on knowledge obtained from the field of pharmacogenomics.

"Pharmacogenomics", as used herein, refers to the application of genomics technologies
such as gene sequencing, statistical genetics, and gene expression analysis to drugs in
clinical development and on the market. More specifically, the term refers the study of

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how a patient's genes determine his or her response to a drug (e.g., a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with either the CSAPK molecules of the present invention or CSAPK modulators according to that individual's drug response genotype. Pharmacogenomics allows a clinician or physician to target prophylactic or therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug-related side effects.

1. Prophylactic Methods

In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with an aberrant CSAPK expression or activity, by administering to the subject a CSAPK or an agent which modulates CSAPK expression or at least one CSAPK activity. Subjects at risk for a disease which is caused or contributed to by aberrant CSAPK expression or activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the CSAPK aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of CSAPK aberrancy, for example, a CSAPK, CSAPK agonist or CSAPK antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating CSAPK expression or activity for therapeutic purposes. Accordingly, in an exemplary embodiment, the modulatory method of the invention involves contacting a cell with a CSAPK or agent that modulates one or more of the activities of CSAPK protein activity associated with the cell. An agent that modulates CSAPK protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of a CSAPK protein (e.g., a CSAPK phosphorylation substrate), a CSAPK

antibody, a CSAPK agonist or antagonist, a peptidomimetic of a CSAPK agonist or antagonist, or other small molecule. In one embodiment, the agent stimulates one or more CSAPK activities. Examples of such stimulatory agents include active CSAPK protein and a nucleic acid molecule encoding CSAPK that has been introduced into the cell. In another embodiment, the agent inhibits one or more CSAPK activites. Examples of such inhibitory agents include antisense CSAPK nucleic acid molecules, anti-CSAPK antibodies, and CSAPK inhibitors. These modulatory methods can be performed in vitro (e.g., by culturing the cell with the agent) or, alternatively, in vivo (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of a CSAPK protein or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) CSAPK expression or activity. In another embodiment, the method involves administering a CSAPK protein or nucleic acid molecule as therapy to compensate for reduced or aberrant CSAPK expression or activity.

Stimulation of CSAPK activity is desirable in situations in which CSAPK is abnormally downregulated and/or in which increased CSAPK activity is likely to have a beneficial effect. For example, stimulation of CSAPK activity is desirable in situations in which a CSAPK is downregulated and/or in which increased CSAPK activity is likely to have a beneficial effect. Likewise, inhibition of CSAPK activity is desirable in situations in which CSAPK is abnormally upregulated and/or in which decreased CSAPK activity is likely to have a beneficial effect.

25 3. Pharmacogenomics

The CSAPK molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on CSAPK activity (e.g., CSAPK gene expression) as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) disorders (e.g, cardiovascular disorders such as congestive heart failure) associated with aberrant CSAPK activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship

between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer a CSAPK molecule or CSAPK modulator as well as tailoring the dosage and/or therapeutic regimen of treatment with a CSAPK molecule or CSAPK modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, for example, Eichelbaum, M. et al. (1996) Clin. Exp. Pharmacol. Physiol. 23(10-11):983-985 and Linder, M.W. et al. (1997) Clin. Chem. 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (e.g., a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants.) Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a

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"SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict a drug response. According to this method, if a gene that encodes a drug target is known (e.g., a CSAPK protein or CSAPK receptor of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (e.g., N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (e.g., a CSAPK molecule or CSAPK modulator of the present invention) can give an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with a CSAPK molecule or CSAPK modulator, such as a modulator identified by one of the exemplary screening assays described herein.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the Figures and the Sequence Listing, are incorporated herein by reference.

EXAMPLES

20 Example 1: Identification and Characterization Human CSAPK cDNA

In this example, the identification and characterization of the genes encoding human CSAPK-1, CSAPK-2, CSAPK-3, CSAPK-4, and CSAPK-5 (also referred to as pjchrb014g05, pjchrb086g01, pJlhba191a08, pjchrb002d04, and pjchrb155a01, respectively) is described. Four splice variants of CSAPK-2 were also identified, referred to herein as CSAPK-2a, CSAK-2b, CSAPK-2c, and CSAPK-2d.

Isolation of the human CSAPK cDNA

The invention is based, at least in part, on the discovery of five human genes encoding members of the CSAPK family. The human CSAPK family members were isolated from cDNA libraries which were prepared from tissue obtained from subjects suffering from congestive heart failure of ischemic and idiopathic origin. Briefly, a

cardiac tissue sample was obtained from a biopsy of a patient suffering from congestive heart failure. mRNA was isolated from the cardiac tissue and a cDNA library was prepared therefrom using art known methods (described in, for example, *Molecular Cloning A Laboratory Manual*, 2nd Ed., ed. by Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory Press: 1989). Positive clones were isolated following comparison to homologs in public protein databases, including a comparison with known kinases and/or examination of the sequence for protein motifs of kinases.

The sequences of the positive clones were determined and found to contain open reading frames. The nucleotide sequence encoding the human CSAPK-1 protein is shown in Figure 1 and is set forth as SEQ ID NO:1. The protein encoded by this nucleic acid comprises about 302 amino acids and has the amino acid sequence shown in Figure 1 and set forth as SEQ ID NO:2. The coding region (open reading frame) of SEQ ID NO:1 is set forth as SEQ ID NO:3. The clone comprising the entire coding region of human CSAPK-1 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203308.

The nucleotide sequence encoding the human CSAPK-2 protein is shown in Figure 2 and is set forth as SEQ ID NO:4. The protein encoded by this nucleic acid comprises about 455 amino acids and has the amino acid sequence shown in Figure 2 and set forth as SEQ ID NO:5. The coding region (open reading frame) of SEQ ID NO:4 is set forth as SEQ ID NO:6. The clone comprising the entire coding region of human CSAPK-2 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209 on October 27, 1998, and assigned Accession No. 203306.

Four splice variants of CSAPK-2 were also identified. These variants are schematically depicted in Figure 7 (for the cDNA) and Figure 8 (for the protein). The start methionines and in-frame stop codons are indicated for each splice variant clone, in relation to the CSAPK-2 clone shown at the top. The dual specificity kinase catalytic domain is also indicated.

The CSAPK-2a (clone 1.1) nucleotide sequence (which includes parts of clones zchr86SA004 and jchrb086g01) is shown in Figure 9 and set forth as SEQ ID NO:16.

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The CSAPK-2b (clone 2) nucleotide sequence (which includes parts of clones jthrc169f03, jthrc089d05, and jchrb086g01) is shown in Figure 10 and set forth as SEQ ID NO:17. This clone was identified and/or is expressed in pulmonary artery smooth muscle.

The CSAPK-2c (clone 3) nucleotide sequence (which includes parts of clones jthvb025c05, jchrb116b08, jthka142df1, and jchrb086g01) is shown in Figure 11 and set forth as SEQ ID NO:18. This clone was identified and/or is expressed in mammary epithelial cells and in keratinocytes.

The CSAPK-2d (clone 4) nucleotide sequence (which includes parts of clones johnc051a01 and jchrb086g01) is shown in Figure 12 and set forth as SEQ ID NO:19. This clone was identified and/or is expressed in colon to liver metastasis tissues.

The nucleotide sequence encoding the human CSAPK-3 protein is shown in Figure 3 and is set forth as SEQ ID NO:7. The protein encoded by this nucleic acid comprises about 581 amino acids and has the amino acid sequence shown in Figure 3 and set forth as SEQ ID NO:8. The coding region (open reading frame) of SEQ ID NO:7 is set forth as SEQ ID NO:9. The clone comprising the entire coding region of human CSAPK-3 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203309.

The nucleotide sequence encoding the human CSAPK-4 protein is shown in Figure 4 and is set forth as SEQ ID NO:10. The protein encoded by this nucleic acid comprises about 160 amino acids and has the amino acid sequence shown in Figure 4 and set forth as SEQ ID NO:11. The coding region (open reading frame) of SEQ ID NO:10 is set forth as SEQ ID NO:12. The clone comprising the entire coding region of human CSAPK-4 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203307.

The nucleotide sequence encoding the human CSAPK-5 protein is shown in Figure 5 and is set forth as SEQ ID NO:13. The protein encoded by this nucleic acid comprises about 444 amino acids and has the amino acid sequence shown in Figure 5 and set forth as SEQ ID NO:14. The coding region (open reading frame) of SEQ ID

NO:13 is set forth as SEQ ID NO:15. The clone comprising the entire coding region of human CSAPK-5 was deposited with the American Type Culture Collection (ATCC®), 10801 University Boulevard, Manassas, VA 20110-2209, on October 27, 1998, and assigned Accession No. 203305.

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Analysis of Human CSAPK Molecule

A BLASTN 1.4.9 search, using a score of 100 and a word length of 12 (Altschul et al. (1990) *J. Mol. Biol.* 215:403) of the nucleotide sequence of human CSAPK-1 revealed that CSAPK-1 is similar to the human protein kinase HPK-1 coding sequence (Accession No. V23831). This nucleic acid molecule is approximately 70% identical to CSAPK-1, over nucleotides 388-1214.

CSAPK-2 is similar to the human MST mRNA for serine/threonine kinase (Accession No. Z48615). This nucleic acid molecule is approximately 54% identical to CSAPK-2, over nucleotides 482-805.

CSAPK-3 is similar to the human cDNA clone IMAGE:1257327 (Accession No. AA746653). This nucleic acid molecule is approximately 99% identical to CSAPK-3, over nucleotides 2068-2430.

CSAPK-4 is similar to the Mus musculus cDNA clone 902193 (Accession No. AA516800) This nucleic acid molecule is approximately 86% identical to CSAPK-4, over nucleotides 321-755.

CSAPK-5 is similar to the non-annotated human mRNA for KIAA0551 protein (Accession No. AB011123). This nucleic acid molecule is approximately 100% identical to CSAPK-5, over nucleotides 1-1333.

25 Tissue Distribution of CSAPK mRNA

This Example describes the tissue distribution of CSAPK mRNA, as determined by Northern blot hybridizations.

Northern blot hybridizations with the various RNA samples were performed under standard conditions and washed under stringent conditions, i.e., 0.2 X SSC at 65° C. The DNA probe was radioactively labeled with ³²P-dCTP using the Prime-It kit (Stratagene, La Jolla, CA) according to the instructions of the supplier. Filters

containing human mRNA (MultiTissue Northern I and MultiTissue Northern II from Clontech, Palo Alto, CA) were probed in ExpressHyb hybridization solution (Clontech) and washed at high stringency according to manufacturer's recommendations.

CSAPK-1 message was detected predominantly in the heart, skeletal muscle, or the placenta. CSAPK-2 message was detected predominantly in muscle, e.g., skeletal or cardiac muscle. CSAPK-3 message was detected predominantly in heart, skeletal muscle, brain, placenta, lung, liver, kidney, and pancreas. CSAPK-4 message was detected predominantly in skeletal muscle.

10 Example 2: Expression of Recombinant Protein in Bacterial Cells

In this example, CSAPK is expressed as a recombinant glutathione-S-transferase (GST) fusion polypeptide in *E. coli* and the fusion polypeptide is isolated and characterized. Specifically, CSAPK is fused to GST and this fusion polypeptide is expressed in *E. coli*, e.g., strain PEB199. Expression of the GST-CSAPK fusion protein in PEB199 is induced with IPTG. The recombinant fusion polypeptide is purified from crude bacterial lysates of the induced PEB199 strain by affinity chromatography on glutathione beads. Using polyacrylamide gel electrophoretic analysis of the polypeptide purified from the bacterial lysates, the molecular weight of the resultant fusion polypeptide is determined.

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Example 3: Expression of Recombinant CSAPK Protein in COS Cells

To express the CSAPK gene in COS cells, the pcDNA/Amp vector by Invitrogen Corporation (San Diego, CA) is used. This vector contains an SV40 origin of replication, an ampicillin resistance gene, an *E. coli* replication origin, a CMV promoter followed by a polylinker region, and an SV40 intron and polyadenylation site. A DNA fragment encoding the entire CSAPK protein and an HA tag (Wilson et al. (1984) *Cell* 37:767) or a FLAG tag fused in-frame to its 3' end of the fragment is cloned into the polylinker region of the vector, thereby placing the expression of the recombinant protein under the control of the CMV promoter.

To construct the plasmid, the CSAPK DNA sequence is amplified by PCR using two primers. The 5' primer contains the restriction site of interest followed by

approximately twenty nucleotides of the CSAPK coding sequence starting from the initiation codon; the 3' end sequence contains complementary sequences to the other restriction site of interest, a translation stop codon, the HA tag or FLAG tag and the last 20 nucleotides of the CSAPK coding sequence. The PCR amplified fragment and the pCDNA/Amp vector are digested with the appropriate restriction enzymes and the vector is dephosphorylated using the CIAP enzyme (New England Biolabs, Beverly, MA). Preferably the two restriction sites chosen are different so that the CSAPK gene is inserted in the correct orientation. The ligation mixture is transformed into *E. coli* cells (strains HB101, DH5a, SURE, available from Stratagene Cloning Systems, La Jolla, CA, can be used), the transformed culture is plated on ampicillin media plates, and resistant colonies are selected. Plasmid DNA is isolated from transformants and examined by restriction analysis for the presence of the correct fragment.

COS cells are subsequently transfected with the CSAPK-pcDNA/Amp plasmid DNA using the calcium phosphate or calcium chloride co-precipitation methods, DEAE-dextran-mediated transfection, lipofection, or electroporation. Other suitable methods for transfecting host cells can be found in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd. ed., Cold Spring Harbor Laboratory,* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989. The expression of the CSAPK polypeptide is detected by radiolabelling (35S-methionine or 35S-cysteine available from NEN, Boston, MA, can be used) and immunoprecipitation (Harlow, E. and Lane, D. *Antibodies: A Laboratory Manual,* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1988) using an HA specific monoclonal antibody. Briefly, the cells are labelled for 8 hours with 35S-methionine (or 35S-cysteine). The culture media are then collected and the cells are lysed using detergents (RIPA buffer, 150 mM NaCl, 1% NP-40, 0.1% SDS, 0.5% DOC, 50 mM Tris, pH 7.5). Both the cell lysate and the culture media are precipitated with an HA specific monoclonal antibody. Precipitated polypeptides are then analyzed by SDS-PAGE.

Alternatively, DNA containing the CSAPK coding sequence is cloned directly into the polylinker of the pCDNA/Amp vector using the appropriate restriction sites.

The resulting plasmid is transfected into COS cells in the manner described above, and

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the expression of the CSAPK polypeptide is detected by radiolabelling and immunoprecipitation using a CSAPK specific monoclonal antibody.

Example 4: Chromosomal Mapping of CSAPK-2 (MLK)

In order to chromosomally map the CSAPK-2 (MLK) gene, the following primers were designed based on the sequence of jchrb086g01 (CSAPK-2): Forward-5'-GTCCGGTGGAAGTATAATACTTTGTC-3' (SEQ ID NO:20) and Reverse-5'-CTATTTTGAGGAGCTTCTTTACAGCC-3' (SEQ ID NO:21). Using these primers and PCR, a 185 bp product was amplified from human control cell line DNA and from control hamster cell line DNA. These primers were subsequently used used to amplify the 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel.

The PCR reactions were performed by using 5μl of template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR Buffer, 1.2μl Pharmacia dNTP mix (2.5mM), 1.15μl Forward primer 6.6μM, 1.15μl Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq (05U/μl) (Hot Start) and subjecting the reaction mix to 95°C for 10 minutes, followed by 35 Cycles of 94°C for 40 minutes, 55°C for 40 minutes, 72°C for 40 minutes, followed by 72°C for 5 minutes.

The products of the PCR reaction were run on 2% agarose gels, post-stained with SYBR Gold (1:10,000 dilution in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

RH linkage analysis was performed using the Map Manager QTb27 software package.

Clone jchrb086g01 (CSAPK-2 or MLK) was found to map to human chromosome 2, 16.5 cR₃₀₀₀ telomeric to the Whitehead Institute framework marker WI-5538, and 16.5 cR₃₀₀₀ centromeric of the Whitehead framework marker WI-5555. LOD

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scores for linkage were 15.3 for both WI-5538 and WI-5555. This region corresponds to the cytogenetic location 2q23-31.

Example 5: Generation of CSAPK-2 (MLK) Transgenic Animals

An N-terminal His-tagged MLK (CSAPK-2) cDNA was ligated to the cardiacspecific α-myosin heavy chain promoter and the human growth hormone polyadenylation signal sequence using art known techniques (see Figure 6). This construct was microinjected into the male pronucleus of fertilized mouse eggs, which were subsequently implanted into a pseudopregnant foster mother. Transgenic animals were identified by Southern blot analysis and founder mice were bred with nontransgenic cohorts for propagation of individual lines.

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

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What is claimed:

- 1. An isolated nucleic acid molecule selected from the group consisting of:
- (a) a nucleic acid molecule comprising the nucleotide sequence set forth
- 5 in SEQ ID NO:1;
 - (b) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:3;
 - (c) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:4;
- 10 (d) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:6;
 - (e) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:7;
 - (f) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:9;
 - (g) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:10;
 - (h) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:12;
- 20 (i) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:13;
 - (j) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:15;
 - (k) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:16;
 - (l) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:17;
 - (m) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:18; and
- 30 (n) a nucleic acid molecule comprising the nucleotide sequence set forth in SEQ ID NO:19.

- An isolated nucleic acid molecule selected from the group consisting of:

 (a) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2;
 (b) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:5;
 (c) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:8;
 (d) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:11; and
 (e) a nucleic acid molecule which encodes a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:14.
- An isolated nucleic acid molecule selected from the group consisting of:

 (a) a nucleic acid molecule comprising the nucleotide sequence contained
 n the plasmid deposited with ATCC® as Accession Number 203305;
 (b) a nucleic acid molecule comprising the nucleotide sequence
 contained in the plasmid deposited with ATCC® as Accession Number 203306;
 - (c) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203307;
 - (d) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203308; and
- 25 (e) a nucleic acid molecule comprising the nucleotide sequence contained in the plasmid deposited with ATCC® as Accession Number 203309.

4. An isolated nucleic acid molecule selected from the group consisting of:

(a) a nucleic acid molecule which encodes a naturally occurring allelic
variant of a polypeptide comprising the amino acid sequence of SEQ ID
NO:2, wherein the nucleic acid molecule hybridizes to a nucleic acid
molecule comprising SEQ ID NO:1 or 3 under stringent conditions;
(b) a nucleic acid molecule which encodes a naturally occurring allelic
variant of a polypeptide comprising the amino acid sequence of SEQ ID
NO:5, wherein the nucleic acid molecule hybridizes to a nucleic acid
molecule comprising SEQ ID NO:4, 6, 16, 17, 18, or 19 under stringent

10 conditions;

- (c) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:8, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:7 or 9 under stringent conditions;
- (d) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:10 or 12 under stringent conditions; and
- 20 (e) a nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:14, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule comprising SEQ ID NO:13 or 15 under stringent conditions.

5. An isolated nucleic acid molecule selected from the group consisting of: a) a nucleic acid molecule comprising a nucleotide sequence which is at least 60% homologous to the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 15, 16, 17, 18, or 19, or a complement thereof; b) a nucleic acid molecule comprising a fragment of at least 200 5 nucleotides of a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19, or a complement thereof; c) a nucleic acid molecule which encodes a polypeptide comprising an amino acid sequence at least about 60% homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14; and 10 d) a nucleic acid molecule which encodes a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14, wherein the fragment comprises at least 15 contiguous amino acid residues of the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.

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- 6. An isolated nucleic acid molecule which hybridizes to the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 under stringent conditions.
- 7. An isolated nucleic acid molecule comprising a nucleotide sequence which is complementary to the nucleotide sequence of the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5.
 - 8. An isolated nucleic acid molecule comprising the nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5, and a nucleotide sequence encoding a heterologous polypeptide.
 - 9. A vector comprising the nucleic acid molecule of any one of claims 1, 2, 3, 4, or
- 30 10. The vector of claim 9, which is an expression vector.

- 11. A host cell transfected with the vector of claim 9.
- 12. A method of producing a polypeptide comprising culturing a host cell transfected with the vector of claim 9 in an appropriate culture medium to, thereby, produce the polypeptide.
- 13. An isolated polypeptide selected from the group consisting of:
 a) a fragment of a polypeptide comprising the amino acid sequence of SEQ ID
 NO:2, 5, 8, 11, or 14, wherein the fragment comprises at least 15 contiguous
 amino acids of SEQ ID NO:2, 5, 8, 11, or 14;
 - b) a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14, wherein the polypeptide is encoded by a nucleic acid molecule which hybridizes to a nucleic acid molecule comprising SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19 under stringent conditions;
 - c) a polypeptide which is encoded by a nucleic acid molecule comprising a nucleotide sequence which is at least 60 % homologous to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, 12, 13, 15, 16, 17, 18, or 19;
- d) a polypeptide comprising an amino acid sequence which is at least 60% homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.
 - 14. The isolated polypeptide of claim 13 comprising the amino acid sequence of SEQ ID NO:2, 5, 8, 11, or 14.
 - 15. The polypeptide of claim 13, further comprising heterologous amino acid sequences.
 - 16. An antibody which selectively binds to a polypeptide of claim 13.

- 17. A method for detecting the presence of a polypeptide of claim 13 in a sample comprising:
 - a) contacting the sample with a compound which selectively binds to the polypeptide; and
- b) determining whether the compound binds to the polypeptide in the sample to thereby detect the presence of a polypeptide of claim 13 in the sample.
 - 18. The method of claim 17, wherein the compound which binds to the polypeptide is an antibody.
 - 19. A kit comprising a compound which selectively binds to a polypeptide of claim 13 and instructions for use.
- 20. A method for detecting the presence of a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 in a sample comprising:
 - a) contacting the sample with a nucleic acid probe or primer which selectively hybridizes to the nucleic acid molecule; and
 - b) determining whether the nucleic acid probe or primer binds to a nucleic acid molecule in the sample to thereby detect the presence of a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 in the sample.
 - 21. The method of claim 20, wherein the sample comprises mRNA molecules and is contacted with a nucleic acid probe.
- 25 22. A kit comprising a compound which selectively hybridizes to a nucleic acid molecule of any one of claims 1, 2, 3, 4, or 5 and instructions for use.

- 23. A method for identifying a compound which binds to a polypeptide of claim 13 comprising:
 - a) contacting the polypeptide, or a cell expressing the polypeptide with a test compound; and
- b) determining whether the polypeptide binds to the test compound.
 - 24. The method of claim 23, wherein the binding of the test compound to the polypeptide is detected by a method selected from the group consisting of:
 - a) detection of binding by direct detection of test compound/polypeptide binding;
- b) detection of binding using a competition binding assay; and
 - c) detection of binding using an assay for CSAPK activity.
 - 25. A method for modulating the activity of a polypeptide of claim 13 comprising contacting the polypeptide or a cell expressing the polypeptide with a compound which binds to the polypeptide in a sufficient concentration to modulate the activity of the polypeptide.
 - 26. A method for identifying a compound which modulates the activity of a polypeptide of claim 13 comprising:
- a) contacting a polypeptide of claim 13 with a test compound; and
 b) determining the effect of the test compound on the activity of the
 polypeptide to thereby identify a compound which modulates the activity
 of the polypeptide.

TCGACCCACGCGTCCGGGAGGATCGGGAGTCGCGGGAGGATGGGCCGCCGCTAGGCTCGCACTCCGGACGCCCTCGC

CGCCCAAGGTCTCTCGCGGGCGGGAGAACGGAAAACTCCCAACTTCCTGAGTTCTAAAGTTCCTGTTGCTTCAGACAA

TGGATGAGCAATCACAAGGAATGCAAGGGCCACCTGTTCCTCAGTTCCAACCACAGAAGGCCTTACGA CCGGATATGG

AAGCACGTGCTGATTGCATCAAAGAAATAGATCTTCTTAAGCAACTCAACCATCCAAATGTAATAAAA
TATTATGCAT

CATTCATTGAAGATAATGAACTAAACATAGTTTTGGAACTAGCAGATGCTGGCGACCTATCCA GAATGATCAAGCATTTTAAGAAGCAAAAGAGGCTAATTCCTGAAAGAACTGTTTGGAAGTATTTTGTT CAGCTTTGCA

GTGCATTGGAACACATGCATTCTCGAAGAGTCATGCATAGAGATATAAAACCAGCTAATGTGTTCATT
ACAGCCACTG

GGGTGGTAAAACTTGGAGATCTTGGGCTTGGCCGGTTTTTCAGCTCAAAAACCACAGCTGCAC ATTCTTTAGTTGGTACGCCTTATTACATGTCTCCAGAGAGAATACATGAAAATGGATACAACTTCAAA TCTGACATCT

GGTCTCTTGGCTGTCTACTATATGAGATGGCTGCATTACAAAGTCCTTTCTATGGTGACAAAATGAAT TTATACTCAC

AGAGGATGCATGCACTGCAAGCAGCTAAACATGCAAGATCATGAAGAGTGTAACCAAAGTAATT
GAAAGTATTT

TGTGCAAAGTCGTACCTSCCCATTTATGTCTGGGTGTTAAGATTAATATTTCAGAGCTAGTGT GCTCTGAATCCTTAACCAGTTTTCATATAAGCTTCATTTTGTACCAGTCACCTAAATCACCTCCTTGC AACCCCCAAA

TGACTTTGGAATAACTGAATTGCATGTTAGGAGAGAAAATGAAACATGATGGTTTTGAATGGCTAAAG GTTTATAGAA

TTTATGGACATTGAGATGAACACAATTGTGAACTTTTGTGAAGATTTTATTTTTAAACGTTTGAAGTA

TTCTTAGCAGAGTAGTTTTCAAATATGATTCTTATGATAAATGTAGACACAAACTATTTGAGA AACATTTAGAACTCTTAGCTTATACATTCAAAATGTAACTATTAAATGTGAAGATTTGGGGACAAAAT GTGAGTCAGA

CACTGAAGAGTTTTTTGTTTTTAATATTTTTTGATATTCTCTTTTGCATTGAAATGGTATAAATGA ATCCATTTAA

AAAGTGGTTAAGGATTTGTTTAGCTGGTGTGATAATAATTTTTAAAGTTGCACATTGCCCAAG GCTTTTTTTGTGTGTTTTTATTGTTGTTTGTACATTTGAAAAATATTCTTTGAATAACCTTGCAGTAC TATATTTCAA

TTTCTTTATAAATTTAAGTGCATTTTAACTCATAATTGTACACTATAATATAAGCCTAAGTTTTTATT CATAAGTTTT

ATTGAAGTTCTGATCGGTCCCCTTCAGAAATTTTTTTATATTATTCTTCAAGTTACTTTCTTA TTTATATTGTATGTGCATTTTATCCATTAATGTTTCATACTTTCTGAGAGTATAATACCCTTTTAAAA GATATTTGGT

ATACCAATACTTTTCCTGGATTGAAAACTTTTTTTAAACTTTTTAAAATTTGGGCCACTCTGTATGCA TATGTTTGGT

CTTGTTAAAGAGGAAGAAAGGATGTGTTATACTGTACCTGTGAATGTTGATACAGTTACAA TTTATTTGACAAGGTTGTAATTCTAGAATATGCTTAATAAAATGAAAACTGGCCATGACTACAGCCAG AACTGTTATG

AGATTAACATTTCTATTGAGAAGCTTTTGAGTAAAGTACTGTATTTGTTCATGAAGATGACTGAGATG

GTGTAGCTTAAGGAAATGGGCAGAATTTCGTAAATGCTGTTGTGCAGATGTGTTTTCCCTGAA TGCTTTCGTATTAGTGGCGACCAGTTTCTCACAGAATTGTGAAGCCTGAAGGCCAAGAGGAAGTCACT GTTAAAGGAC

TCTGTGCCATCTTACAACCTTGGATGAATTATCCTGCCAACGTGAAAACCTCATGTTCAAAGAACACT TCCCTTTAGC

CGATGTAACTGCTGGTTTTGTTTTTCATATGTGTTTTTCTTACACTCATTTGAATGCTTTCAA GCATTTGTAAACTTAAAAAAANWAWAAAGGGCAAAAAGTCTGAACCCTTGTTTTCTGAAATCTAATC AGTTATGTAT

GGTTTCTGAAGGGTAATTTTATTTTGGAATAGGTAAAGCGAAACCTGTTTTGTCWTGTTTTTCCTGAG GGCTAGATGC

ATTTTTTTTTCTCACACTCTTAATGACTTTTAACATTTATACTGAGCATCCATAGATATATTCC
TAGAAGTATGAGAAGAATTATTCTTATTGACCATTAATGTCATGTTCATTTTAATGTAATATATTGA
GATGAAATGT

TCTCTGGTTGGAACAGATACTCTCTTTTTTTTTTTTTCTTGCAATCTTTAAGAATACATAGATCTAAAATTC
ATTAGCTTGA

CCCCTCAAAGTAACTTTTAAGTAAAGATTAAAGCTTTTCTTCTCAGTGAATATATCTGCTAGA AGGAAATAGCTGGGAAGAATTTAATGATCAGGGAAATTCATTATTTCTATATGTGGAAACTTTTTGCT TCGAATATTG

TATCTTTTTAAATCTAAATGTTCATATTTTTCCTGAAGAAACCACTGTGTAAAAATCAAATTTTAATT
TTGAATGGAA

TAATTTCAAAGAACTATGAAGATGATTTGAAGCTCTAATTTATATAGTCACCTATAAAATGTT CTTTATATGTGTTCATAAGTAAATTTTATATTGATTAAGTTAAACTTTTGAATTGAATTGAGGAGCAG TAAAATGAAA

GCTATATCTATTNCTAAACCYTATTTAGACATTGGKACCAGTTACCCAGGTGAAAATAKGGAGTAACT TTGTTTTGTA

TGGTAAGGTTTAGGAATGGNGGATGAAGGGTATCTCTATATAAATAAAGTGCTCAACAATGTG CAATGATTGTAAATTTAGTAAGATATTACAGCCATTTCATGAATGCTTTACCATTCAACATAGTATCT ATTACAAAAC

CATTAAATGTTTATTCTTTAAAATGAATGTATTATGTTTTTAACCCACAAATGCATACTTACC
CTGTGCCTCATATTTCAATAGTACTGTAATATGGACATCTTTTTGTGAAATACTTTTATTTTGTTATGC
TTTAAATATA

Fig. 1 (continued)

SUBSTITUTE SHEET (RULE 26)

MDEQSQGMQGPPVPQFQPQKALRPDMGYNTLANFRIEKKIGRGQFSEVYRAACLLDGVPVALKKVQIF DLMDAKARAD CIKEIDLLKQLNHPNVIKYYASFIEDNELNIVLELADAGDLSRMIKHFKKQKRLIPERTVWKYFVQLC SALEHMHSRR VMHRDIKPANVFITATGVVKLGDLGLGRFFSSKTTAAHSLVGTPYYMSPERIHENGYNFKSDI WSLGCLLYEMAALQSPFYGDKMNLYSLCKKIEQCDYPPLPSDHYSEELRQLVNMCINPDPEKRPDVTY VYDVAKRMHA CTASS

Fig. 1 (continued)

GTCGACCCACGCGTCCGGTGGAAGTATAATACTTTGTCATTATGAGATGTCGTCTCTCGG TGCCTCCTTTGTGCAAATTAAATTTGATGACTTGCAGTTTTTTGAAAACTGCGGTGGAGG AAGTTTTGGGAGTGTTTATCGAGCCAAATGGATATCACAGGACAAGGAGGTGGCTGTAAA GAAGCTCCTCAAAATAGAGAAAGAGGCAGAAATACTCAGTGTCCTCAGTCACAGAAACAT CATCCAGTTTTATGGAGTAATTCTTGAACCTCCCAACTATGGCATTGTCACAGAATATGC TTCTCTGGGATCACTCTATGATTACATTAACAGTAACAGAAGTGAGGAGATGGATATGGA TCACATTATGACCTGGGCCACTGATGTAGCCAAAGGAATGCATTATTTACATATGGAGGC TCCTGTCAAGGTGATTCACAGAGACCTCAAGTCAAGAAACGTTGTTATAGCTGCTGATGG AGTACTGAAGATCTGTGACTTTGGTGCCTCTCGGTTCCATAACCATACAACACACATGTC CTTGGTTGGAACTTTCCCATGGATGGCTCCAGAAGTTATCCAGAGTCTCCCTGTGTCAGA **AACTTGTGACACATATTCCTATGGTGTGGTTCTCTGGGAGATGCTAACAAGGGAGGTCCC** CTTTAAAGGTTTGGAAGGATTACAAGTAGCTTGGCTTGTAGTGGAAAAAAACGAGAGATT AACCATTCCAAGCAGTTGCCCCAGAAGTTTTGCTGAACTGTTACATCAGTGTTGGGAAGC TGATGCCAAGAAACGGCCATCATTCAAGCAAATCATTTCAATCCTGGAGTCCATGTCAAA TGACACGAGCCTTCCTGACAAGTGTAACTCATTCCTACACAACAAGGCGGAGTGGAGGTG CGAAATTGAGGCAACTCTTGAGAGGCTAAAGAAACTAGAGCGTGATCTCAGCTTTAAGGA GCAGGAGCTTAAAGAACGAGAAAGACGTTTAAAGATGTGGGAGCAAAAGCTGACAGAGCA GTCCAACACCCCGCTTCTCTTGCCTCTTGCTGCAAGAATGTCTGAGGAGTCTTACTTTGA **ATCTAAAACAGAGGAGTCAAACAGTGCAGAGATGTCATGTCAGATCACAGCAACAAGTAA** CGGGGAGGGCCATGGCATGAACCCAAGTCTGCAGGCCATGATGCTGATGGGCTTTGGGGA TATCTTCTCAATGAACAAAGCAGGAGCTGTGATGCATTCTGGGATGCAGATAAACATGCA AGCCAAGCAGAATTCTTCCAAAACCACATCTAAGAGAAGGGGGAAGAAAGTCAACATGGC TCTGGGGTTCAGTGATTTTGACTTGTCAGAAGGTGACGATGATGATGATGATGACGGTGA GGAGGAGGATAATGACATGGATAATAGTGAATGAAAGCAGAAAGCAAAGTAATAAAATCA **CAAATGTTTGGAAAACACAAAAGTAACTTGTTTATCTCAGTCTGTACAAAAACAGTAAGG** AGGCAGAAAGCCAAGCACTGCATTTTTAGGCCAATCACATTTACATGACCGTAATTTCTT ATCAATTCTACTTTTATTTTTGCTTACAGAAAAACGGGGGGAGAATTAAGCCAAAGAAGT **ATATTTATGAATCAGCAAATGTGGTGCCTGATTATAGAAATTTGTGATCCTATATACAAT** ATAGGACTTTTAAAGTTGTGACATTCTGGCTTTTTTCTTTTAATGAATACTTTTTAGTTTG TATTTGACTTTATTTCCTTTATTCAAATCATTTTTAAAAACTTACATTTTGAACAAAACAC TAGCAATACACTGTAATATCAGAAATGGTTGGCCTGAGCAACCTAGTAAGACCTCGTCTC TACTAATAATTAAAAAACTAGCTGGCATGGTAGCACACCTGTAGTCCCAGATACTTGG GATGGCACCACTGCACTCTAGCCTGGGCAAGAGAACAAGATCCTGTCTCAAAAAAACAAAA **AAAAAAAAAAGGGCGGCCG**

MSSLGASFVQIKFDDLQFFENCGGGSFGSVYRAKWISQDKEVAVK KLLKIEKEAEILSVLSHRNIIQFYGVILEPPNYGIVTEYASLGSLYDYINSNRSEEMDMD HIMTWATDVAKGMHYLHMEAPVKVIHRDLKSRNVVIAADGVLKICDFGASRFHNHTTHMS LVGTFPWMAPEVIQSLPVSETCDTYSYGVVLWEMLTREVPFKGLEGLQVAWLVVEKNERL TIPSSCPRSFAELLHQCWEADAKKRPSFKQIISILESMSNDTSLPDKCNSFLHNKAEWRC EIEATLERLKKLERDLSFKEQELKERERRLKMWEQKLTEQSNTPLLLPLAARMSEESYFE SKTEESNSAEMSCQITATSNGEGHGMNPSLQAMMLMGFGDIFSMNKAGAVMHSGMQINMQ AKQNSSKTTSKRRGKKVNMALGFSDFDLSEGDDDDDDDGEEEDNDMDNSE

GACAGGCGCTGGGCCGGCCTGCAGCTGGGTCGAGCGCTGCTGCTGCGCTTCACGGGCA AGCCCGGCCGGGCCTACGGCTTGGGGCGGCCGGGCCGGCGGCGGCTGTGTCCGCGGGG AGCGTCCAGGCTGGGCCGCAGGACCGGGCGCGGAGCCTCGCAGGGTCGGGCTCGGGCTTC CTAACCGTCTCCGCCTTCTTCCGCCAGTCGGTGGCCGGGCTGGCGGCGCGCGGTTGCAGCGGC **AGTTCGTGGTGCGGGCCTGGGGCTGCGCGGGCCCTTGCGGCCGGGCAGTCTTTCTGGCCT** TCGGGCTAGGGCTGGGCCTCATCGAGGAAAAACAGGCGGAGAGCCGGCGGCGGCGGTCTCGG ACACGAGACGCTTGCAGGGCTTTCGGCTGGAGGAGTATCTGATAGGGCAGTCCATTGGTA AGGGCTGCAGTGCTGTGTATGAAGCCACCATGCCTACATTGCCCCAGAACCTGGAGG TGACAAAGAGCACCGGGTTGCTTCCAGGGAGAGGCCCCAGGTACCAGTGCACCAGGAGAAG GGCAGGAGCGAGCTCCGGGGGCCCCTGCCTTCCCCTTGGCCATCAAGATGATGTGGAACA TCTCGGCAGGTTCCTCCAGCGAAGCCATCTTGAACACAATGAGCCAGGAGCTGGTCCCAG CGAGCCGAGTGGCCTTGGCTGGGGAGTATGGAGCAGTCACTTACAGAAAATCCAAGAGAG GTCCCAAGCAACTAGCCCCTCACCCCAACATCATCCGGGGTTCTCCGCGCCTTCACCTCTT CCGTGCCGCTGCTGCCAGGGGCCCTGGTCGACTACCCTGATGTGCTGCCCTCACGCCTCC ACCCTGAAGGCCTGGGCCATGGCCGGACGCTGTTCCTCGTTATGAAGAACTATCCCTGTA CCCTGCGCCAGTACCTTTGTGTGAACACACCCAGCCCCGCCTCGCCGCCATGATGCTGC TGCAGCTGCTGGAAGGCGTGGACCATCTGGTTCAACAGGGCATCGCGCACAGAGACCTGA AATCCGACAACATCCTTGTGGAGCTGGACCCAGACGGCTGCCCCTGGCTGATCGCAG ATTTTGGCTGCCTGGCTGATGAGAGCATCGGCCTGCAGTTGCCCTTCAGCAGCTGGT ACGTGGATCGGGGCGGAAACGGCTGTCTGATGGCCCCAGAGGTGTCCACGGCCCGTCCTG GCCCCAGGGCAGTGATTGACTACAGCAAGGCTGATGCCTGGGCAGTGGGAGCCATCGCCT ATGAAATCTTCGGGCTTGTCAATCCCTTCTACGGCCAGGGCAAGGCCCACCTTGAAAGCC GCAGCTACCAAGAGGCTCAGCTACCTGCACTGCCCGAGTCAGTGCCTCCAGACGTGAGAC AGTTGGTGAGGGCACTGCTCCAGCGAGAGGCCCAGCAAGAGACCATCTGCCCGAGTAGCCG CAAATGTGCTTCATCTAAGCCTCTGGGGTGAACATATTCTAGCCCTGAAGAATCTGAAGT TAGACAAGATGGTTGGCTGGCTCCTCCAACAATCGGCCGCCACTTTGTTGGCCAACAGGC TCACAGAGAAGTGTTGTGTGGAAACAAAAATGAAGATGCTCTTTCTGGCTAACCTGGAGT GTGAAACGCTCTGCCAGGCAGCCCTCCTCCTCTCTCATGGAGGGCAGCCCTGTGATGTC CCTGCATGGAGCTGGTGAATTACTAAAAGAACTTGGCATCCTCTGTGTCGTGATGGTCTG TGAATGGTGAGGGTGGGAGTCAGGAGACAAGACAGCGCAGAGAGGGGCTGGTTAGCCGGAA AAGGCCTCGGGCTTGGCAAATGGAAGAACTTGAGTGAGAGTTCAGTCTGCAGTCCTCTGC TCACAGACATCTGAAAAGTGAATGGCCAAGCTGGTCTAGTAGATGAGGCTGGACTGAGGA GGGGTAGGCCTGCATCCACAGAGAGGATCCAGGCCAAGGCACTGGCTGTCAGTGGCAGAG TTTGGCTGTGACCTTTGCCCCTAACACGAGGAACTCGTTTGAAGGGGGCAGCGTAGCATG TCTGATTTGCCACCTGGATGAAGGCAGACATCAACATGGGTCAGCACGTTCAGTTACGGG **AGTGGGAAATTACATGAGGCCTGGGCCTCTGCGTTCCCAAGCTGTGCGTTCTGGACCAGC** GGGATTTAAACTTGAGGGTTTCCCTCCTGACTAGCCTCTCTTACAGGAATTGTGAAATAT

Val Arg Gln Ala Leu Gly Arg Gly Leu Gln Leu Gly Arg Ala Leu Leu 10 Leu Arg Phe Thr Gly Lys Pro Gly Arg Ala Tyr Gly Leu Gly Arg Pro Gly Pro Ala Ala Gly Cys Val Arg Gly Glu Arg Pro Gly Trp Ala Ala 40 Gly Pro Gly Ala Glu Pro Arg Arg Val Gly Leu Gly Leu Pro Asn Arg Leu Arg Phe Phe Arg Gln Ser Val Ala Gly Leu Ala Ala Arg Leu Gln Arg Gln Phe Val Val Arg Ala Trp Gly Cys Ala Gly Pro Cys Gly Arg Ala Val Phe Leu Ala Phe Gly Leu Gly Leu Gly Leu Ile Glu Glu Lys 105 100 Gln Ala Glu Ser Arg Arg Ala Val Ser Ala Cys Gln Glu Ile Gln Ala 125 115 120 Ile Phe Thr Gln Lys Ser Lys Pro Gly Pro Asp Pro Leu Asp Thr Arg Arg Leu Gln Gly Phe Arg Leu Glu Glu Tyr Leu Ile Gly Gln Ser Ile 160 Gly Lys Gly Cys Ser Ala Ala Val Tyr Glu Ala Thr Met Pro Thr Leu 175 165 170 Pro Gln Asn Leu Glu Val Thr Lys Ser Thr Gly Leu Leu Pro Gly Arg 185 180 Gly Pro Gly Thr Ser Ala Pro Gly Glu Gly Gln Glu Arg Ala Pro Gly 205 200 195 Ala Pro Ala Phe Pro Leu Ala Ile Lys Met Met Trp Asn Ile Ser Ala 220 215

Fig. 3 (continued)

Gly	Ser	Ser	Ser 230	Glu	Ala	Ile	Leu	Asn 235	Thr	Met	Ser	Gln	Glu 240	Leu	Val
Pro	Ala	Ser 245	Arg	Val	Ala	Leu	Ala 250	Gly	Glu	Tyr	Gly	Ala 255	Val	Thr	Tyr
Arg	Lys 260	Ser	Lys	Arg	Gly	Pro 265	Lys	Gln	Leu	Ala	Pro 270	His	Pro	Asn	Ile
Ile 275	Arg	Val	Leu	Arg	Ala 280	Phe	Thr	Ser	Ser	Val 285	Pro	Leu	Leu	Pro	Gly 290
Ala	Leu	Val	Asp	Tyr 295	Pro	Asp	Val	Leu	Pro 300	Ser	Arg	Leu	His	Pro 305	Glu
Gly	Leu	Gly	His 310	Gly	Arg	Thr	Leu	Phe 315	Leu	Val	Met	Lys	Asn 320	Tyr	Pro
Cys	Thr	Leu 325	Arg	Gln	Tyr	Leu	Cys 330	Val	Asn	Thr	Pro	Ser 335	Pro	Arg	Leu
Ala	Ala 340	Met	Met	Leu	Leu	Gln 345	Leu	Leu	Glu	Gly	Val 350	Asp	His	Leu	Val
Gln 355	Gln	Gly	Ile	Ala	His 360	Arg	Asp	Leu	Lys	Ser 365	Asp	Asn	Ile	Leu	Val 370
Glu	Leu	Asp	Pro	Asp 375	Gly	Cys	Pro	Trp	Leu 380	Val	Ile	Ala	Asp	Phe 385	Gly
Cys	Cys	Leu	Ala 390	Asp	Glu	Ser	Ile	Gly 395	Leu	Gln	Leu	Pro	Phe 400	Ser	Ser
Trp	Tyr	Val 405		Arg	Gly	Gly	Asn 410	Gly	Cys	Leu	Met	Ala 415	Pro	Glu	Va1
Ser	Thr 420		Arg	Pro	Gly	Pro 425	Arg	Ala	Val	Ile	Asp 430	Tyr	Ser	Lys	Ala

Fig. 3 (continued)

Asp 435	Ala	Trp	Ala	Val	Gly 440	Ala	Ile	Ala	Tyr	Glu 445	Ile	Phe	Gly	Leu	Val 450
Asn	Pro	Phe	Tyr	Gly 455	Gln	Gly	Lys	Ala	His 460	Leu	Glu	Ser	Arg	Ser 465	Tyr
Gln	Glu	Ala	Gln 470	Leu	Pro	Ala	Leu	Pro 475	<u>Glu</u>	Ser	Val	Pro	Pro 480	Asp	Val
Arg	Gln	Leu 485	Val	Arg	Ala	Leu	Leu 490	Gln	Arg	Glu	Ala	Ser 495	Lys	Arg	Pro
Ser	Ala 500	Arg	Val	Ala	Ala	Asn 505	Val	Leu	His	Leu	Ser 510	Leu	Trp	Gly	Glu
His 515	Ile	Leu	Ala	Leu	Lys 520	Asn	Leu	Lys	Leu	Asp 525	Lys	Met	Val	Gly	Trp 530
Leu	Leu	Gln	Gln	Ser 535	Ala	Ala	Thr	Leu	Leu 540	Ala	Asn	Arg	Leu	Thr 545	Glu
Lys	Cys	Cys	Val 550	Glu	Thr	Lys	Met	Lys 555	Met	Leu	Phe	Leu	Ala 560	Asn	Leu
Glu	Cys	Glu 565	Thr	Leu	Суз	Gln	Ala 570	Ala	Leu	Leu	Leu	Cys 575	Ser	Trp	Arg
Ala	Ala	Leu													

Fig. 3 (continued)

GTCGACCCACGCGTCCGCCCACGCGTTCCGGAGACATGTCTCTGTGTTTC TCTCCCCTCCGCTTTTGAGTCCGTTGAAGACACAATTTCTCTCTGTCGGGT GCTTAGGAGGAGCTCCATGAACATGTATTGAATTGGACTTAGCTGAACAG GCTGCTGGTTGGCTGCCCAGAGGGGGCAGGCTGTGTTGCTGGGAGCCTTC CAGCTCCCTGCAGCAGTCATGGGGCAGGGTTCCCCGAGTCCGTAATCCCC TTAAGTGGAGCATCCCCTTTCCTGGGAGACACGAAGCAGGAAACACTGGC **AAATATCACAGCAGTGAGTTACGACTTTGATGAGGAATTCTTCAGCCAGA** CGAGCGAGCTGGCCAAGGACTTTATTCGGAAGCTTCTGGTTAAAGAGACC CGGAAACGGCTCACAATCCAAGAGGCTCTCAGACACCCCTGGATCACGCC GGTGGACAACCAGCAAGCCATGGTGCGCAGGGAGTCTGTGGTCAATCTGG AGAACTTCAGGAAGCAGTATGTCCGCAGGCGGTGGAAGCTTTCCTTCAGC **ATCGTGTCCCTGTGCAACCACCTCACCCGCTCGCTGATGAAGAAGGTGCA** CCTGAGGCCGGATGAGGACCTGAGGAACTGTGAGAGTGACACTGAGGAG GACATCGCCAGGAGGAAAGCCCTCCACCCACGGAGGAGGAGCAGCACCT CCTAACTGGCCTGACCTGCAGTGGCCGCCAGGGAGGTCTGGGCCCAGCGG GGCTCCCTTCTGTGCAGACTTTTGGACCCAGCTCAGCACCAGCACCCGGGC GTCCTGAGCACTTTGCAAGAGAGATGGGCCCAAGGAATTCAGAAGAGCTT GCAGGCAAGCCAGGAGACCCTGGGAGCTGTGGCTGTCTTCTGTGGAGGAG GCTCCAGCATTCCCAAAGCTCTTAATTCTCCATAAAATGGGCTTTCCTCTG TCTGCCATCCTCAGAGTCTGGGGTGGGAGTGTGGACTTAGGAAAACAATA TAAAGGACATCCTCATCATCACGGGGTGAAGGTCAGACTAAGGCAGCCTT CTTCACAGGCTGAGGGGGTTCAGAACCAGCCTGGCCAAAAATTACACCAG AGAGACAGAGTCCTCCCCATTGGGAACAGGGTGATTGAGGAAAGTGAACC TTGGGTGTGAGGGACCAATCCTGTGACCTCCCAGAACCATGGAAGCCAGG ACGTCAGGCTGACCAACACCTCAGACCTTCTGAAGCAGCCCATTGCTGGC CCGCCATGTTGTAATTTTGCTCATTTTTATTAAACTTCTGGTTTACCTGATG CTTGGCTTCTTTTAGGGCTACCCCCATCTCATTTCCTTTAGCCCGTGTGCCT GTAACTCTGAGGGGGGGCACCCAGTGGGGTGCTGAGTGGGCAGAATCTCA CTGGGAGTGGTGGCTCACGCCTATAATCCCTGCGCTTTGGCAGTCCGAGG CAGGGGAATTGCTTGAACTCAGGAGTTGGAGACCAGCCTGGGCAACATGG CAAAACGCAGTCTGTACAAAAAATACAAAAAATTAGCCAGGTGTAGGGGT AGGCACCTGGCATCCCAGCTACTCCAGGGGCTGAGGTGACAGCATTGCTT **AAGCCCAGAAGGTCGAGGCTGCAGTGAGCTGAGATCACGCCACTGCACTC** CGGCCGC

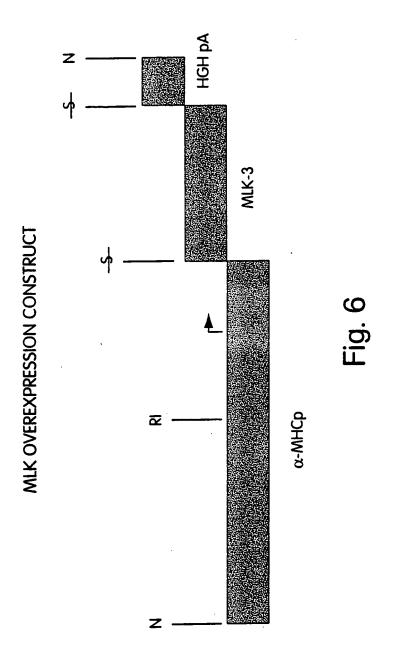
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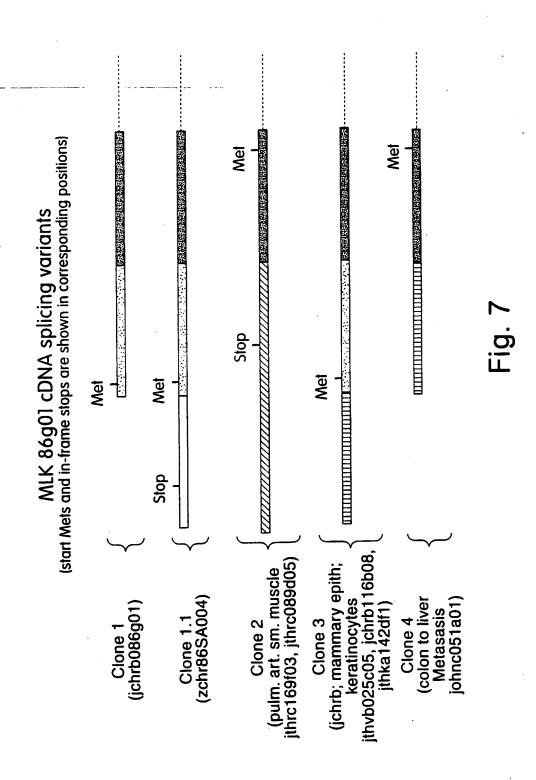
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K	K	V	T	D	Y	S	S	S	S	E	E	S	E	S	S	e	e	E	E	39
AAG	AAG	GTG	ACT	GAT	TAC	TCC	TCC	TCC	AGT	GAG	GAG	TCA	GAA	AGT	AGC	gag	gaa	GAG	GAG	117
e	D	G	E	S	E	T .	H	D	G	T	V	A	V	S	D	I	P	R	L	59
gaa	GAT	GGA	GAG	AGC	GAG	ACC	CAT	GAT	GGG	ACA	GTG	GCT	GTC	AGC	GAC	ATA		AGA	CTG	177
I	P	T	G	A	P	G	S	n	E	Q	Y	n	V	G	M	V	G	T		79
ATA	CCA	ACA	GGA	GCT	CCA	GGC	AGC	aac	GAG	CAG	TAC	aat	GTG	GGA	ATG	GTG	GGG	ACG		237
G GGG	L CTG	e gag	T ACC	S TCT	H CAT	A GCG	D GAC	s agt	F TTC	S AGC	G GGC	S AGT	I ATT	S TCA	R AGA	e gaa		T ACC		99 297
M	I	R	E	T	S	G	E	K	K	R	S	G	H	S	D	S	n	G	F	119
ATG	ATT	AGA	GAG	ACG	TCT	GGA	GAG	AAG	AAG	CGA	TCT	GGC	CAC	AGT	GAC	AGC	aat	GGC	TTT	357
A GCT	G GGC	H CAC	I ATC	N AAC	L CTC	P CCT	D GAC	L CTG	V GTG	Q CAG	Q CAG	S AGC	H CAT	S TCT	P CCA	A GCT		T ACC	PCCG	139 417
T	E	G	L	G	R	V	S	T	H	S	Q	e	M		S	G	T	e	Y	159
ACT	GAG	GGA	CTG	GGG	CGC	GTC	TCA	ACC	ÇAT	TCC	CAG	Gag	ATG		TCT	GGG	ACT	gaa	TAT	477
G	M	G	S	S	T	K	A	S	F	T	P	F	V	D	P	r	V	Y	Q	179
GGC	ATG	GGG	AGC	AGC	ACC	AAA	GCC	TCC	TTC	ACC	CCC	TTT	GTG	GAC		aga	GTA	TAC	CAG	537
T	S	P	T	D	e	D	E	E	D	E	e	s	S	A	A	a	L	F	T	199
ACG	TCT		ACT	GAT	gaa	GAT	GAÀ	GAG	GAT	GAG	gaa	TCA	TCA	GCC	GCA	GCT	CTG	TTT	ACT	597
S	E	L	L	R	Q	E	Q	A	K	L	n	e	A	R	K	I	S	V	V	219
AGC	GAA	CTT	CTT	AGG	CAA	GAA	CAG	GCC	AAA	CTC	aat	gaa	GCA	AGA	AAG		TCG	GTG	GTA	657
n	V	N	P	T	N	I	R	P	H	S	D	T	P	e	I	R	K	Y	K	239
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K	R	F	N	S	E	I	L	C	A	A	L	W	g	V	N	L	L	V	G.	259
AAA	CGA	TTC	AAC	TCA	GAA	ATA	CTT	TGT	GCA	GCT	CTG	TGG	ggt	GTA	AAC	CTT	CTG	GTG	GGG	777
T	E	n	G	L	M	L	L	D	R	s	G	Q	G	K	V	Y	n	L	I	279
ACT	GAA	aat	GGC	CTG	ATG	CTT	TTG	GAC	CGA	agt	GGG	CAA	GGC	AAA	GTC	TAT	aat	CTG	ATC	837
N	R	R	R	P	Q	Q	M	D	V	L	e	G	L	n	V	L	V	T	I	299
AAC	CGG	AGG	GGA	TTT	CAG	CAG	ATG	GAT	GTG	CTA	Gag	GGA	CTG	aat	GTC	CTT	GTG	ACA	ATT	897
s	G	K	K	n	K	L	R	V	Y	Y	L	S	W	L	R	n	R	I	L	319
TCA	GGA	AAG	AAG	aat	AAG	CTA	CGA	GTT	TAC	TAT	CTT	TCA	TGG	TTA	AGA	aac	AGA	ATA	CTA	957
H	N	D	P	E	V CTA	E	K	K	Q CAA	G GGC	W TGG	I ATC	T ACT	V GTT	G GGG	D GAC	L TTG	E Gaa	G GGC	339 1017

Fig. 5

C	I	H	Y	K	V	V	K	Y	e	r	I	K	F	L	V	I	A	L	K	359
TGT	ATA	CAT	TAT	AAA	GTT	GTT	AAA	TAT	Gaa	agg	ATC	Aaa	TTT	TTG	GTG	ATT	GCC	TTA	AAG	1077
n	A	V	E	I	Y	A	W	A	P	K	P	Y	H	K	F	M	A	F	K	379
aat	GCT	GTG	GAA	ATA	TAT	GCT	TGG	GCT	CCT	AAA	CCG	TAT	CAT	AAA	TTC	atg	GCA	TTT	AAG	1137
S TCT	F TTT	A GCA	D GAT	L	Q CAG	H_CAC	K AAG	P	L CTG	L_CTA	V GTT	D GAT	L CTC	T ACG	V GTA	E GAA	E GAA	g Ggt	Q CAA	399 1197
R	L	K	V	I	F	g	S	H	T	g	F	H	V	I	D	V	D	S	G	419
AGA	TTA	AAG	GTT	ATT	TTT	ggt	TCA	CAC	ACT	GGT	TTC	CAT	GTA	ATT	GAT	GTT	GAT	TCA	GGA	1257
N	S	Y	D	I	Y	I	P	s	H	I	Q	G	n	I	T	P	H	A	I	439
AAC	TCT	TAT	GAT	ATC	TAC	ATA		TCT	CAT	ATT	CAG	GGC	aat	ATC	ACT	CCT	CAT	GCT	ATT	1317
		L																		444 1332

Fig. 5 (continued)





SUBSTITUTE SHEET (RULE 26)

Full-length nos no? yes yes Resulting proteins from MLK86g01 splice variants Fig. 8 HMM pkinase domain match active site Clone 3
(jchrb; mammary epith; keratinocytes jthvb025c05, jchrb116b08, jthka142df1) Clone 2 (pulm. art. sm. muscle jthrc169f03, jthrc089d05) Clone 4 (colon to liver Metasasis johnc051a01) Clone 1 (jchrb086g01)

SUBSTITUTE SHEET (RULE 26)

Clone 1.1 (includes parts of zchr86SA004, jchrb086g01) TGGGGATATCTGAGCTATTTCTCTGTGGGCAGCTGTGTTTCAAAGTCTGGG CAGGTTGTTGTTGAATTTTGCGTGGGCTGCCAGGATTTTGTGGAAGTATAA TACTTTGTCATTATGAGATGTCGTCTCTCGGTGCCTCCTTTGTGCAAATTAA **ATTTGATGACTTGCAGTTTTTTGAAAACTGCGGTGGAGGAAGTTTTGGGAG** TGTTTATCGAGCCAAATGGATATCACAGGACAAGGAGGTGGCTGTAAAGA AGCTCCTCAAAATAGAGAAAGAGGCAGAAATACTCAGTGTCCTCAGTCAC **AGAAACATCATCCAGTTTTATGGAGTAATTCTTGAACCTCCCAACTATGGC ATTGTCACAGAATATGCTTCTCTGGGATCACTCTATGATTACATTAACAGT** AACAGAAGTGAGGAGATGGATATGGATCACATTATGACCTGGGCCACTGA TGTAGCCAAAGGAATGCATTATTTACATATGGAGGCTCCTGTCAAGGTGA TTCACAGAGACCTCAAGTCAAGAAACGTTGTTATAGCTGCTGATGGAGTA CTGAAGATCTGTGACTTTGGTGCCTCTCGGTTCCATAACCATACAACACAC ATGTCCTTGGTTGGAACTTTCCCATGGATGGCTCCAGAAGTTATCCAGAGT CTCCCTGTGTCAGAAACTTGTGACACATATTCCTATGGTGTGGTTCTCTGG GAGATGCTAACAAGGGAGGTCCCCTTTAAAGGTTTGGAAGGATTACAAGT **AGCTTGGCTTGTAGTGGAAAAAACGAGAGATTAACCATTCCAAGCAGTT** GCCCCAGAAGTTTTGCTGAACTGTTACATCAGTGTTGGGAAGCTGATGCC **AAGAAA**CGGCCATCATTCAAGCAAATCATTTCAATCCTGGAGTCCATGTC **AAATGACACGAGCCTTCCTGACAAGTGTAACTCATTCCTACACAACAAGG** CGGAGTGGAGGTGCGAAATTGAGGCAACTCTTGAGAGGCTAAAGAAACT AGAGCGTGATCTCAGCTTTAAGGAGCAGGAGCTTAAAGAACGAGAAAGA CGTTTAAAGATGTGGGAGCAAAAGCTGACAGAGCAGTCCAACACCCCGCT TCTCTTGCCTCTTGCTGCAAGAATGTCTGAGGAGTCTTACTTTGAATCTAA **AACAGAGGAGTCAAACAGTGCAGAGATGTCATGTCAGATCACAGCAACA AGTAACGGGGAGGGCCATGGCATGAACCCAAGTCTGCAGGCCATGATGCT** GATGGGCTTTGGGGATATCTTCTCAATGAACAAAGCAGGAGCTGTGATGC **ATTCTGGGATGCAGATAAACATGCAAGCCAAGCAGAATTCTTCCAAAACC** ACATCTAAGAGAAGGGGGAAGAAAGTCAACATGGCTCTGGGGTTCAGTG **ATTTTGACTTGTCAGAAGGTGACGATGATGATGATGACGGTGAGGAG** GAGGATAATGACATGGATAATAGTGAATGAAAGCAGAAAGCAAAGTAAT AAAATCACAAATGTTTGGAAAACACAAAAGTAACTTGTTTATCTCAGTCT GTACAAAACAGTAAGGAGGCAGAAAGCCAAGCACTGCATTTTTAGGCC **AATCACATTTACATGACCGTAATTTCTTATCAATTCTACTTTTATTTTTGCT** TACAGAAAAACGGGGGGAGAATTAAGCCAAAGAAGTATATTTATGAATC AGCAAATGTGGTGCCTGATTATAGAAATTTGTGATCCTATATACAATATAG GACTTTTAAAGTTGTGACATTCTGGCTTTTTCTTTTAATGAATACTTTTTAG TTTGTATTTGACTTTATTTCCTTTATTCAAATCATTTTTAAAAACTTACATT TTGAACAAACACTCTTAACTCCTAATTGTTCTTTGACACGTAGTAATTCTG TGACATACTTTTTTTTTTTTATAGCAATACACTGTAATATCAGAAATGGTT GGCCTGAGCAACCTAGTAAGACCTCGTCTCTACTAATAATTAAAAAAACTA GCTGGCATGGTAGCACACCTGTAGTCCCAGATACTTGGGAGGCCAAGG CAGGAGGATTGCTTGAGACCTAGCAATCAGTCAGGGCTGCAGTGAGCCAT GATGGCACCACTGCACTCTAGCCTGGGCAAGAGAACAAGATCCTGTCTCA AAAACAAAAAAAAAAAAAAAAGGGCGGCCGC

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		-	tca Ser 100		-	-		-				_	-	-	336
	_	_	gct Ala	 _			-	-		-			-	-	. 384
			cta Leu												432
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			cca Pro												528
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	His		tta Leu				Tyr							ata Ile	624
His	_		gga Gly			-		_						_	672

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Gln	Cys	Trp	Glu	Ala 245	Asp	Ala	Lys	Lys	Arg 250	Pro	Ser	Phe	Ly	ີ ກ 255	Ile
Ile	Ser	Ile	Leu 260	Glu	Ser	Met	Ser	Asn 265	Asp	Thr	Ser	Leu	Pro 270	Asp	Lys

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- 12 -

•																•
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							cta Leu									392
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Pro	Gly 210	Ala	Pro	Ala	Phe	Pro 215	Leu	Ala	Ile	Lys	Met 220	Met	Trp	Asn	Ile
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Thr	Tyr	Arg	Lys 260	Ser	Lys	Arg	Gly	Pro 265	Lys	Gln	Leu	Ala	Pro 270	His	Pro

ASI	TTE	275	-	val	Leu	Arg	280		2 1111	. sei	. Sei	285		o re	и те
Pro	Gly 290		Leu	Val	Asp	Tyr 295		Asp	Val	l Lei	300		Arg	g Le	u Hi
Pro 305		Gly	Leu	Gly	His 310	_	Arg	Thr	Lev	1 Phe 315		ı Val	. Met	Lys	32
Tyr	Pro	Cys	Thr	Leu 325	Arg	Gln	Tyr	Leu	330		. Asn	Thr	Pro	335	
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Phe 385	Gly	Cys	Cys	Leu	Ala 390	Asp	Glu	Ser	Ile	Gly 395		Gln	Leu	Pro	Phe 400
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Asp	Val	Arg	Gln	Leu 485	Val	Arg	Ala	Leu	Leu 490	Gln	Arg	Glu	Ala	Ser 495	Lys
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Gly	Glu	His 515	Ile	Leu	Ala	Leu	Lys 520	Asn	Leu	Lys	Leu	Asp 525	Lys	Met	Val
Gly	Trp 530	Leu	Leu	Gln	Gln	Ser 535	Ala	Ala	Thr	Leu	Leu 540	Ala	Asn	Arg	Leu
Thr	Glu	Lys	Cys	Cys	Val	Glu	Thr	Lys		Lys	Met	Leu	Phe	Leu	Ala

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- 19 -

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								Pro					Glu		gct Ala	624
												Met			atc Ile	672
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		cca Pro								Gly						768
		aga Arg												His		816
		atc Ile 275														864
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		ggc Gly														960
		tgt Cys														1008
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Lys Asp Phe Ile Arg Lys Leu Leu Val Lys Glu Thr Arg Lys Arg Leu 50 55 60

Thr Ile Gln Glu Ala Leu Arg His Pro Trp Ile Thr Pro Val Asp Asn 65 70 75 80

Gin Cin Ala Met Val Arg Arg Glu Ser Val Val Asn Leu Glu Asn Phe 35 90 95

Arg Lys Gln Tyr Val Arg Arg Trp Lys Leu Ser Phe Ser Ile Val 100 105 110

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			Phe												gcc Ala	144
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	Leu					acc Thr										384
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Leu Val Gly Thr Glu Asn Gly Leu Met Leu Leu Asp Arg Ser Gly Gln 265

Gly Lys Val Tyr Asn Leu Ile Asn Arg Arg Arg Phe Gln Gln Met Asp 275

Val Leu Glu Gly Leu Asn Val Leu Val Thr Ile Ser Gly Lys Lys Asn 295

Lys Leu Arg Val Tyr Tyr Leu Ser Trp Leu Arg Asn Arg Ile Leu His 305

Asn Asp Pro Glu Val Glu Lys Lys Gln Gly Trp Ile Thr Val Gly Asp

Leu Glu Gly Cys Ile His Tyr Lys Val Val Lys Tyr Glu Arg Ile Lys

Phe Leu Val Ile Ala Leu Lys Asn Ala Val Glu Ile Tyr Ala Trp Ala

Pro Lys Pro Tyr His Lys Phe Met Ala Phe Lys Ser Phe Ala Asp Leu

Gln His Lys Pro Leu Leu Val Asp Leu Thr Val Glu Glu Gly Gln Arg

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<211> 26
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<213> Artificial Sequence

- 34 -

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                                                                    26
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<211> 37
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      is any amino acid
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             20
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Xaa Arg Xaa His Xaa
         35
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<213> synthetic construct
<223> at positions 2,4,5,7-x is any amino acid
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Gly Xaa Gly Xaa Xaa Gly Xaa Lys
  1
<210> 24
<211> 49
<212> PRT
<213> synthetic construct
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<220>
<223> at positions
1,3,5,7,9,11,13,15,19,21,23,25,29,31,35,37,39,41,4
4,47,49-x is any amino acid

Arg Asp Xaa Lys Xaa Asn Xaa Lys Xaa Asp Phe Gly Xaa Trp Xaa Ala 20 25 30

Pro Glu Xaa Trp Xaa Glu Xaa Pro Xaa Cys Trp Xaa Arg Pro Xaa Phe 35 40 45

Xaa

INTERNATIONAL SEARCH REPORT

International application No. PCT/US99/20631

			101/00///200	J.
IPC(6) US CL	SSIFICATION OF SUBJECT MATTER:Please See Extra Sheet.:435/194, 6, 15, 7.1, 320.1, 325, 252.3; 530/387.1; to International Patent Classification (IPC) or to both	536/23.2 h national classification	and IPC	
B. FIEI	LDS SEARCHED			
Minimum d	ocumentation searched (classification system follow	ed by classification sym	bols)	
U.S. :	435/194, 6, 15, 7.1, 320.1, 325, 252.3; 530/387.1; 5	36/23.2		
Documenta	tion searched other than minimum documentation to t	he extent that such docum	nents are included	in the fields searched
	lata base consulted during the international search (if T, MEDLINE, EMBASE, BIOSIS, SCISEARCH, B			
C. DOC	UMENTS CONSIDERED TO BE RELEVANT		······································	
Category*	Citation of document, with indication, where a	ppropriate, of the relevan	nt passages	Relevant to claim No.
X	WO 9811234 A2 (INCYTE PHARMA 1998, see abstract and their SEQ ID acid sequence with 67% homology to comprising a fragment encoding at least of SEQ ID NO:2 of this invention.	NO:1 for encoding claimed SEQ ID	g an amino NO:2 and	5-26
Furth	er documents are listed in the continuation of Box (See patent	family annex.	
Spe	ocial categories of cited documents:	*T* later document p	ublished after the inter	national filing date or priority
	nument defining the general state of the art which is not considered be of particular relevance	date and not in	conflict with the applications the	eation but cited to understand
	oe or perusuant resevence lier document published on or after the international filing date	"X" document of per	ticular relevance; the	claimed invertion cannot be
°L° doc cite	nument which may throw doubts on priority claim(s) or which is d to establish the publication date of another citation or other cial reason (se specified)	when the docum	ent is taken alone	ed to involve an inventive step
•	nument referring to an oral disclosure, use, exhibition or other	considered to ir	volve en inventive	claimed invention cannot be step when the document is documents, such combination e art
	ument published prior to the international filing dete but later than priority date claimed	-	or of the same patent (1
	actual completion of the international search MBER 1999	Date of mailing of the	international sear 3 FEB 200	· I
Commission Box PCT Washington	sailing address of the ISA/US ser of Patents and Trademarks , D.C. 20231		WENCE HIPOURI	
Facsimile No	o. (703) 305-3230	Telephone No. (703	37 378 77104	į

INTERNATIONAL SEARCH REPORT

International application No. PCT/US99/20631

<u> </u>
Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Claims Nos.: 1-2, 4-26 partially because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
These claims refer to SEQ ID NO:s but the case does not comply with sequence rules. Only SEQ ID NOs: 1-3 which were elected and searched in the parent U.S. application could be searched in the instant case.
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search sees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest
No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No. PCT/US99/20631

12N 15/00, 5	/00, 1/20,	9/12; C07K 1	.6/00; C07H 2	21/04; G01N 33	/53		
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